

This document is the Accepted Manuscript version of a Published Work that appeared in final form in **JOURNAL OF PHYSICAL CHEMISTRY B**, copyright 2011 American Chemical Society after peer review and technical editing by the publisher. To access the final edited and published work see <https://doi.org/10.1021/jp200748u>.

Postprint of: Śmiechowski M. , Gojło E. , Stangret J., Hydration of Simple Carboxylic Acids from Infrared Spectra of HDO and Theoretical Calculations, JOURNAL OF PHYSICAL CHEMISTRY B, Vol. 115, Iss. 16 (2011), pp. 4834–4842

Hydration of Simple Carboxylic Acids from Infrared Spectra of HDO and Theoretical Calculations

Journal:	<i>The Journal of Physical Chemistry</i>
Manuscript ID:	jp-2011-00748u.R1
Manuscript Type:	Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Śmiechowski, Maciej; Gdansk University of Technology, Department of Physical Chemistry Gojło, Emilia; Gdansk University of Technology, Department of Physical Chemistry Stangret, Janusz; Gdansk University of Technology, Department of Physical Chemistry

SCHOLARONE™
Manuscripts

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Hydration of Simple Carboxylic Acids from Infrared Spectra of HDO and Theoretical Calculations

Maciej Śmiechowski, Emilia Gojło, Janusz Stangret*

Department of Physical Chemistry, Chemical Faculty, Gdańsk University of Technology,

Narutowicza 11/12, 80-233 Gdańsk, Poland

E-mail address: msmiech@chem.pg.gda.pl

RECEIVED DATE:

* To whom correspondence should be addressed. Phone: (+48) 58 347 1283. Fax: (+48) 58 347 2694.

E-mail address: msmiech@chem.pg.gda.pl.

1
2 The hydration of carboxylic acids in dilute aqueous solutions is important for our understanding of their
3
4 functioning in the biochemical context. Here we apply vibrational spectra of HDO isotopically diluted in
5
6 H₂O to study this phenomenon, using the difference spectra method for analysis and interpretation of the
7
8 results. The spectra of HDO affected by formic, acetic and propionic acid display characteristic
9
10 component bands, significantly red-shifted from the bulk HDO band position. The appearance of these
11
12 component bands is linked with isotopic substitution on the carboxylic acid molecule, which forms a
13
14 short and strong hydrogen bond with a water molecule. Additionally, a charge separation due to the
15
16 proton transfer in the neutral form of the complex leading to a contact ion pair formation may be
17
18 inferred from the affected HDO spectra. Apart from the contraction of the principal acid-water hydrogen
19
20 bond, it results in other major structural changes in the hydration shell, as revealed by Density
21
22 Functional Theory (DFT) calculations of optimal geometries of aqueous clusters of the studied acids.
23
24
25
26
27
28

29 Keywords: Carboxylic acids, aqueous solutions, hydration, FTIR spectroscopy, DFT calculations.
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Introduction

Carboxylic acids form an important class of biomolecules and their hydration in dilute aqueous solutions has important implications for our understanding of their role in the biochemical context. Owing to the presence of both an alkyl chain and a hydrophilic $-\text{COOH}$ head group, carboxylic acids often experience a mixed type of hydration, i.e., the hydrophobic hydration of the alkyl chain combined with the hydrophilic hydration of the head group. The cooperative effects of both types of hydration are supposed to determine the overall interaction of the molecule with water.¹

Carboxylic acids belong to moderately weak ones ($\text{p}K_{\text{a}} = 3.75, 4.76, 4.87$ for formic, acetic, and propionic acid, respectively)² and are mostly undissociated in an aqueous solution. Strong acids, upon dissolution in water, release the proton into the surrounding medium, giving rise to the hydrated proton species with unique properties. The debate, whether it is the so-called Eigen cation (H_3O^+) or the Zundel cation (H_5O_2^+) that correctly describes the hydrated individual, now seems to reach a conclusion that H^+_{aq} forms a “fluxional defect” in the hydrogen-bonded (H-bonded) network of water³ with proton transfer (PT) acts facilitated by complicated rearrangements of the H-bond network.⁴ The existence of some intermediate forms, variously referred to as a “deformed Eigen” or “deformed Zundel” cation has been also proposed on the basis of computational and spectroscopic studies.⁵⁻⁹

Infrared spectra of aqueous solutions of strong acids are characterized by the so-called “continuum of absorption” (very broad, essentially structureless low-intensity absorption in the $3000\text{-}1000\text{ cm}^{-1}$ region). The concept of “extremely polarisable” hydrogen bonds was proposed by Zundel and his co-workers to justify its appearance.¹⁰ However, in relatively dilute aqueous solutions of strong acids this region of the vibrational spectrum could be adequately resolved into analytical component bands.^{7,8} For weaker acids (such as HCOOH) the intensity of the “continuum” is much weaker and the protons in the acid-water H-bonds are supposed to reside preferentially at the acid site, with the protonic polarisability being much lower.¹¹

Although this picture of strong acid-water H-bonding with acid as proton donor and water as proton acceptor in the H-bond prevailed in the interpretation of infrared data,^{11,12} the possibility of carboxylic

1 acids to form relatively stable ion pairs in water was also investigated.¹³⁻²⁰ Molecular dynamics (MD)
2 simulations led to a conclusion that the formation of a contact ion pair (CIP) is the first step leading
3 ultimately to the act of acid dissociation^{16-18,20} (by contact ion pair we denote a structure, in which the
4 acidic anion and the hydrated proton are directly adjacent after the occurrence of dissociation).
5
6
7
8

9
10 Simple carboxylic acids in aqueous environment have been thus far studied with diverse theoretical
11 and experimental methods. Their aqueous clusters were investigated with static *ab initio*
12 calculations,^{14,15,19,21-26} as well as with matrix isolation vibrational spectroscopy²⁶ and mass
13 spectrometry.²⁷ Aqueous solutions of carboxylic acids were studied computationally with MD^{16-20,28,29}
14 and Monte Carlo (MC) simulations.^{30,31} Experimental studies of such solutions were mainly conducted
15 with vibrational spectroscopy, both infrared^{11,12} and Raman spectroscopy,^{13,32-38} but also dielectric
16 spectroscopy,³⁹ X-ray scattering studies⁴⁰ and volumetric measurements⁴¹ were applied for this purpose.
17
18
19
20
21
22
23
24
25

26 Vibrational spectroscopy, although regarded as one of the best methods to study the solute hydration,⁴²
27 in the case of aqueous solutions of protic solutes faces the added complication of the presence of
28 different –OH groups in the system, originating not exclusively from the water molecules. The
29 spectroscopy of HDO isotopically diluted in H₂O (D₂O), which is widely considered an ideally suited
30 method in hydration studies,⁴³ in this case offers no benefit by itself, since isotopic substitution in
31 principle proceeds on all protic groups in the system. One of the possible solutions is the application of
32 factor analysis techniques to the vibrational spectra of aqueous solutions, which allow extraction of
33 spectra of chemically relevant species present in the system. This approach was already applied in the
34 studies of aqueous carboxylic acids in H₂O.¹² Another approach, recently used in our group for the
35 interpretation of HDO spectra, is to couple the band shape analysis with the static *ab initio* quantum
36 mechanical (QM) calculations of optimal structures of small aqueous clusters. The ascription of
37 component bands in the HDO spectra to particular hydrogen bonds in aqueous clusters is then possible
38 resorting to empirical H-bond length vs. stretching frequency correlations.⁴⁴ Such approach proved
39 successful in discriminating different deuterium substitutions in systems like aqueous solutions of the
40 hydroxide anion⁴⁵ or anions of the phosphate(V) family.⁴⁶ Additionally, the aforementioned
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 “continuum” is absent in the HDO spectra in H₂O in the OD stretching range, due to isotopic decoupling
2 of the OD oscillators from the surrounding bath.⁹ Therefore, the application of HDO spectroscopy to
3 study the hydration of carboxylic acids seems a preferable method.
4
5

6
7 For the present study, the three simplest homologous carboxylic acids, i.e., formic, acetic, and
8 propionic acid have been selected. To study their hydration, we used vibrational (FTIR) spectroscopy of
9 HDO isotopically diluted in aqueous solutions of the studied solutes in H₂O. The quantitative method of
10 spectral data analysis formulated by Lindgren and coworkers, and independently in our laboratory,⁴⁷⁻⁴⁹
11 allows separation of solute-affected water spectrum basing on the series of spectra measured at
12 increasing concentration of the solute. The transformation of the obtained HDO band contour to water
13 oxygen-oxygen distance probability distribution function, basing on the published procedure,^{47,49} reveals
14 also the structural state of the hydration spheres. To further link the structural and spectral data, we also
15 applied static *ab initio* QM calculations of optimal structures of small aqueous clusters of the solutes in
16 the gas phase and in the water solvent simulated by the polarizable continuum model (PCM).⁵⁰
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32

33 **Experimental Section**

34
35 Formic acid (80% aq., pure p.a.), acetic acid (pure p.a.) and propionic acid (97%) from POCh Poland
36 were used as supplied. D₂O (99.84% isotopic purity) was provided by Institute of Nuclear Investigation,
37 Poland. Stock solutions were prepared by dissolving weighted amounts of respective solutes in double-
38 distilled water. The series of solutions were prepared by dissolving weighted amounts of respective
39 stock solution in double-distilled water. The solution series of carboxylic acids included samples from
40 the 0.1-1.0 mol·kg⁻¹ molality range. The natural pH of the solutions was not adjusted and its range was
41 1.88-2.37 for formic acid, 2.40-2.89 for acetic acid and 2.46-2.94 for propionic acid. Sample solutions
42 were made by adding 4% (by weight) of D₂O relative to H₂O and reference solutions by adding the same
43 molar amounts of H₂O. Densities of the solutions were measured with an Anton Paar DMA 5000
44 density meter at 25.000±0.001 °C.
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 FTIR spectra were recorded on IFS 66 Bruker spectrometer. 256 scans were made with 4 cm^{-1}
2 resolution. A cell with CaF_2 windows was employed. The path length was 0.0299 mm, as determined
3 interferometrically. The spectrometer was purged with dry air free of carbon dioxide. The temperature,
4 monitored by a thermocouple inside the cell, was kept at $25.0\pm 0.1\text{ }^\circ\text{C}$ by circulating thermostated water
5 through mounting plates of the cell.
6
7
8
9
10

11 **Spectral Data Analysis**

12
13
14
15
16
17 The spectra have been handled and analyzed by commercial programs GRAMS/32 4.01 (Galactic
18 Industries Corporation, Salem, USA) and RAZOR (Spectrum Square Associates, Inc., Ithaca, USA) run
19 under GRAMS/32.
20
21
22

23
24 Spectral data have been analyzed following the published procedures, allowing the separation of the
25 spectrum of solute-affected water, basing on the spectra of the entire solution series and the bulk HDO
26 spectrum.^{48,49} The algorithm is based on the assumption that water in solution may be divided into
27 additive contributions of bulk (b) and solute-affected (a) water. The vibrational spectrum of the latter,
28 ε_a , may be calculated for each wavenumber using Eq. (1a) from the pure HDO spectrum, ε_b , and the
29 solution spectrum, ε , assuming that the “affected number”, N , is known (its significance will be
30 discussed below). In the infinite dilution limit ($m \rightarrow 0$) the bracketed fraction may be approximated by
31 the derivative of molar absorptivity vs. molality as in Eq. (1b). In both equations M denotes the mean
32 molar mass of water ($\text{H}_2\text{O} + 4\% \text{D}_2\text{O}$).
33
34
35
36
37
38
39
40
41
42
43
44

$$45 \varepsilon_a = \frac{1}{NM} \left(\frac{\varepsilon - \varepsilon_b}{m} \right) + \varepsilon_b \quad (1a)$$

$$46 \varepsilon_a = \frac{1}{NM} \left(\frac{\partial \varepsilon}{\partial m} \right)_{m=0} + \varepsilon_b \quad (1b)$$

47
48
49
50
51
52
53
54
55 An approximation of the experimental spectra $\varepsilon(\nu_i)$ vs. molality m at each wavenumber ν_i by the least
56 squares method allows the calculation of the respective derivative.
57
58
59
60

1 The N parameter is equal to the number of moles of water affected by one mole of solute in the
2 infinite dilution conditions. The proper value of it is found basing on the published algorithm.⁴⁹ Briefly,
3 the trial solute-affected water spectrum for a given N value is fitted using the baseline, analytical bands
4 and the bulk water spectrum. The product of Gaussian and Lorentzian peak functions is normally used
5 as the starting analytical band shape, but it might be replaced later by a pure function for a given band if
6 the contribution of the other component is found negligible. The maximum value of N , for which the
7 solute-affected water spectrum still contains a negligible amount of the bulk water spectrum is
8 considered as the “true” value of N (usually, the threshold value for bulk water contribution is set at
9 0.5% of the total integrated intensity of the ε_a spectrum). The ε_a spectrum corresponding to this N is
10 regarded as the “true” affected water spectrum. Thus, both unknowns are obtained simultaneously.
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25

26 Computational Details

27
28 The model systems in the calculations were aqueous clusters $\text{RCOOH}(\text{H}_2\text{O})_n$ ($\text{R} = \text{H}, \text{CH}_3, \text{C}_2\text{H}_5, n =$
29 1–8). Sixteen clusters were considered for each acid, and thus the total number of cluster structures
30 discussed here is 48.
31
32
33
34

35 QM calculations were performed in the framework of Density Functional Theory (DFT), utilizing the
36 hybrid B3LYP exchange-correlation functional with the 6-311++G(d,p) basis set.⁵¹ All calculations
37 were performed with the Gaussian 09 system.⁵² *Tight* convergence criteria were used for optimizations.
38 Vibrational analysis was performed on optimized clusters to check for absence of imaginary frequencies
39 and thus confirm the existence of local energetic minima. Zero-point vibrational energies (ZPVE) and
40 thermal corrections to energy, enthalpy and Gibbs free energy were simultaneously obtained during the
41 vibrational analysis. The ZPVE values were empirically scaled by the recommended scale factor equal
42 to 0.9877.⁵³
43
44
45
46
47
48
49
50
51
52
53

54 Gas phase cluster structures were subsequently used as starting points for geometry optimization in the
55 self-consistent reaction field approach. The polarizable continuum solvation model (PCM) in the
56 integral equation formalism was used.⁵⁰ The atomic radii for molecular cavity definition were taken
57
58
59
60

1 from the UFF force field.⁵⁴ Additionally, the new SMD parameterization for PCM was used.⁵⁵ The
2 united atom approach was not applied in the PCM calculations.
3
4
5
6
7
8

9 **Results and Discussion**

10 Figure 1a presents the molar absorptivity derivatives $(\partial\varepsilon/\partial m)_{m=0}$ obtained on the basis of the measured
11 spectra for the solution series of the three studied acids (available as Supporting Information). These
12 derivatives correspond to the linear regression of $\varepsilon(\nu)$ values vs. solution molality. Together with the
13 bulk HDO spectrum, they were used to determine the solute-affected HDO spectra according to eq 1.
14 Spectra of HDO affected by formic, acetic and propionic acids are shown in Figure 1b. The affected
15 number was found to be almost constant for the three studied acids ($N = 3.9 \pm 0.1$). A similar value for
16 the total number of acid-water interactions was obtained previously for dilute acetic acid-water mixtures
17 in X-Ray diffraction experiments.⁴⁰ The decomposition of the affected spectra into analytical component
18 bands is illustrated in Figure 2 and their most important parameters (excluding baseline correction
19 bands) are gathered in Table 1.
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34

35 As seen in Table 1, component band positions at maximum (ν°_{OD}) are similar for all the acids.
36 Excluding the band at $\sim 2125\text{ cm}^{-1}$ for acetic acid in Figure 2b (which arises from the uncompensated
37 intramolecular absorption of the acid), other bands seem to correspond closely. In the case of acetic and
38 propionic acids of very similar strength, the band positions are almost identical. For the stronger formic
39 acid, a slight blue shift of the $\sim 2550\text{ cm}^{-1}$ and the $\sim 2430\text{ cm}^{-1}$ bands is apparent, accompanied by a red
40 shift of the $\sim 2260\text{ cm}^{-1}$ and the $\sim 2050\text{ cm}^{-1}$ bands. Additionally, a well-developed band at ca. 1950 cm^{-1}
41 is apparent for formic acid and is slightly less intense for acetic acid, while for propionic acid it is not
42 observed.
43
44
45
46
47
48
49
50
51
52

53 As we have recently shown, analogous carboxylates display two component bands of affected HDO at
54 ~ 2550 and $\sim 2420\text{ cm}^{-1}$.⁵⁵ These locations correspond well to those observed in this work. We previously
55 interpreted them in terms of instantaneous charge localization on the carboxylate anion, with one oxygen
56
57
58
59
60

1 atom negatively charged as in the hydroxide anion (for which $\nu^{\circ}_{\text{OD}} = 2437 \text{ cm}^{-1}$)⁴⁵ and the other one
2 resembling the carboxylic group in ketones (for which $\nu^{\circ}_{\text{OD}} = 2540 \text{ cm}^{-1}$).⁵⁶ In an undissociated
3 carboxylic acid molecule, the localization of the single and double bond is obvious and the band at
4
5
6
7 $\sim 2550 \text{ cm}^{-1}$ may be ascribed to the water molecules H-bonded to the oxygen atom engaged in the C=O
8
9 bond. This, however, does not justify the presence of the band at $\sim 2430 \text{ cm}^{-1}$. The H-bond donated by
10
11 the acid to the accepting water molecule may be suspected to be much stronger. For phosphoric(V) acid,
12
13 an analogous H-bond gives rise to an HDO band at 2310 cm^{-1} ,⁴⁶ while for carboxylic acids in H_2O , a
14
15 strong and short acid-water H-bond is responsible for the ν_{OH} band at $\sim 3000 \text{ cm}^{-1}$.¹² The latter figure,
16
17 transformed to OD intramolecular stretching frequency⁵⁷ gives $\nu^{\circ}_{\text{OD}} = \sim 2280 \text{ cm}^{-1}$. Therefore, the ~ 2260
18
19 cm^{-1} band may be with high certainty ascribed to the C-O-D \cdots OH₂ hydrogen bond. The appearance of
20
21 the $\sim 2430 \text{ cm}^{-1}$ band in the carboxylic acid-affected HDO spectra will be discussed later.
22
23
24

25
26 The more elusive is the origin of the $\sim 2625 \text{ cm}^{-1}$ and $\sim 2050 \text{ cm}^{-1}$ bands. The former may be traced in
27
28 the H_3PO_4 -affected HDO spectrum, interpreted previously by us.⁴⁶ It has been ascribed to the water-acid
29
30 H-bond to the acidic oxygen atom (i.e., the one that has the acidic proton directly bonded to it). The
31
32 engagement in a strong H-bond via the acidic hydrogen makes this oxygen atom a poor electron pair
33
34 donor and weakens the accepted H-bond in an anticooperative way. A similar H-bonding pattern for
35
36 carboxylic acids is expected.
37
38

39
40 The remaining $\sim 2050 \text{ cm}^{-1}$ band (and also the $\sim 1950 \text{ cm}^{-1}$ one) is unusually red-shifted for a
41
42 component band of the HDO spectrum. Previously, such situation was encountered by us only in the
43
44 case of the H^+ -affected HDO spectrum in strongly acidic solutions.⁹ On the basis of the ab initio
45
46 optimized cluster geometries, the possibility of the existence of the regular Eigen (H_3O^+) cation in such
47
48 aqueous solutions was ruled out by the absence in the mentioned affected spectra of the characteristic
49
50 OD stretching band ($\nu^{\circ}_{\text{OD}} = \sim 2050 \text{ cm}^{-1}$), corresponding to strong H-bonds in the H_3O^+ first hydration
51
52 sphere.⁹ Its presence in the carboxylic acid-affected HDO spectra strongly suggests the possibility of
53
54 existence of $\text{RCOO}^-\cdots\text{H}_3\text{O}^+$ CIPs in a measurable proportion. The resulting Eigen-type cation would
55
56 then be responsible for the appearance of the $\sim 2050 \text{ cm}^{-1}$ band. Let us mention briefly that previous
57
58
59
60

1 experimental evidence supporting the existence of CIPs in aqueous solutions of carboxylic acids is
2 rather scarce. They were indirectly detected in Raman spectra by splitting of the C–C stretching band
3 only. These spectral features were observed solely in heavy water, so the higher acidity of carboxylic
4 acids in D₂O was suggested as the possible explanation of the phenomenon.¹³
5
6
7
8

9 The tentative interpretation of the component bands in the carboxylic acid-affected HDO spectra
10 outlined above may be strengthened in confrontation with electronic structure calculations. However,
11 when such a comparison is made, a direct comparison of H-bond lengths' distribution is beneficial. The
12 vibrational spectra of HDO in the OD stretching range may be transformed to the interatomic oxygen-
13 oxygen distance probability distribution via generalization⁴⁹ of the well-known empirical distance-
14 frequency relationships (see also below).⁴⁴ This provides a direct structure-spectrum relationship in
15 terms of the H-bond length distribution in the affected solvent. The probability distributions resulting
16 from the calculated affected spectra are shown in Figure 3 and compared to the bulk HDO case. It is
17 evident that these curves are highly structured and once again discriminate between the very similar
18 acetic and propionic acids and the stronger formic acid. The strongly red-shifted component bands in the
19 affected spectra are responsible for the appearance of probability distribution features at ca. 2.53 Å and
20 slightly above 2.6 Å, indicating the presence of short hydrogen bonds.
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36

37 The existence of such strong and short hydrogen bonds may be verified in static QM calculations.
38 Representative carboxylic acids aqueous clusters studied in this work are illustrated in Figure 4 (more
39 are available as Supporting Information). While increasing the size of the clusters, we noticed that water
40 molecules tend to form H-bonded rings with the acid molecule: one around the –OH group of the acid,
41 one around the =O group and a third one connecting the two H-bonded networks in between the two
42 functional groups. Adopting the previously proposed nomenclature, these rings are designated by
43 counting the number of heavy (i.e., non-hydrogen) atoms.⁵⁸ Therefore, the cluster symbol XYZ denotes
44 X heavy atoms forming the ring around the –OH site, Z heavy atoms forming the ring around the =O
45 site, and Y heavy atoms forming the “bridging” ring. Each of the clusters presented was optimized for
46 the three studied acids, however, no major structural changes were observed, apart from slight changes
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 in hydrogen bond lengths (see Table 2). The major difference in the gas phase between formic acid and
2 the other two acids comes from the 474 cluster, which is a CIP for formic acid and neutral otherwise.
3

4 We found previously that the parameter most useful in the comparison with the experimental affected
5 HDO spectra is the hydrogen bond length, measured by the interatomic oxygen-oxygen distance (R_{OO}).
6 The R_{OO} value can be easily transformed to the OD band position of HDO (ν_{OD}^o) with the aid of
7 empirical relations. Eq. (2) linking ν_{OD}^o to R_{OO} was established on the basis of vibrational spectra and
8 neutron diffraction measurements on crystalline hydrates.⁴⁴
9

$$10 \nu_{OD}^o / \text{cm}^{-1} = 2727 - \exp[16.01 - 3.73(R_{OO} / \text{\AA})] \quad (2)$$

11 Although this procedure seems like a roundabout route to the OD stretching frequency, which might
12 as well be simply determined from QM frequency calculations, we found formerly that the stretching
13 frequencies determined from the molecular geometry with the aid of the empirical relation (2) correlate
14 much better with the experimental component bands.^{9,45,46} The hydrogen bond lengths and the
15 corresponding calculated frequencies (ν_{OD}^c) for the studied solutes are shown in Table 2, averaged over
16 different molecular situations in all clusters (individual H-bond lengths are available in the Supporting
17 Information). The hydrogen bond types encountered in the studied clusters in both neutral and contact
18 ion pair form are schematically illustrated in Fig. 5.
19

20 The most noteworthy feature of our gas phase calculations is the existence of stable ion pairs (the 464,
21 474 and 574 structures for formic acid, and the 464 and 574 structures for the other acids). Previously
22 investigated aqueous clusters of carboxylic acids were usually too small to allow for stable energetic
23 minima corresponding to CIP structures.²¹⁻²⁵ However, explicit consideration of more solvating water
24 molecules led to stable minima matching CIP and solvent-separated ion pair (SSIP) cases for both
25 formic¹⁴ and acetic acid¹⁵ (by solvent-separated ion pair we denote a structure, in which at least one
26 solvent molecule is positioned directly between the acidic anion and the hydrated proton). A formic acid
27 cluster with 4 H₂O allowed for a stable SSIP structure, while an additional water molecule stabilized
28 also the CIP structure.¹⁴ For both types of ion pairs, the hydrated proton was clearly an Eigen-type cation
29 (H_3O^+).^{14,15}
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 The difference between the previous computational results and those obtained herein lies in the
2 relative stability of the different isomers. To illustrate this, selected thermodynamic data for the studied
3 clusters are included in Table 3. For example, the 464 cluster is a stable minimum on the potential
4 energy surface (PES) at the B3LYP/6-311++G(d,p) level of theory and has a lower energy (including
5 ZPVE) than the corresponding 464t cluster, which is also a stable local minimum at the same level. This
6 is the first time that such an observation is made and all previously reported CIP or SSIP structures lay
7 above the respective neutral clusters on the PES.^{14,15} The 464t cluster is formed from 464 by a simple
8 proton transfer through the inner ring of three water molecules. Such an arrangement for proton transfer
9 was observed formerly in MD simulations.¹⁶ At the same time, the reaction free energy for the 464t →
10 464 isomerisation in the gas phase is very close to zero (Table 3) and the process is almost at an ideal
11 equilibrium. In a previous QM investigation, gas phase ΔG for isomerisation of acetic acid to an ion pair
12 was also found to be $-0.1 \text{ kJ}\cdot\text{mol}^{-1}$.¹⁵ However, inclusion of the solvation free energy of the reactants
13 obtained in the PCM/SMD approximation makes free energy more positive and the resulting
14 equilibrium constant is on the order of 10^{-3} (Table 3).

15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33 The most interesting fact from the point of view of correlation with the spectral data is the dramatic
34 structural change when going from neutral to CIP clusters. The C-OH...OH₂ hydrogen bond length is
35 2.615 Å in the gas phase for formic acid and 2.625 Å for the other acids (averaging over all the studied
36 acid clusters, see Table 2, bond type IV). When in turn the cluster attains a CIP configuration, the
37 resulting C-O...H₃O⁺ hydrogen bond shortens considerably to ~2.55 Å (see Table 2). The even more
38 important difference between these two isomers is the equally pronounced shortening of the two
39 hydrogen bonds formed with the other water molecules by the water molecule that is the proton acceptor
40 from the acid (although the anion-cation H-bond still remains the shortest one of the three). Therefore,
41 the Eigen-type H₃O⁺ cation is a valid description of the formed individual, as the H-bond length of
42 2.50-2.55 Å is characteristic for this case.⁹

43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Several concomitant phenomena due to the proton transfer in the CIP structures can be observed. The
shortening of the two water-acid H-bonds to the C=O group, when it becomes a C-O⁻ group in a CIP

1 structure, is apparent (from ~ 2.87 Å for formic acid and ~ 2.85 Å for the other two acids to ~ 2.79 - 2.8 Å).
2
3 It can be linked to the increased negative charge on the oxygen atom due to the charge separation in the
4
5 CIP structure. The CHelpG charge⁵⁹ of the relevant oxygen atom changes from -0.590 in 464t to -0.748
6
7 in 464 (for formic acid). At the same time, the charge of the C-O-H oxygen atom changes from -0.689
8
9 in 464t to -0.795 after dissociation, with a similar change in the hydration structure. Apart from the
10
11 obvious shortening of the H-bond connected with proton transfer, described above, the long H-bond
12
13 donated by a water molecule to the $-OH$ oxygen atom of the acid also shortens from 2.88 - 2.86 Å to
14
15 ~ 2.73 Å, when the acid dissociates. Averaging over all three H-bonds to $C-O^-$ (two to the former $C=O$
16
17 oxygen atom and one to the former $-OH$ oxygen atom), the value ~ 2.77 Å is obtained for all three acids.
18
19 Previously for analogous carboxylates we found a much more pronounced asymmetry in the hydrogen
20
21 bond lengths between water and the two carboxylic oxygens, leading to instantaneous charge
22
23 localization on the carboxylate anion visible also in the affected HDO spectra.⁵⁵ The charge differences
24
25 reported previously for the two oxygens were on the order of 0.1 ,⁵⁵ which is observed here only for the
26
27 neutral form of acidic clusters. Therefore, the presence of the hydronium cation leads to the marked
28
29 charge delocalization in the CIP clusters, even though the cation itself is positioned asymmetrically with
30
31 respect to the carboxylic anion in the 464, 474 and 574 clusters. However, the cation may transiently
32
33 fluctuate within the water chain connecting the two carboxylic oxygens¹⁶ and possible transition states
34
35 with the hydronium cation located symmetrically between the two oxygens were reported before.¹⁹
36
37 Consequently, this may lead to symmetrization of the interactions with the hydrating water and the
38
39 picture observed in the static infrared spectra is a statistical average of the rapidly fluctuating Eigen
40
41 cation.
42
43
44
45
46
47
48

49 The geometry of an aqueous cluster obtained in the gas phase is usually overstructured with respect to
50
51 the situation in the solvent. We found previously that the application of the PCM approach to solvation
52
53 provides much better agreement with the interatomic oxygen-oxygen distances obtained from the
54
55 experimental spectra.^{9,45,46} Also here, the H-bond lengths obtained with PCM are greater than the
56
57 respective gas phase values, with the SMD model systematically predicting longer H-bonds on average
58
59
60

1 than the UFF atomic radii-based model (Table 2). The main deficiency of the UFF model is the lack of
2 Eigen-type CIPs for acetic and propionic acids (clusters 464, 474, and 574). The SMD model performs
3
4 better in this respect, failing to predict a CIP structure only for 474 (the same as in the gas phase results
5
6 described above).
7
8

9
10 The average H-bond lengths, especially those obtained using the PCM/SMD approach, can now
11 enable us to thoroughly interpret the observed analytical bands in the carboxylic acid-affected HDO
12 spectra. Especially the structural changes between neutral and CIP clusters are reflected as equally
13 pronounced changes in the position of the analytical component bands. The band positions at maximum
14 after empirical transformation to the intermolecular oxygen-oxygen distance (Table 1) can be directly
15 compared with the respective average distances obtained from QM calculations (Table 2).
16
17
18
19
20
21
22

23
24 As already mentioned above, the proton transfer from acid to water results in a red shift of the band
25 position from $\sim 2260\text{ cm}^{-1}$ to $\sim 2050\text{ cm}^{-1}$ and the decrease of the H-bond length from 2.62-2.65 Å to
26 $\sim 2.54\text{ Å}$, which is adequately reproduced in the PCM/SMD calculations, even with respect to the
27 difference between formic acid and the other two acids. Simultaneously the bands at $\sim 2625\text{ cm}^{-1}$ and
28 $\sim 2550\text{ cm}^{-1}$ shift to $\sim 2430\text{-}2440\text{ cm}^{-1}$, in the latter case from the position typical to the C=O group to the
29 position typical to a negatively charged C-O⁻ group.⁵⁵ This time the agreement with QM calculations is
30 less impressive, however, the characteristic changes in H-bond lengths in this process are preserved,
31 with H-bonds from the calculations generally shorter on average than the values obtained from the
32 experimental band positions, especially for the two blue-shifted bands.
33
34
35
36
37
38
39
40
41
42
43
44

45 As all these bands are observed simultaneously in a static vibrational spectrum, equilibrium between
46 the neutral and the CIP structure must be present. At the very least, the lifetime of each species should
47 be longer than the characteristic time of the OD stretching vibration ($\sim 13.3\text{ fs}$). Actually, the lifetime of
48 the CIP form as predicted from MD simulations is much longer. The estimation based on Car-Parrinello
49 MD was ca. 0.2-0.3 ps,¹⁶ although it was also termed a “metastable intermediate” only.¹⁷ The
50 deprotonation of formic acid (FA) could also be followed by subsequent reprotonation in $\sim 0.7\text{ ps}$,
51 leading through a SSIP intermediate and with proton transfer facilitated by a ring of 3 water molecules¹⁶
52
53
54
55
56
57
58
59
60

(cf. the arrangement of water molecules in the clusters 464 and 464t). In a single classical MD simulation with an algorithm allowing proton transfer acts, frequent proton shuttling between acetic acid and H-bonded water molecule was observed and this process seemed to be driven by environmental fluctuations.²⁰ Therefore, the observation of the OD stretching bands characteristic for CIP structures is entirely possible.

The actual proportion of the CIP to neutral form of the acid may be obtained from integrated intensities of the respective component bands, viz. $\sim 2260\text{ cm}^{-1}$ and $\sim 2050\text{ cm}^{-1}$. However, the presence of an additional band at $\sim 1950\text{ cm}^{-1}$ for formic and acetic acid makes such comparison difficult. This band most probably reflects the asymmetry of the Eigen cation formed in a CIP structure and its integrated intensity grows with increasing acid strength. On the other hand, the absolute value of this intensity is uncertain, as the band is located at the edge of the studied region and is artificially broadened in the fitting procedure to account for baseline corrections. Therefore, only for propionic acid a truly meaningful integrated intensity comparison may be performed. Before such a comparison is made, the intensities have to be rescaled taking into account the well-known effect that the integrated intensity of the HDO stretching bands increases approximately linearly with a shift to lower wavenumbers.^{60,61} The general form of these relationships is expressed in Eq. (3), where the zero-intensity reference point $\nu^{\circ}_{\text{OD}} = 2727\text{ cm}^{-1}$ is the gas phase OD stretching band position.⁶²

$$I = C \cdot (2727 - \nu^{\circ}_{\text{OD}}) \quad (3)$$

On this basis, both bands' intensities were rescaled to their hypothetical values at the bulk HDO band center of gravity (2505 cm^{-1}) using Eq. (4) before calculating their ratio.

$$I^c = I \frac{2727 - 2505}{2727 - \nu^{\circ}_{\text{OD}}} \quad (4)$$

After such procedure the proportion of CIPs in aqueous solution as measured by the corrected intensity ratio of the two characteristic bands ($I^c_{2050} / I^c_{2260} + I^c_{2050}$) is $\sim 12\%$ for propionic acid. The proportion of CIPs basing on the PCM/SMD value of $\Delta G^{\circ}_{\text{aq}}$ (Table 3) is an order of magnitude lower and amounts $\sim 0.9\%$. Results comparable to our experimental data were obtained in CPMD simulations,

1 where the free-energy barrier for dissociation of neutral formic acid to CIP was $\sim 4 \text{ kcal mol}^{-1}$.¹⁶ In a
2 different study, the proportion of the CIP forms was estimated at $\sim 3\%$.²⁰ In the latter case, the pH of the
3 simulated solution was equal to 1, so a shift of the dissociation equilibrium towards the neutral form
4 might be expected in comparison to our results. All these figures are much higher than the dissociation
5 degree of propionic acid, which is ca. 0.37% at the standard concentration of 1 mol dm^{-3} . However, this
6 might be expected if the CIP formation is the first step in the proton transfer away from the anion that
7 ultimately results in the acid dissociation.^{16-18,20}

19 Conclusions

20 The hydration of the $-\text{COOH}$ group viewed from the perspective of the affected HDO spectra might
21 be roughly described as an “additive” phenomenon. In particular, the hydration of the neutral form of the
22 acid is well described as a combination of HDO affected by the $\text{C}=\text{O}$ group and the $\text{C}-\text{OH}$ group. The
23 former was previously studied by us in aqueous solutions of simple ketones (acetone and 2-butanone)⁵⁶
24 and gives rise to a component band at $\nu^{\circ}_{\text{OD}} \approx 2540 \text{ cm}^{-1}$, as also observed here. In turn, two component
25 bands are attributable to the $\text{C}-\text{OH}$ group affected HDO. The $\text{C}-\text{OH}$ hydrogen bond to water gives rise to
26 a component at ca. $\sim 2260 \text{ cm}^{-1}$ (cf. analogous $\text{P}-\text{OH}$ hydrogen bond to water in aqueous phosphoric(V)
27 acid at 2310 cm^{-1})⁴⁶, while the H -bond from water to the $\text{C}-\text{OH}$ oxygen atom is accountable for a
28 component at 2625 cm^{-1} (also here there is a close correspondence to our previous study of H_3PO_4).⁴⁶
29 Water engaged in hydrophobic hydration of the alkyl chain is expected to contribute to the band at
30 $\sim 2540 \text{ cm}^{-1}$, as evidenced before for aqueous *sec*-butylamine.¹ The increase of the integrated intensity of
31 this band for acetic and propionic acids with respect to formic acid (Table 2) seems to confirm this
32 hypothesis.

33 For the contact ion pair isomer of a carboxylic acid, the component bands characteristic to the $\text{C}-\text{O}^-$
34 group and the Eigen form of the hydrated proton (H_3O^+) are visible in the spectra. The latter was
35 previously not found by us in the strongly acidic solutions, where the “deformed Zundel” form of the
36 hydrated proton prevails.⁹ However, the characteristic ν°_{OD} value, predicted for it on the basis of ab
37

1 initio calculations⁹ ($\sim 2050\text{ cm}^{-1}$) is readily identified in the present affected HDO spectra (Table 1). In
2
3 turn, the hydration of the C-O⁻ group is closely related to the hydration of hydroxide anion (OH⁻). The
4
5 negative charge on the oxygen atom causes a red shift of the OD stretching vibration and the value
6
7 found previously for OH⁻ (2437 cm^{-1})⁴⁵ and carboxylate anions⁵⁵ is strikingly close to the component
8
9 band position at $2430\text{-}2440\text{ cm}^{-1}$ found in this work.

10
11 The presence of both sets of the discussed component bands in the affected HDO spectra implies that
12
13 the interaction of the carboxylic acid with the hydrating water molecules leads to the formation of
14
15 contact ion pairs in an appreciable proportion ($\sim 12\%$ as inferred from the respective bands' integrated
16
17 intensities), in agreement with numerous MD studies.^{16-18,20} The neutral to CIP interconversion in
18
19 aqueous clusters seems to be facilitated by a ring of three water molecules, as also evidenced by the
20
21 recent MD study.¹⁶ This process ultimately leads to the acid dissociation and its equilibrium constant
22
23 obtained in the PCM/SMD approximation is on the order of 10^{-3} (Table 3), at least an order of
24
25 magnitude greater than the macroscopic acid dissociation constant.² Apart from the computational
26
27 evidence, the changes in the hydration structure between the neutral and CIP acid forms are directly
28
29 visible in the affected HDO spectra, as mentioned above, which offer possibly the first experimental
30
31 proof that such equilibrium actually exists in aqueous solution, since only some indirect clues were
32
33 reported before.¹³
34
35
36
37
38
39
40
41
42

43 **Acknowledgments.** This work was supported from the Republic of Poland scientific funds as a
44
45 research project, within grant no. N N204 3799 33. Calculations were carried out at the Academic
46
47 Computer Center in Gdańsk (TASK).
48
49
50

51 **Supporting Information Available:** Experimental spectra of the studied aqueous solutions in the OD
52
53 stretching range of HDO. Gas phase optimized structures of the carboxylic acids' aqueous clusters not
54
55 included in Figure 4. Individual hydrogen bond lengths in the optimized aqueous clusters. This material
56
57 is available free of charge via the Internet at <http://pubs.acs.org>.
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

References

- (1) Gojło, E.; Śmiechowski, M.; Stangret, J. *J. Mol. Struct.* **2005**, *744-747*, 809.
- (2) Lide, D. R., Ed. *CRC Handbook of Chemistry and Physics*, 90th Ed., CRC Press, Boca Raton, FL, 2010, p. 8–42.
- (3) Marx, D.; Tuckerman, M. E.; Hutter, J.; Parrinello, M. *Nature* **1999**, *397*, 601.
- (4) Voth, G. A. *Acc. Chem. Res.* **2006**, *39*, 143.
- (5) Botti, A.; Bruni, F.; Ricci, M. A.; Soper, A. K. *J. Chem. Phys.* **2006**, *124*, 014508.
- (6) Asthagiri, D.; Pratt, L. R.; Kress, J. D. *Proc. Natl. Acad. Sci. U.S.A.* **2005**, *102*, 6704.
- (7) Stoyanov, E. S.; Stoyanova, I. V.; Reed, C. A. *J. Am. Chem. Soc.* **2010**, *132*, 1484.
- (8) Śmiechowski, M.; Stangret, J. *J. Mol. Struct.* **2008**, *878*, 104.
- (9) Śmiechowski, M.; Stangret, J. *J. Chem. Phys.* **2006**, *125*, 204508.
- (10) Zundel, G. In *The Hydrogen Bond - Recent Developments in Theory and Experiments*; Schuster, P.; Zundel, G.; Sandorfy, C., Eds.; North-Holland: Amsterdam, 1976; Vol. 2, pp 683-766.
- (11) Leuchs, M.; Zundel, G. *J. Chem. Soc. Faraday Trans. II* **1980**, *76*, 14.
- (12) Max, J.-J.; Chapados, C. *J. Phys. Chem. A* **2004**, *108*, 3324.
- (13) Génin, F.; Quilès, F.; Burneau, A. *Phys. Chem. Chem. Phys.* **2001**, *3*, 932.
- (14) Chuhev, K.; BelBruno, J. J. *J. Mol. Struct. (Theochem)* **2006**, *763*, 199.
- (15) Yamabe, S.; Tsuchida, N. *J. Comput. Chem.* **2003**, *24*, 939.

- 1 (16) Lee, J.-G.; Ascitutto, E.; Babin, V.; Sagui, C.; Darden, T.; Roland, C. *J. Phys. Chem. B* **2006**,
2
3 *110*, 2325.
4
5
6 (17) Park, J. M.; Laio, A.; Iannuzzi, M.; Parrinello, M. *J. Am. Chem. Soc.* **2006**, *128*, 11318.
7
8
9 (18) Thomas, V.; Maurer, P.; Iftimie, R. *J. Phys. Chem. B* **2010**, *114*, 8147.
10
11
12 (19) Wei, D.; Truchon, J.-F.; Sirois, S.; Salahub, D. *J. Chem. Phys.* **2002**, *116*, 6028.
13
14
15 (20) Gu, W.; Frigato, T.; Straatsma, T. P.; Helms, V. *Angew. Chem. Int. Ed.* **2007**, *46*, 2939.
16
17
18 (21) Aloisio, S.; Hintze, P. E.; Vaida, V. *J. Phys. Chem. A* **2002**, *106*, 363.
19
20
21 (22) Zhou, Z.; Shi, Y.; Zhou, X. *J. Phys. Chem. A* **2004**, *108*, 813.
22
23
24 (23) Priem, D.; Ha, T.-K.; Bauder, A. *J. Chem. Phys.* **2000**, *113*, 169.
25
26
27 (24) Hsieh, Y.-H.; Weinberg, N.; Yang, K.; Kim, C.-K.; Shi, Z.; Wolfe, S. *Can. J. Chem.* **2005**, *83*,
28
29 769.
30
31
32 (25) Gao, Q.; Leung, K. T. *J. Chem. Phys.* **2005**, *123*, 074325.
33
34
35 (26) George, L.; Sander, W. *Spectrochim. Acta A* **2004**, *60*, 3225.
36
37
38 (27) Yamamoto, K.; Nishi, N. *J. Am. Chem. Soc.* **1990**, *112*, 549.
39
40
41 (28) Ruderman, G.; Caffarena, E. R.; Mogilner, I. G.; Tolosa, E. J. *J. Solution Chem.* **1998**, *27*, 935.
42
43
44 (29) Billeter, S. R.; van Gunsteren, W. F. *J. Phys. Chem. A* **2000**, *104*, 3276.
45
46
47 (30) SanRomán–Zimbrón, M. L.; Ortega–Blake, I. *J. Chem. Phys.* **1997**, *107*, 3253.
48
49
50 (31) Iftimie, R.; Salahub, D.; Wei, D.; Schofield, J. *J. Chem. Phys.* **2000**, *113*, 4852.
51
52
53 (32) Nishi, N.; Nakabayashi, T.; Kosugi, K. *J. Phys. Chem. A* **1999**, *103*, 10851.
54
55
56 (33) Kosugi, K.; Nakabayashi, T.; Nishi, N. *Chem. Phys. Lett.* **1998**, *291*, 253.
57
58
59
60

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
- (34) Ng, J. B.; Shurvell, H. F. *J. Phys. Chem.* **1987**, *91*, 496.
- (35) Tanaka, N.; Kitano, H.; Ise, N. *J. Phys. Chem.* **1990**, *94*, 6290.
- (36) Tanaka, N.; Kitano, H.; Ise, N. *J. Phys. Chem.* **1991**, *95*, 1503.
- (37) Semmler, J.; Irish, D. E. *J. Solution Chem.* **1988**, *17*, 805.
- (38) Waldstein, P.; Blatz, L. A. *J. Phys. Chem.* **1967**, *71*, 2271.
- (39) Kaatze, U.; Menzel, K.; Pottel, R. *J. Phys. Chem.* **1991**, *95*, 324.
- (40) Takamuku, T.; Kyoshoin, Y.; Noguchi, H.; Kusano, S.; Yamaguchi, T. *J. Phys. Chem. B* **2007**, *111*, 9270.
- (41) Granados, K.; Gracia-Fadrique, J.; Amigo, A.; Bravo, R. *J. Chem. Eng. Data* **2006**, *51*, 1356.
- (42) Verrall, R.E.; Lilley, T.H.; Conway, B.E.; Luck, W.A.P. In *Water, a Comprehensive Treatise*, Franks, F., Ed., Plenum, New York, 1973, Vol. 3.
- (43) Falk, M.; Ford, T. A. *Can. J. Chem.* **1966**, *44*, 1699.
- (44) Berglund, B.; Lindgren, J.; Tegenfeldt, J. *J. Mol. Struct.* **1978**, *43*, 179.
- (45) Śmiechowski, M.; Stangret, J. *J. Phys. Chem. A* **2007**, *111*, 2889.
- (46) Śmiechowski, M.; Gojłło, E.; Stangret, J. *J. Phys. Chem. B* **2009**, *113*, 7650.
- (47) Kristiansson, O.; Lindgren, J.; de Villepin, J. *J. Phys. Chem.* **1988**, *92*, 2680.
- (48) Stangret, J. *Spectrosc. Lett.* **1988**, *21*, 369.
- (49) Stangret, J.; Gampe, T. *J. Phys. Chem. B* **1999**, *103*, 3778.
- (50) Tomasi, J.; Mennucci, B.; Cancès, M. T. *J. Mol. Struct. (Theochem)* **1999**, *464*, 211.
- (51) Stephens, P. J.; Devlin, F. J.; Chabalowski, C. F.; Frisch, M. J. *J. Phys. Chem.* **1994**, *98*, 11623.

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
- (52) Gaussian 09, Revision A.02, Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, Jr., J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, N. J.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian, Inc., Wallingford CT, 2009.
- (53) Merrick, J. P.; Moran, D.; Radom, L. *J. Phys. Chem. A* **2007**, *111*, 11683.
- (54) Rappe, A. K.; Casewit, C. J.; Colwell, K. S.; Goddard III, W. A.; Skiff, W. M. *J. Am. Chem. Soc.* **1992**, *114*, 10024.
- (55) Gojłto, E.; Śmiechowski, M.; Panuszko, A.; Stangret, J. *J. Phys. Chem. B* **2009**, *113*, 8128.
- (56) Gojłto, E.; Gampe, T.; Krakowiak, J.; Stangret, J. *J. Phys. Chem. A* **2007**, *111*, 1827.
- (57) Berglund, B.; Lindgren, J.; Tegenfeldt, J. *J. Mol. Struct.* **1978**, *43*, 169.
- (58) Pu, L.; Wang, Q.; Zhang, Y.; Miao, Q.; Kim, Y. S.; Zhang, Z. B. *Adv. Quantum Chem.* **2008**, *54*, 271.
- (59) Breneman, C. M.; Wiberg, K. B. *J. Comp. Chem.* **1990**, *11*, 361.
- (60) Glew, D. N.; Rath, N. S. *Can. J. Chem.* **1971**, *49*, 837.
- (61) Bergström, P.-Å.; Lindgren, J.; Kristiansson, O. *J. Phys. Chem.* **1991**, *95*, 8575.

(62) Morozow, W. P.; Kwasha, N. T.; Caune, A. J.; Lisowienko, W. A. *Opt. Spectrosc.* **1963**, *15*, 617.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

TABLE 1: Selected Parameters of Analytical Component Bands from Decomposition of Spectra of HDO Affected by the Studied Carboxylic Acids

Solute	$\nu_{\text{OD}}^{\text{o}}$ ^a	fwhh ^b	<i>I</i> ^c	<i>R</i> _{OO} ^d
HCOOH	2627	76	916	3.058
	2558	104	2798	2.917
	2444	200	3680	2.779
	2212	227	1570	2.618
	2037	88	287	2.540
CH ₃ COOH	1956	182	874	2.510
	2625	86	1066	3.052
	2546	122	4818	2.899
	2432	143	2683	2.768
	2261	191	1587	2.645
C ₂ H ₅ COOH	2052	101	356	2.546
	1923	176	494	2.499
	2625	83	636	3.052
	2550	125	4844	2.905
	2432	166	2568	2.768
	2264	206	1399	2.647
	2054	111	278	2.546

^a Band position at maximum (cm⁻¹). ^b Full-width at half-height (cm⁻¹). ^c Integrated intensity (dm⁻³·mol⁻¹·cm⁻²). ^d Interatomic oxygen-oxygen distance obtained from $\nu_{\text{OD}}^{\text{o}}$ by an empirical relation.⁴⁴

TABLE 2: Average Hydrogen Bond Lengths in Different Structural Situations for the Optimized Aqueous Clusters of Carboxylic Acids (R-COOH)

solute	assignment ^a	gas		SMD		UFF	
		m ^b	R _{OO} ^c	m ^b	R _{OO} ^c	m ^b	R _{OO} ^c
R = H	I) C-O(H) ⋯ H-OH	7	2.885(69)	5	2.911(29)	5	2.884(25)
	II) C=O ⋯ H-OH	12	2.866(94)	11	2.883(63)	11	2.868(62)
	III) C-O ⁻ ⋯ H-OH	9	2.771(34)	9	2.806(46)	9	2.788(33)
	IV) C-OH ⋯ OH ₂	13	2.615(55)	13	2.620(25)	13	2.622(35)
	V) OH ₃ ⁺ ⋯ O	9	2.545(57)	9	2.567(25)	9	2.547(48)
R = CH ₃	I) C-O(H) ⋯ H-OH	8	2.859(54)	7	2.919(85)	9	2.906(90)
	II) C=O ⋯ H-OH	14	2.847(85)	13	2.864(52)	18	2.857(50)
	III) C-O ⁻ ⋯ H-OH	6	2.775(40)	6	2.796(36)	0	
	IV) C-OH ⋯ OH ₂	14	2.626(70)	14	2.641(28)	16	2.631(48)
	V) OH ₃ ⁺ ⋯ O	6	2.546(77)	6	2.565(60)	0	
R = C ₂ H ₅	I) C-O(H) ⋯ H-OH	8	2.857(56)	7	2.921(88)	9	2.904(93)
	II) C=O ⋯ H-OH	14	2.845(83)	13	2.859(46)	18	2.856(49)
	III) C-O ⁻ ⋯ H-OH	6	2.775(34)	6	2.799(35)	0	
	IV) C-OH ⋯ OH ₂	14	2.625(68)	14	2.647(29)	16	2.634(48)
	V) OH ₃ ⁺ ⋯ O	6	2.548(86)	6	2.565(71)	0	

^a The hydrogen bond in question is marked by ellipsis, the Roman numbers refer to Fig. 5. ^b Total number of hydrogen bonds of specific type in all clusters of a given solute. ^c Average intermolecular oxygen-oxygen distance (Å); standard deviation (as final digits of the value) given in parentheses.

TABLE 3: Selected Thermodynamic Data for the Optimized Aqueous Clusters of Carboxylic Acids (R-COOH)

solute	cluster ^a	E_0^b	ΔE_0^c	ΔH_{298}^d	ΔG_{298}^e	K^f	ΔG_{aq}^g	K_{aq}^h
R = H	464	-648.488809	-7.7	-11.5	0.1	0.95	13.0	$5.2 \cdot 10^{-3}$
	464t	-648.485881						
R = CH ₃	464	-687.796377	-3.2	-6.2	1.4	0.57	13.9	$3.6 \cdot 10^{-3}$
	464t	-687.795170						
R = C ₂ H ₅	464	-727.092638	-6.7	-9.6	-0.4	1.18	11.8	$8.6 \cdot 10^{-3}$
	464t	-727.090103						

^a The clusters are identified by their symbols from Figure 4. ^b Total energy, including scaled zero-point vibrational energy (hartree). ^c Energy difference between 464 and 464t at 0 K (kJ·mol⁻¹). ^d Enthalpy difference between 464 and 464t at 298 K (kJ·mol⁻¹). ^e Gibbs free energy difference between 464 and 464t at 298 K (kJ·mol⁻¹). ^f Equilibrium constant for the 464t → 464 process at 298 K. ^g Gibbs free energy difference between 464 and 464t in aqueous solution (SMD solvation model) at 298 K (kJ·mol⁻¹). ^h Equilibrium constant for the 464t → 464 process in aqueous solution (SMD solvation model) at 298 K.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Figure 1. (a) The derivatives $(\partial\epsilon/\partial m)_{m=0}$ and (b) the affected HDO spectra based on them for formic (green line), acetic (red line) and propionic (blue line) acid. The bulk HDO spectrum indicated with black dashed line in (b).

Figure 2. The spectra of HDO affected by (a) formic (green line), (b) acetic (red line) and (c) propionic (blue line) acid, and their decomposition into analytical bands (dashed lines).

Figure 3. Interatomic oxygen-oxygen distance distribution obtained from the affected HDO spectra in Fig. 1(b) for formic (green line), acetic (red line) and propionic (blue line) acid. The bulk HDO distance distribution indicated with dashed line.

Figure 4. Representative R-COOH clusters (R = H, CH₃, C₂H₅) studied in this work with both neutral and contact ion pair arrangement, optimized in the gas phase at the B3LYP/6-311++G(d,p) level for R = H. Hydrogen bonds indicated by thin lines, light grey spheres indicate hydrogens, dark grey – carbons, red – oxygens, blue – R substituents. See text for the explanation of cluster symbols.

Figure 5. Schematic R-COOH aqueous clusters in (a) neutral and (b) contact ion pair arrangement, with hydrogen bonds indicated by dashed lines. The Roman numbers over hydrogen bonds refer to hydrogen bond types listed in Table 2.

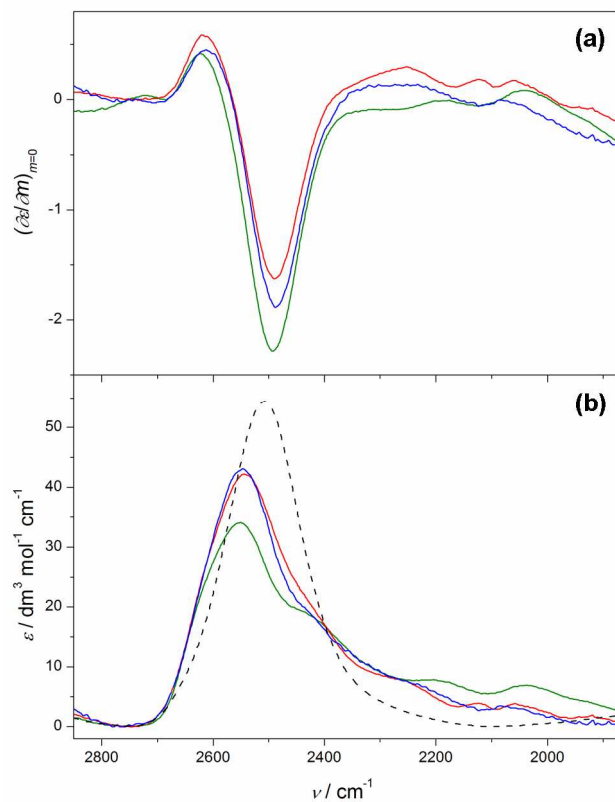


Figure 1. (a) The derivatives $(\partial \epsilon / \partial m)_{m=0}$ and (b) the affected HDO spectra based on them for formic (green line), acetic (red line) and propionic (blue line) acid. The bulk HDO spectrum indicated with black dashed line in (b).

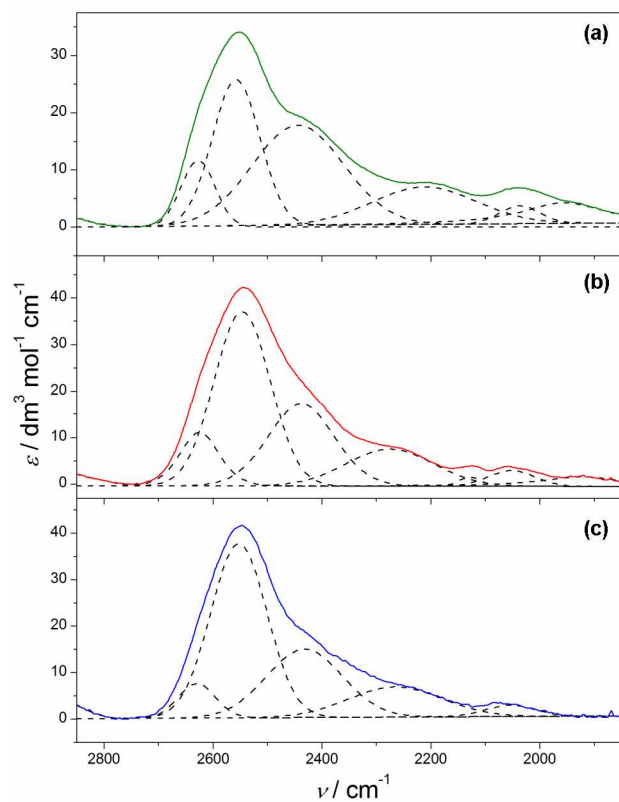


Figure 2. The spectra of HDO affected by (a) formic (green line), (b) acetic (red line) and (c) propionic (blue line) acid, and their decomposition into analytical bands (dashed lines).

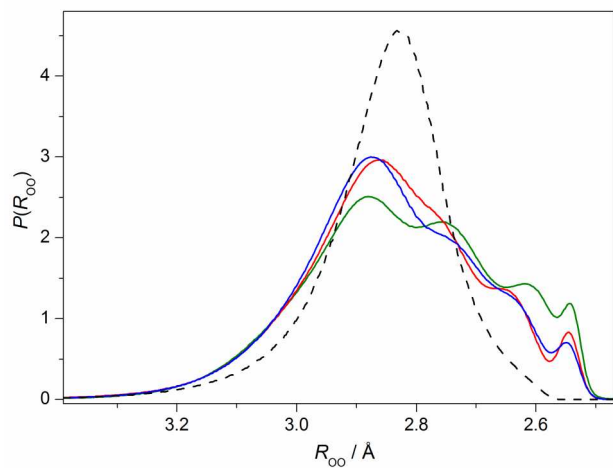


Figure 3. Interatomic oxygen-oxygen distance distribution obtained from the affected HDO spectra in Fig. 1(b) for formic (green line), acetic (red line) and propionic (blue line) acid. The bulk HDO distance distribution indicated with dashed line.

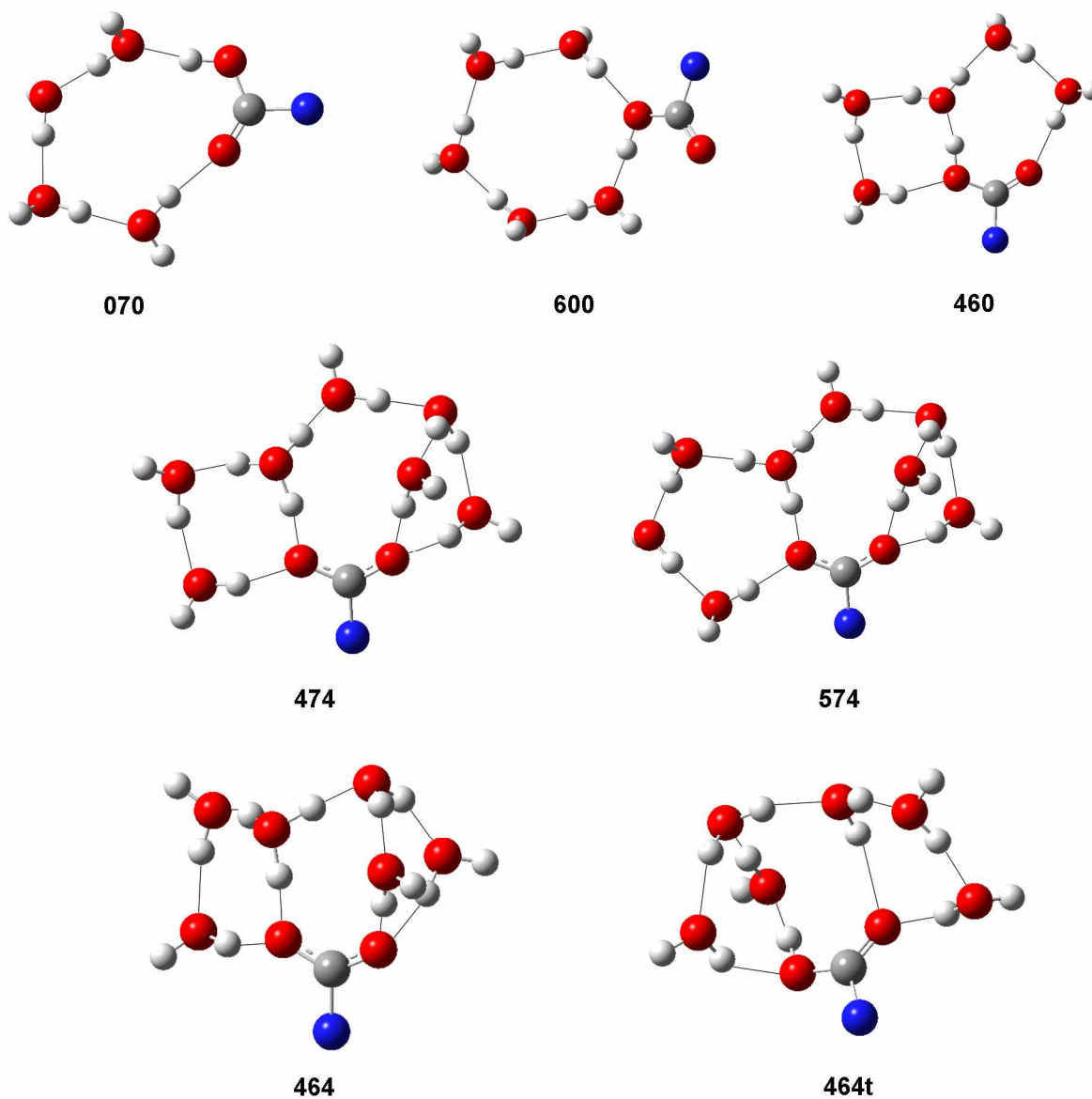
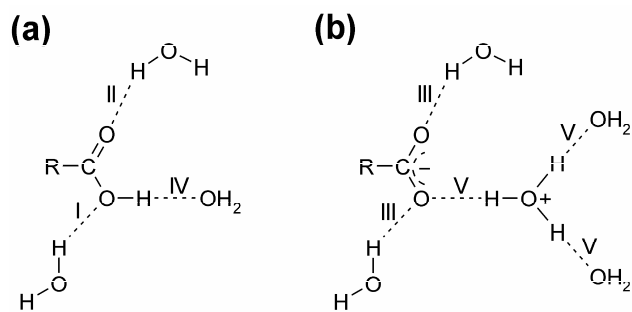


Figure 4. Representative R-COOH clusters (R = H, CH₃, C₂H₅) studied in this work with both neutral and contact ion pair arrangement, optimized in the gas phase at the B3LYP/6-311++G(d,p) level for R = H. Hydrogen bonds indicated by thin lines, light grey spheres indicate hydrogens, dark grey – carbons, red – oxygens, blue – R substituents. See text for the explanation of cluster symbols.



14 **Figure 5.** Schematic R-COOH aqueous clusters in (a) neutral and (b) contact ion pair arrangement, with
15 hydrogen bonds indicated by dashed lines. The Roman numbers over hydrogen bonds refer to hydrogen
16 bond types listed in Table 2.
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

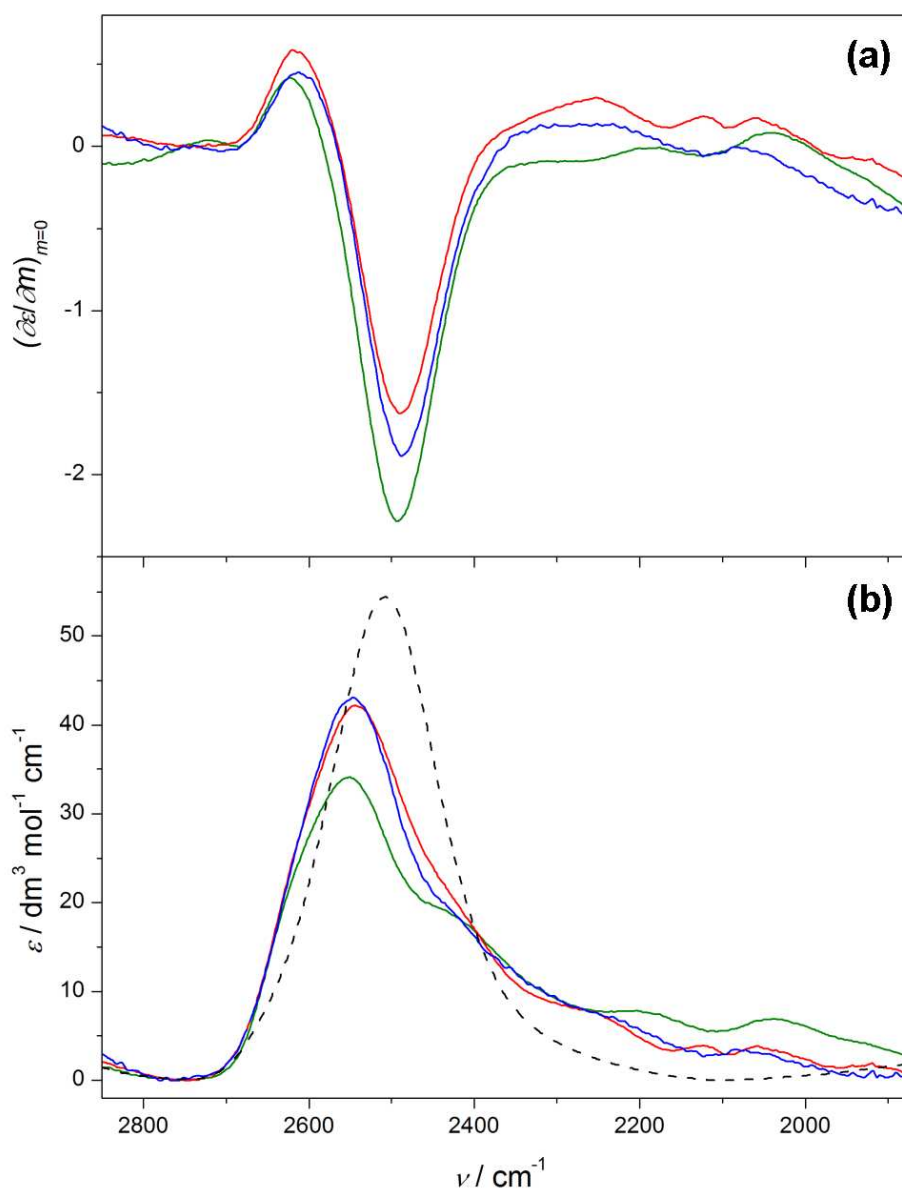


Figure 1. (a) The derivatives $(\partial\epsilon/\partial m)_{m=0}$ and (b) the affected HDO spectra based on them for formic (green line), acetic (red line) and propionic (blue line) acid. The bulk HDO spectrum indicated with black dashed line in (b).

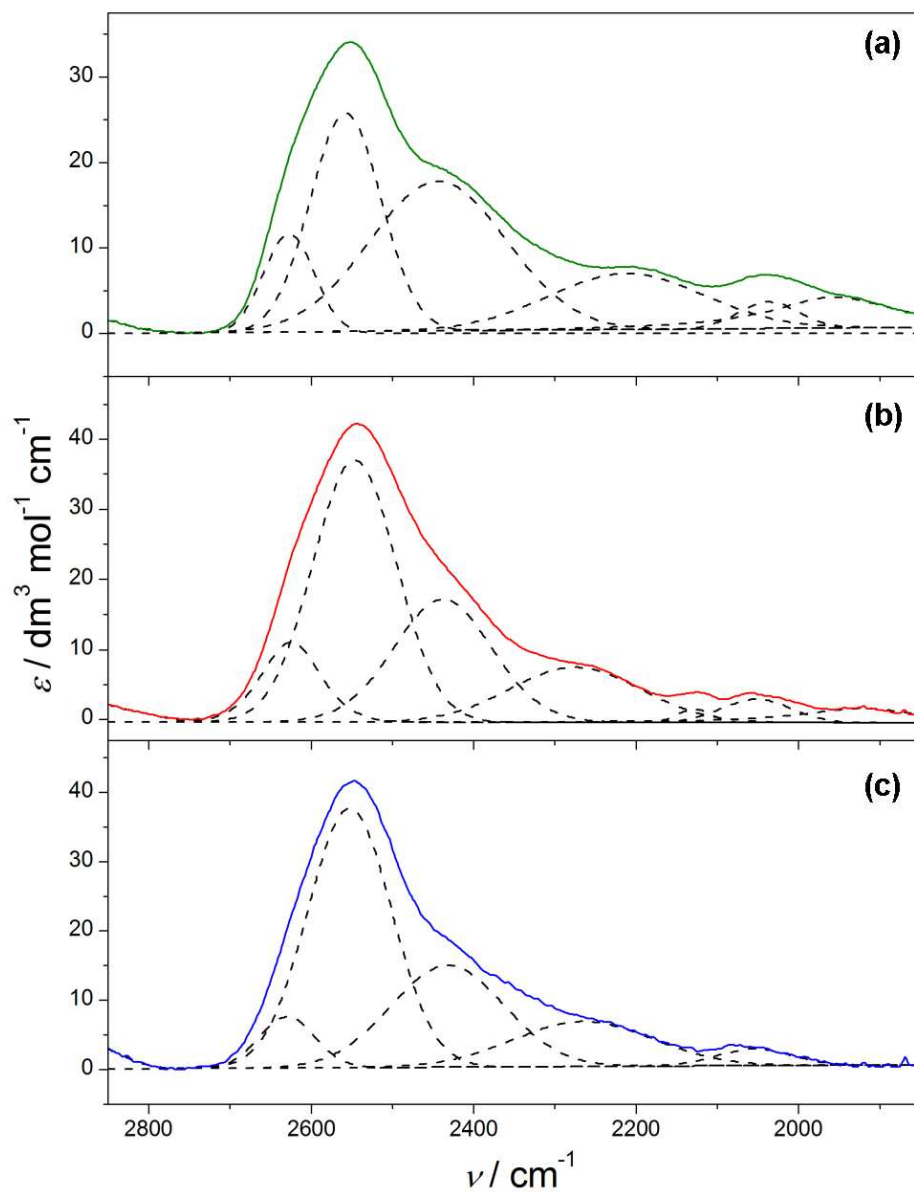


Figure 2. The spectra of HDO affected by (a) formic (green line), (b) acetic (red line) and (c) propionic (blue line) acid, and their decomposition into analytical bands (dashed lines).

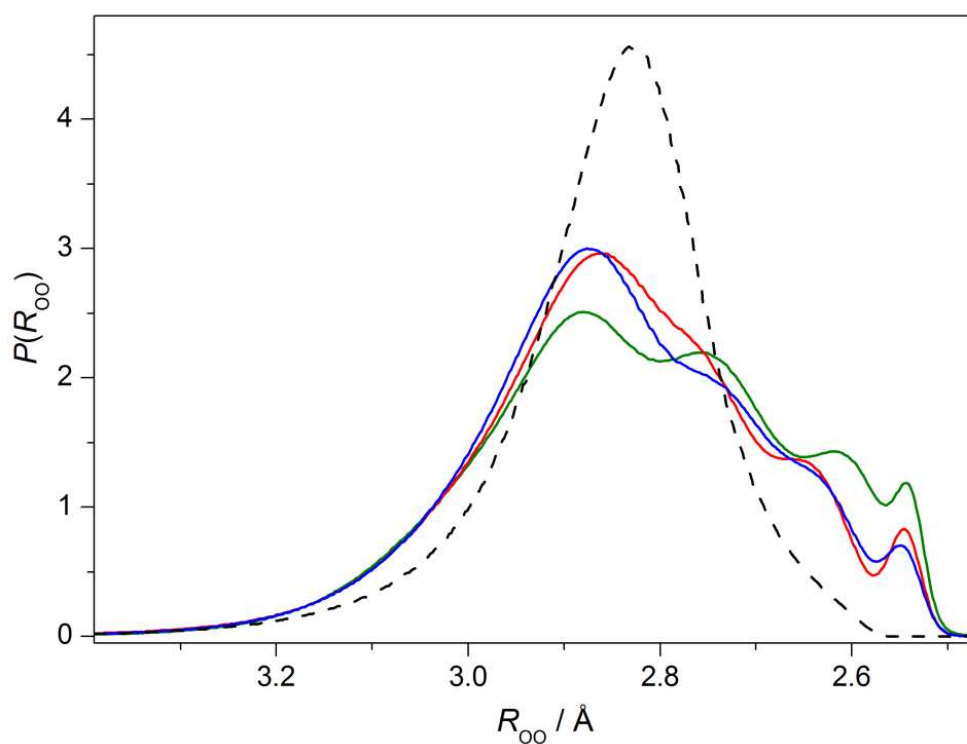


Figure 3. Interatomic oxygen-oxygen distance distribution obtained from the affected HDO spectra in Fig. 1(b) for formic (green line), acetic (red line) and propionic (blue line) acid. The bulk HDO distance distribution indicated with dashed line.

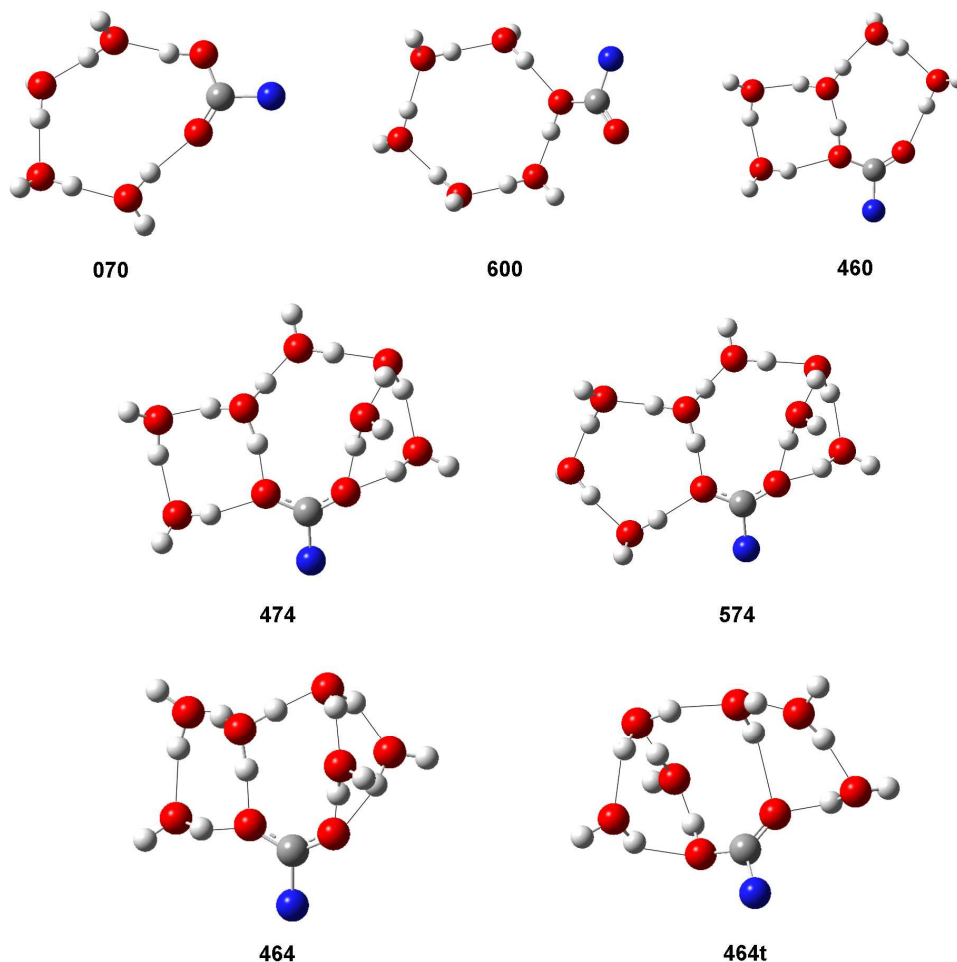


Figure 4. Representative R-COOH clusters (R = H, CH₃, C₂H₅) studied in this work with both neutral and contact ion pair arrangement, optimized in the gas phase at the B3LYP/6-311++G(d,p) level for R = H. Hydrogen bonds indicated by thin lines, light grey spheres indicate hydrogens, dark grey – carbons, red – oxygens, blue – R substituents. See text for the explanation of cluster symbols.

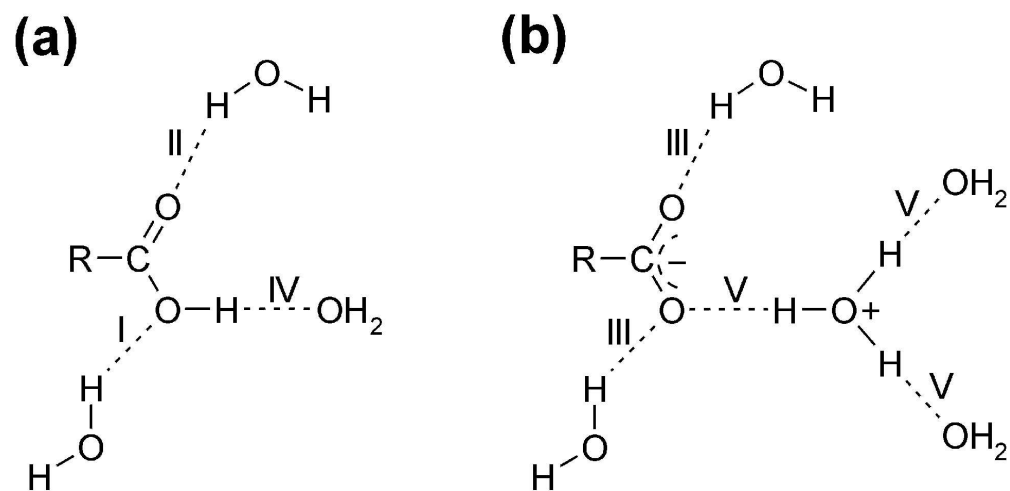


Figure 5. Schematic R-COOH aqueous clusters in (a) neutral and (b) contact ion pair arrangement, with hydrogen bonds indicated by dashed lines. The Roman numbers over hydrogen bonds refer to hydrogen bond types listed in Table 2.