

# Production of essential oils from in vitro cultures of *Caryopteris* species and comparison of their concentrations with in vivo plants

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**Abstract** The chemical composition of hydrodistilled essential oils obtained from aerial parts and roots of selected *Caryopteris* ('bluebeard') species (*C. incana*, *C. mongolica*, *Caryopteris* × *clandonensis*), as well as the newly established in vitro shoot and adventitious root cultures of the above plants, was analyzed by gas chromatography–mass spectrometry. Essential oil content and composition differed significantly depending on the type of plant material analyzed. Adventitious roots were characterized by the highest essential oil yield, reaching 1.8 % V/DW in *Caryopteris* × *clandonensis*. Limonene and cedrol were the main components of the essential oil derived from aerial parts of the intact plants (11.9–16.0 and 10.7–10.9 %, respectively), whereas the volatile fractions

of the in vivo roots of all species contained large amounts of 3,5-bis(1,1-dimethyl)-phenol (12.9–26.2 %). 1,8-cineole, absent in the intact plant materials, was the dominating volatile constituent of the essential oils obtained from in vitro shoots (24.8–34.2 %). The volatile oil derived from adventitious root cultures consisted primarily of 1-octen-3-ol (19.7–31.5 %) and medicinally relevant diterpenoids: abietatriene and *trans*-totarol, which were accumulated in considerable quantities, especially in the adventitious roots of *C. clandonensis* (21.6 and 29.2 %, respectively).

**Keywords** Abietatriene · Adventitious roots · *Caryopteris* · In vitro shoots · Totarol

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## Introduction

The genus *Caryopteris* [formerly placed in Verbenaceae, currently assigned to Labiatae (Cantino et al. 1999)] includes hardy, small, deciduous shrubs, native to eastern and southern Asia, and particularly well represented in China, Mongolia and Japan. Bluebeards, as they are commonly named, mostly grow on sunny mountain slopes in well-drained soil. Fragrant foliage and violet-blue flowers formed in the late summer and autumn are the characteristics of *Caryopteris* (Miller 2007). Bluebeards are included in traditional medicine systems as remedies for arthritic pains, cough, sore throat, bronchitis and eczema (Park et al. 2014).

Phytochemical studies of *Caryopteris* revealed the presence of iridoids, phenylpropanoids, phenolic acids, flavonoids and the biologically active essential oil (Park et al. 2014). Besides antimicrobial (Nedorostova et al. 2009) and insecticidal (Chu et al. 2011) properties, volatile

constituents of the investigated plants were shown to exhibit cytotoxic effects (Gao and Han 1997; Kim 2008), and can therefore be considered potential therapeutic agents.

In vitro cultures of higher plants can be regarded as an alternative, renewable source of high-value secondary metabolites (Murthy et al. 2014), with ginsenosides (Kochan et al. 2014; Liu et al. 2014), hypericin (Wu et al. 2014) and the anticancer lignan podophyllotoxin (Rajesh et al. 2014) being some of the well-known examples. As compared to field cultivation, in vitro techniques offer the possibility of continuous production of large amounts of chemically uniform biomass independently of wild resources and environmental factors (Murthy et al. 2014). As far as essential oils are concerned, in vitro cultivation also gives the possibility to obtain novel compounds, previously not reported in intact plants (Gounaris 2010; Gonçalves and Romano 2013; Marchev et al. 2014). Since the accumulation of volatile constituents was shown to be positively correlated with cell differentiation, most experiments aimed at establishing plant in vitro systems for essential oil production were conducted with the use of organ cultures. In this regard, adventitious and hairy roots are especially useful due to their fast growth and high yield of secondary metabolites (Gounaris 2010; Baque et al. 2012). Nevertheless, essential oils are often found in substantial amounts also in shoot cultures, such as those of *Arnica montana* (Petrova et al. 2012), *Thymus caespititius* (Mendes et al. 2013) and *Mentha* sp. (Hilton et al. 1995).

The aim of the study was to establish, for the first time, *Caryopteris* plant tissue cultures and evaluate their essential oil content, as well as chemical composition of the in vitro-derived volatile fractions. Although bluebeards are sometimes cultivated for ornamental value, their natural habitat, restricted due to climatic conditions, limits their utilization for medicinal purposes. Furthermore, the extraction of the volatile oil from soil-grown roots involves the destruction of the whole plant. For these reasons, the biotechnological research into plant cell cultures of *Caryopteris* was carried out. Three species were selected for the study: *Caryopteris incana* (Thunb.) Miq., *Caryopteris mongolica* Bunge and *Caryopteris × clandonensis* Simmonds, a hybrid between the two previous plants which so far has not been studied with respect to essential oil production. Given that organogenesis is known to positively affect the accumulation of volatile compounds (Gounaris 2010; Gonçalves and Romano 2013), the current research focused especially on establishing organ cultures. Gas chromatography coupled with mass spectrometry (GC–MS) was used to investigate the composition of hydro-distilled essential oil obtained from aerial and underground parts of *Caryopteris* intact plants, as well as shoots and roots maintained under in vitro conditions. The results were discussed and compared with the available literature data.

## Materials and methods

**Reagents and general procedures.** All reagents used for plant cell culture experiments were from Sigma-Aldrich (St. Louis, MO, USA). Water for media preparation and essential oil hydrodistillation was prepared using REL5 double water still (Mera-Polna, Przemysl, Poland). Unless otherwise stated, the in vitro cultures were maintained at  $26 \pm 1$  °C under white fluorescent light (16/24 photoperiod,  $40 \mu\text{mol m}^{-2} \text{s}^{-1}$ , TLD 35 W tubes, Philips, Amsterdam, the Netherlands). The rate of growth of the respective in vitro biomasses was expressed as the Growth index (Gi), calculated using the following formula:  $G_i = [(G_1 - G_0)/G_0] \times 100 \%$  where  $G_1$  is the fresh weight at the end of the growth cycle and  $G_0$  is the fresh weight of the inoculum.

### Seeds and in vivo plant material

Seeds of *Caryopteris incana* (Thunb.) Miq., *Caryopteris × clandonensis* Simmonds and *Caryopteris mongolica* Bunge were obtained from Sandeman seeds (London, UK) and used to grow the plants in vivo and to develop in vitro shoot and adventitious root cultures. The seeds for in vivo material were planted in the Medicinal Plants Garden of Medical University of Gdansk, Poland. The field-grown plants, interchangeably referred to as ‘intact plants’ or ‘in vivo plants’, were collected in October 2010 and separated to roots (referred to as ‘in vivo roots’) and aerial parts (also referred to as ‘herbs’). The intact plant materials were dried at 30 °C and subjected to essential oil analysis.

For in vitro culture initiation, the seeds of all species were sterilized with 0.1 %  $\text{HgCl}_2$  for 30 min, rinsed 3 times with sterile, double distilled water and placed in Petri dishes on absorbent paper soaked in a gibberellin  $A_3$  (28.9  $\mu\text{M}$ ) and kinetin (46.0  $\mu\text{M}$ ) solution. The dishes were kept in the dark at  $26 \pm 1$  °C. After 2 weeks, the seedlings were obtained and subsequently used for in vitro shoot and root culture initiation.

### Shoot in vitro cultures

For the induction of shoot cultures, the obtained *Caryopteris* seedlings were first placed on stationary (0.7 % w/v agar), phytohormone-free SH (Schenk and Hildebrandt 1972) medium supplemented with 3 % w/v sucrose and matured for 30 days. During that time, the seedlings formed 3–4 axillary shoots which were subsequently excised and placed on SH media supplemented with different sets of growth regulators (five explants per culture jar, three containers per modification). After 30 days, the cultures were evaluated for shoot induction (Table 1) and the medium modification providing the highest multiplication rates was

selected for further experiments. All the shoot cultures were maintained in baby food jars (Sigma-Aldrich) containing 25 ml medium and subcultured at 30-day intervals. For the analysis of volatiles, the plant material was repeatedly collected, on 30 days of the each growth cycle, dried at 30 °C and subjected to hydrodistillation.

#### Root in vitro cultures

For the induction of root cultures, the sterile *Caryopteris* seedlings were first placed on the stationary (0.7 % w/v agar), phytohormone-free SH medium supplemented with 3 % w/v sucrose and grown for 30 days. The roots were subsequently excised and cut into 1 cm fragments which were placed on SH media supplemented with different auxins. After 30 days, the explants were evaluated for root induction (Table 2) and the medium providing the highest number of roots per explant was selected for further experiments. For liquid culture initiation, the roots were

transferred to 250 ml Erlenmeyer flasks containing 100 ml of the selected medium, supplemented with 3 % w/v sucrose, and placed on orbital shaker (INNOVA 2300, Eppendorf, Enfield, US-CT, 25.4 mm stroke, 150 rpm). The obtained liquid root cultures were subcultured in 4-week intervals. For the analysis of essential oil, the plant material was repeatedly collected, on 30 days of the each growth cycle, dried at 30 °C and subjected to hydrodistillation.

**Isolation of volatile fraction.** For the isolation of volatiles, the dried (30 °C, 48 h), coarsely ground plant materials of the investigated *Caryopteris* spp. were subjected to 3 h hydrodistillation (20.0 g of material + 200 g of water) using the Deryng-type distillation apparatus (Baj et al. 2013) in accordance with the method quoted in Polish Pharmacopeia VI (2002). For all biomass types, the experiment was repeated in triplicate. After evaluating the essential oil yield (expressed as percentage V/DW ± SD), the volatile fraction was collected, dried over anhydrous sodium sulfate and subjected to GC–MS analysis.

**Table 1** The influence of plant growth regulators on shoot induction in *Caryopteris* in vitro cultures

Growth regulators (μM)	Number of shoots per explant <sup>a</sup>		
	<i>Caryopteris incana</i>	<i>Caryopteris × clandonensis</i>	<i>Caryopteris mongolica</i>
TDZ (1.00)	1 ± 0	2 ± 0	1 ± 0
2iP (9.84)	6 ± 1	5 ± 1	4 ± 1
BAP (8.88)	7 ± 1	7 ± 1	6 ± 1
TDZ (1.00) + 2iP (9.84)	15 ± 2	16 ± 2	14 ± 1
TDZ (1.00) + BAP (8.88)	25 ± 3	30 ± 3	27 ± 3

All cultures were maintained on solidified (0.7 % w/v agar) SH media supplemented with 3.0 % w/v sucrose

BAP 6-benzylaminopurine, 2iP 2-isopentenyladenine, TDZ thidiazuron

<sup>a</sup> Values represent the mean ± SD of 15 samples

**Table 2** The influence of plant growth regulators on root induction in *Caryopteris* in vitro cultures

Growth regulators (μM)	Number of roots per explant <sup>a</sup>		
	<i>Caryopteris incana</i>	<i>Caryopteris × clandonensis</i>	<i>Caryopteris mongolica</i>
NAA (5.37)	1 ± 0	2 ± 0	1 ± 0
IAA (5.71)	1 ± 0	0 ± 0	0 ± 0
IBA (4.92)	15 ± 2	10 ± 1	21 ± 3

All cultures were maintained on solidified (0.7 % w/v agar) SH media supplemented with 3.0 % w/v sucrose

IAA 3-indoleacetic acid, IBA indole-3-butyric acid, NAA 1-naphthaleneacetic acid

<sup>a</sup> Values represent the mean ± SD of 15 samples

#### GC–MS analysis

Chromatographic analysis was carried out on HP-1 30 m × 0.25 mm × 0.25 μm m capillary column, using gas chromatograph HP 5971A (Hewlett-Packard, Palo Alto, CA, USA). The analysis parameters were as follows: ionization energy 70 eV, carrier gas: helium, flow 0.5 ml min<sup>-1</sup>, temperature increase from 80 to 300 °C, at a rate of 8 °C min<sup>-1</sup>. The resulting spectra were compared with the data from the Mass Finder 2.1. library (König et al. 2001). All experiments were repeated three times.

#### Results and discussion

In view of the available literature data, the essential oil of *Caryopteris* is characterized by a great composition variability (Pu et al. 1984; Wen et al. 1990; Shatar and Adams 1999; Yang et al. 2005; Kim 2008; Upadhyaya et al. 2009; Yan and Wang 2009; Chu et al. 2011). To obtain a renewable source of standardized oil, in vitro shoot and adventitious root cultures of *Caryopteris* plants were initiated and established for the first time [except *C. incana* tissue cultures described by Zhang et al. (2008)].

In vitro shoot cultures of the respective species were obtained from sterile seedling fragments with the use of cytokinin-supplemented media. The plant growth regulators applied included two purine cytokinins (6-benzylaminopurine, BAP and isopentenyladenine, 2iP) and one synthetic phenylurea derivative (thidiazuron, TDZ). Due to possible synergistic effects between these two types of cytokinins in terms of shoot induction rate and positive

influence on explant morphology (Nielsen et al. 1995; Łuczkiwicz and Piotrowski 2005; Kokotkiewicz et al. 2012), BAP and 2iP were also applied jointly with TDZ. As presented in Table 1, the combination of 8.88  $\mu\text{M}$  BAP jointly with 1.00  $\mu\text{M}$  TDZ provided the highest number of shoots per explant in all the examined species. These cultures were also characterized by relatively fast growth, as indicated by their Gi values ranging from over 500 % for *C. mongolica* and *C. × clandonensis* to over 700 % for *C. incana* (Fig. 1).

Adventitious root cultures of *Caryopteris* were obtained from root fragments of sterile bluebeard seedlings. To promote root growth, the excised roots were placed on auxin-supplemented media. Among the modifications used, only indole-3-butyric acid-supplemented medium (4.92  $\mu\text{M}$  IBA) was shown to induce root formation (Table 2). Consequently, adventitious roots of the investigated species were further propagated in liquid SH medium of the above composition, providing biomass for phytochemical studies.

The established in vitro cultures were analyzed in terms of growth rate, essential oil content and composition, and the results were compared with the respective intact plants. As presented in Fig. 1, *C. incana* in vitro shoots and *C. mongolica* adventitious roots (depicted in Fig. 2a, b, respectively) showed the highest growth rates among the established shoot and root cultures, respectively. Adventitious roots of all investigated species were also characterized by higher growth index (Gi) values, as compared to the respective shoot cultures (Fig. 1).

The hydrodistillation of the dried plant materials gave essential oil yields ranging from ca. 0.4 to 0.8 % (herbs), 1.1 to 1.3 % (in vivo roots), 0.4 to 0.6 % (in vitro shoots) and 1.4 to 1.8 % (adventitious roots) (Fig. 3).

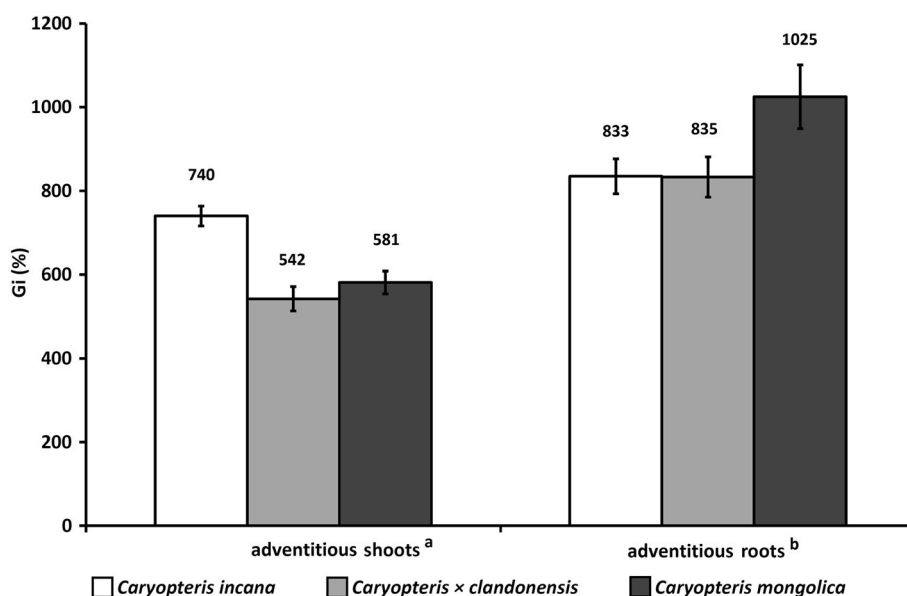
GC–MS of the examined essential oils revealed the presence of 64 compounds, listed in Table 3. All analyzed volatile fractions were characterized by high percentage of monoterpenes, followed by sesquiterpenes, which is in agreement with previous studies on *Caryopteris* (Upadhyaya et al. 2009). In general, the differences in terpenoid composition, observed between the particular materials, were mostly of quantitative nature. However, some of the compounds found in in vitro cultures were previously unreported, or present only in trace quantities in the intact plants.

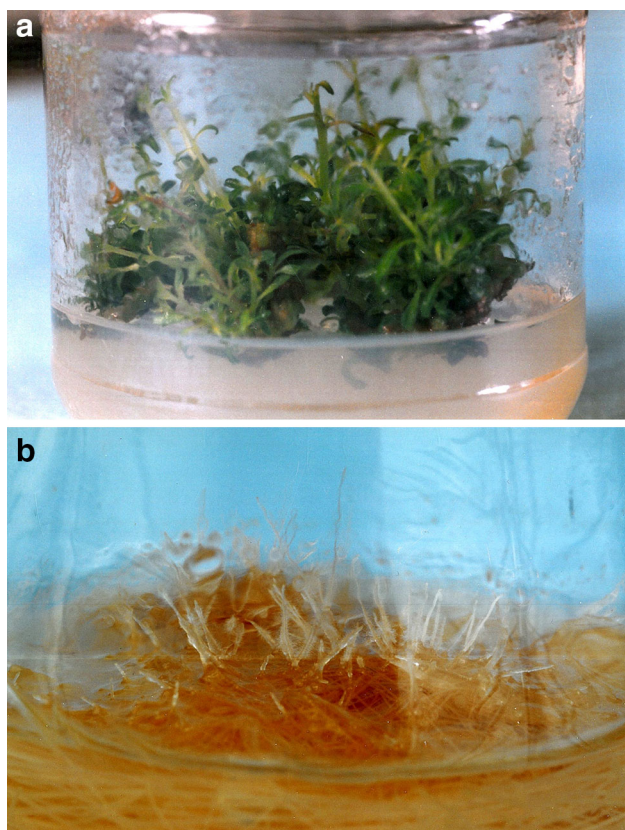
As depicted in Fig. 4a and Table 3, limonene and cedrol were the dominating constituents in aerial parts of the intact plants. Contrary to cedrol, which was previously found only in *Caryopteris tangutica* (Yang et al. 2005), limonene was frequently mentioned in previous studies on *Caryopteris* (Pu et al. 1984; Wen et al. 1990; Shatar and Adams 1999). As compared to the respective herbs, in vitro shoots accumulated lower amounts of cedrol and limonene (Fig. 4b; Table 3), whereas in vivo and adventitious roots were either devoid of or produced only traces of the above constituents (Fig. 5; Table 3). The exception to this rule was the roots of *Caryopteris × clandonensis* intact plant which accumulated significant amounts of cedrol (Fig. 5a).

GC–MS analyses showed that beside limonene and cedrol, volatile fractions of *Caryopteris* herbs contained substantial amounts of  $\alpha$ -pinene,  $\beta$ -pinene, myrtenal, *trans*-pinocarveol, carvone and caryophyllene oxide (Table 3). These compounds were previously identified in aerial parts of several *Caryopteris* plants (Pu et al. 1984; Shatar and Adams 1999; Wen et al. 1990; Yan and Wang 2009). However, the essential oil composition varied significantly,

**Fig. 1** Growth indices (Gi) of the investigated *Caryopteris* in vitro cultures. Values the mean  $\pm$  SD of three replicates.

**a** In vitro cultures grown on solidified SH medium supplemented with 8.88  $\mu\text{M}$  BAP and 1.0  $\mu\text{M}$  TDZ. **b** In vitro cultures grown in liquid SH medium supplemented with 4.92  $\mu\text{M}$  IBA

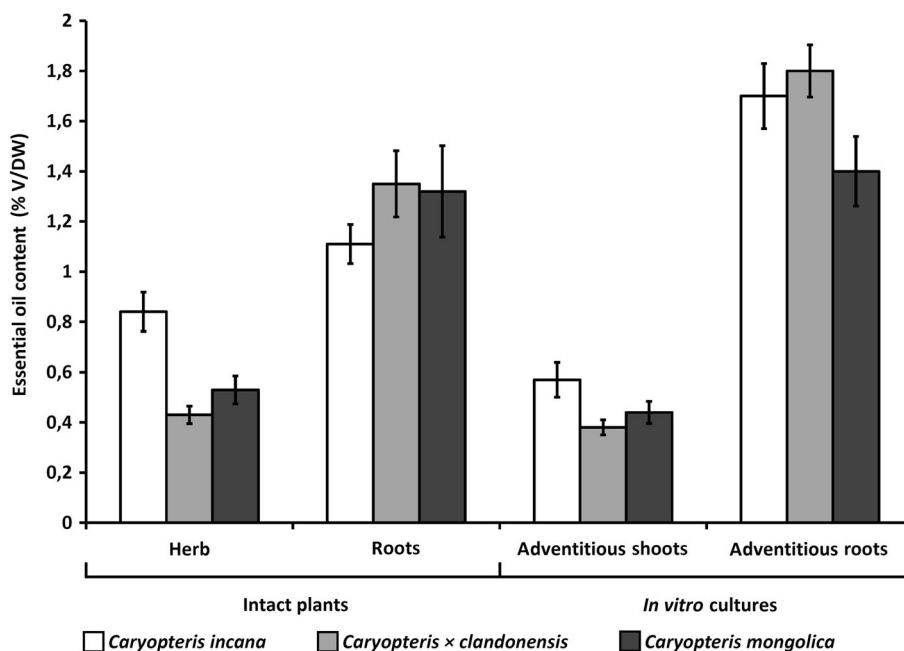




**Fig. 2** In vitro cultures of *Caryopteris* sp.: **a** shoots of *C. incana* and **b** adventitious roots of *C. mongolica*

depending not only on the species studied, but also on the plant habitat. For instance, the presence of  $\alpha$ -thujene and  $\beta$ -ocimene in *C. mongolica*, reported by Shatar and Adams

**Fig. 3** Percentage (V/DW) content of volatile oil in intact plants and in vitro cultures of various *Caryopteris* plants. Values the mean  $\pm$  SD of three replicates



(1999), was not confirmed by our research. The variability of essential oil composition was also observed in aerial parts of *C. incana*. Chu et al. (2011) reported estragole, linalool and 1,8-cineole as its major volatile constituents, whereas the study by Kim (2008) demonstrated the presence of 4,6,6-trimethyl-[1S-(1 $\alpha$ ,2 $\beta$ ,5 $\alpha$ )]-bicyclo[3.1.1]-hept-3-en-2-ol,  $\tau$ -cadinol, myrtenyl acetate, pinocarvone and  $\delta$ -3-carene in the investigated oil. According to Sun et al. (2004), *C. incana* oil contains linalool, perillalcohol and carvone. On the other hand, limonene prevailed in the essential oil investigated by Pu et al. (1984), which corresponds with our results (Fig. 4a). The essential oils examined by the above authors were collected from various locations in China and Korea. Apart from the local climatic and seasonal factors, harvesting time and storage conditions may influence the chemical composition of the *Caryopteris* essential oil, contributing to its great variability (Chu et al. 2011). Furthermore, it was suggested that stems, leaves and flowers of *Caryopteris* plants should be considered separately because of the different composition of volatiles (Yan and Wang 2009).

Given the described variation of *Caryopteris* volatile fraction, establishing in vitro cultures of these plants may be regarded as a step towards the standardization of the essential oil composition. As presented in Fig. 3, shoots grown in vitro had less volatile oil than herbs of the respective intact plants. However, the in vitro-derived essential oil was characterized by altered composition, manifested by the occurrence of new compounds (Table 3). For instance, 1,8-cineole, absent in intact plant materials, was the dominating (24.8–34.2 %) volatile compound of in vitro shoots (Fig. 4b). It was also surprising that the

**Table 3** The composition of hydrodistilled volatile oils obtained from intact plants and in vitro cultures of various *Caryopteris* plants

Compound	RI	Relative area (%) <sup>a</sup>											
		<i>Caryopteris incana</i>				<i>Caryopteris × clandonensis</i>				<i>Caryopteris mongolica</i>			
		In vivo		In vitro		In vivo		In vitro		In vivo		In vitro	
		Herb	Roots	Shoots	Roots	Herb	Roots	Shoots	Roots	Herb	Roots	Shoots	Roots
$\alpha$ -Pinene	925	<b>3.4</b>	tr	0.1	–	<b>4.3</b>	–	tr	–	<b>6.0</b>	tr	0.1	tr
Camphene	942	0.3	tr	tr	–	0.3	–	tr	–	0.3	–	tr	–
$\beta$ -Pinene	972	<b>4.3</b>	tr	0.5	0.2	<b>4.2</b>	–	tr	–	<b>3.5</b>	–	2.1	–
4-Octen-3-one	973	–	<b>2.6</b>	0.2	<b>9.8</b>	–	–	tr	–	–	<b>0.4</b>	tr	<b>3.8</b>
1-Octen-3-ol	978	0.6	–	<b>10.2</b>	<b>31.5</b>	0.5	<b>7.8</b>	<b>7.4</b>	<b>25.4</b>	0.6	<b>2.4</b>	<b>6.0</b>	<b>19.7</b>
Myrcene + 2-pentylfuran	985	0.2	–	tr	–	0.2	0.1	0.3	0.7	0.2	tr	0.2	0.7
3-Octanol	995	tr	–	0.3	tr	tr	tr	0.3	tr	tr	tr	0.1	0.1
1,3,8- <i>p</i> -Menthatriene	1000	0.1	–	0.1	–	0.3	–	0.2	–	0.1	–	0.2	–
<i>p</i> -Cymene	1019	0.3	–	0.3	tr	0.4	–	0.3	–	0.6	tr	0.5	0.2
Limonene	1025	<b>16.0</b>	tr	<b>7.0</b>	–	<b>14.7</b>	tr	<b>1.6</b>	tr	<b>11.9</b>	tr	<b>1.7</b>	tr
1,8-Cineole	1029	tr	–	<b>34.2</b>	–	tr	–	<b>24.8</b>	–	–	–	<b>28.6</b>	–
$\gamma$ -Terpinene	1054	tr	–	0.1	–	0.2	–	tr	–	0.2	–	0.3	–
Terpinolene	1081	tr	tr	tr	–	0.1	–	tr	–	0.1	tr	0.1	tr
<i>p</i> -Cymenene	1087	0.3	–	0.6	–	0.3	tr	0.7	0.2	0.2	tr	0.4	tr
Linalool	1098	tr	tr	0.1	<b>7.6</b>	tr	tr	0.1	<b>0.9</b>	0.1	tr	tr	<b>1.4</b>
$\alpha$ -Campholenal	1124	0.3	–	tr	–	0.4	–	tr	–	0.5	–	0.2	–
<i>trans-p</i> -2,8-Menthadien-1-ol	1134	<b>2.3</b>	–	<b>2.9</b>	–	<b>1.2</b>	–	<b>2.5</b>	–	<b>1.5</b>	–	<b>0.6</b>	–
<i>trans</i> -Pinocarveol	1137	<b>3.3</b>	–	<b>0.8</b>	–	<b>4.0</b>	–	<b>1.1</b>	–	<b>4.8</b>	–	<b>6.7</b>	–
Camphor + <i>trans</i> -verbenol	1144	0.3	–	0.1	–	0.5	–	0.6	–	0.8	0.1	–	0.6
Pinocarvone	1159	<b>2.0</b>	–	<b>0.8</b>	–	<b>2.7</b>	–	<b>0.7</b>	–	<b>3.1</b>	–	<b>6.2</b>	–
( <i>Z</i> )-Cinnamaldehyde	1168	tr	–	–	–	tr	–	–	–	tr	–	–	–
4-Terpineol	1178	0.4	–	0.1	–	0.6	–	0.1	–	0.7	–	0.6	–
Methylphenyl-ethanone	1182	0.3	–	0.2	–	0.4	–	0.2	–	0.6	–	–	–
Myrtenal + myrtenol	1193	<b>4.8</b>	–	<b>2.3</b>	–	<b>4.7</b>	–	<b>1.1</b>	–	<b>4.9</b>	–	<b>10.1</b>	–
4,7-Dimethylbenzopyran	1211	tr	–	–	–	tr	–	–	–	tr	–	–	–
<i>trans</i> -Carveol	1217	<b>3.5</b>	–	1.3	–	<b>2.5</b>	–	0.9	–	<b>2.7</b>	–	0.4	–
Verbenon	1224	0.1	–	1.3	–	0.1	–	0.8	–	0.1	–	tr	–
Carvone	1242	<b>3.7</b>	–	1.0	–	<b>3.8</b>	–	0.6	–	<b>3.5</b>	–	0.2	–
Perillaldehyde	1274	0.3	–	0.2	tr	0.4	–	tr	–	0.4	–	tr	0.5
Bornyl acetate	1282	0.1	–	tr	–	0.2	tr	0.1	tr	0.1	–	0.1	–
Mentha-1,3-dien-7-al	1285	–	–	–	–	–	–	–	–	–	–	–	–
$\alpha$ -Copaene	1373	0.4	<b>2.3</b>	0.4	tr	0.3	<b>5.4</b>	0.7	0.5	0.2	<b>0.5</b>	0.7	0.4
$\beta$ -Bourbonene	1389	0.2	tr	tr	0.3	0.1	–	–	–	0.2	–	–	–
$\alpha$ -Cedrene	1413	tr	tr	tr	–	0.2	0.5	0.1	0.4	0.2	0.1	0.2	0.2
$\beta$ -Caryophyllene	1417	<b>2.3</b>	tr	0.6	tr	<b>2.2</b>	tr	1.2	tr	<b>1.5</b>	–	1.4	–
$\beta$ -Cedrene	1422	0.2	–	0.1	–	1.2	–	0.9	–	0.9	–	0.7	–
$\alpha$ -Humulene	1453	0.7	–	tr	–	0.6	–	0.3	–	0.4	–	0.3	–
Germacrene D	1478	0.5	–	1.2	–	0.5	–	tr	–	0.2	–	0.5	–
4-Epi-cubebol	1493	0.1	–	0.3	–	0.1	tr	0.2	tr	0.1	–	0.1	–
1-(2,3,6-Trimethylphenyl)-3-buten-2-one	1501	tr	–	–	–	tr	–	–	–	tr	–	–	–
3,5-Bis(1,1-dimethyl)-phenol	1504	tr	<b>26.2</b>	0.1	<b>6.0</b>	tr	<b>15.7</b>	0.2	<b>3.5</b>	tr	<b>12.9</b>	0.5	<b>4.1</b>
$\beta$ -Bisabolene	1506	0.2	–	0.2	–	0.1	–	–	–	0.2	–	–	–
Cubebol	1513	tr	–	tr	–	tr	–	0.2	–	tr	tr	tr	tr

**Table 3** continued

Compound	RI	Relative area (%) <sup>a</sup>											
		<i>Caryopteris incana</i>				<i>Caryopteris × clandonensis</i>				<i>Caryopteris mongolica</i>			
		In vivo		In vitro		In vivo		In vitro		In vivo		In vitro	
		Herb	Roots	Shoots	Roots	Herb	Roots	Shoots	Roots	Herb	Roots	Shoots	Roots
$\sigma$ -Cadinene	1516	0.4	–	0.3	–	0.4	1.5	1.1	0.1	0.2	tr	0.6	0.2
$\alpha$ -Calacorene	1539	0.1	–	tr	–	0.1	1.3	0.3	0.1	0.1	tr	0.2	0.2
Caryophyllene- $\beta$ -oxide	1549	0.2	–	–	–	0.2	–	tr	–	0.2	–	tr	–
$\gamma$ -Calacorene	1560	tr	–	tr	–	tr	tr	0.2	tr	tr	–	0.1	–
Caryolan-1-ol	1574	–	–	–	–	–	–	–	–	–	–	–	–
Caryophyllene oxide	1580	<b>4.8</b>	–	0.5	–	<b>4.8</b>	0.7	1.2	0.2	<b>5.4</b>	–	1.5	–
Lemnalol	1586	0.4	–	–	–	0.4	–	1.6	–	0.3	–	0.6	–
Cedrol	1607	<b>10.9</b>	–	<b>1.4</b>	–	<b>10.9</b>	<b>8.4</b>	<b>6.1</b>	0.9	<b>10.7</b>	–	<b>6.4</b>	–
$\tau$ -Muurolol	1642	tr	–	0.2	–	tr	tr	0.2	0.5	tr	0.2	0.4	0.8
6,10,14-Trimethyl-2-pentadecanone; =hexahydrofarnesyl acetone	1840	–	–	0.6	–	tr	tr	1.0	0.7	0.1	–	0.6	–
Isophytol	1944	–	–	0.1	–	–	–	0.1	tr	–	–	tr	–
Hexadecanoic acid	1961	–	–	<b>0.8</b>	–	–	2.1	<b>1.3</b>	3.8	–	2.4	<b>8.0</b>	7.1
Geranyllinalool	2020	–	–	0.1	–	–	–	0.2	–	–	–	0.2	–
Abietatriene	2052	tr	<b>13.5</b>	<b>16.0</b>	<b>5.0</b>	tr	<b>12.7</b>	<b>10.9</b>	<b>21.6</b>	tr	<b>4.5</b>	<b>5.0</b>	<b>7.3</b>
Heneicosane	2100	tr	<b>14.1</b>	0.5	<b>5.3</b>	tr	<b>8.4</b>	0.3	<b>2.5</b>	tr	<b>10.5</b>	–	<b>3.6</b>
Phytol	2105	–	–	<b>4.2</b>	–	tr	–	<b>8.6</b>	1.2	tr	–	<b>4.8</b>	–
Docosane	2200	–	0.1	tr	0.9	tr	0.2	tr	0.7	–	0.9	–	1.5
Tricosane	2300	–	<b>12.4</b>	0.2	<b>14.0</b>	tr	<b>5.1</b>	0.1	<b>6.6</b>	tr	<b>7.5</b>	0.3	<b>15.0</b>
<i>trans</i> -Totarol	2318	–	<b>2.5</b>	<b>7.0</b>	<b>3.9</b>	tr	<b>5.7</b>	<b>4.5</b>	<b>29.2</b>	tr	<b>6.9</b>	<b>4.3</b>	<b>13.5</b>
Tetracosane	2400	–	tr	tr	tr	–	tr	tr	0.3	–	tr	tr	0.6
Pentacosane	2500	–	<b>3.1</b>	0.1	<b>6.7</b>	tr	<b>3.1</b>	0.1	<b>6.7</b>	tr	<b>2.2</b>	0.2	<b>7.3</b>

<sup>a</sup> GC–MS data, major constituents presented in bold text

– Compound not detected, *tr* traces

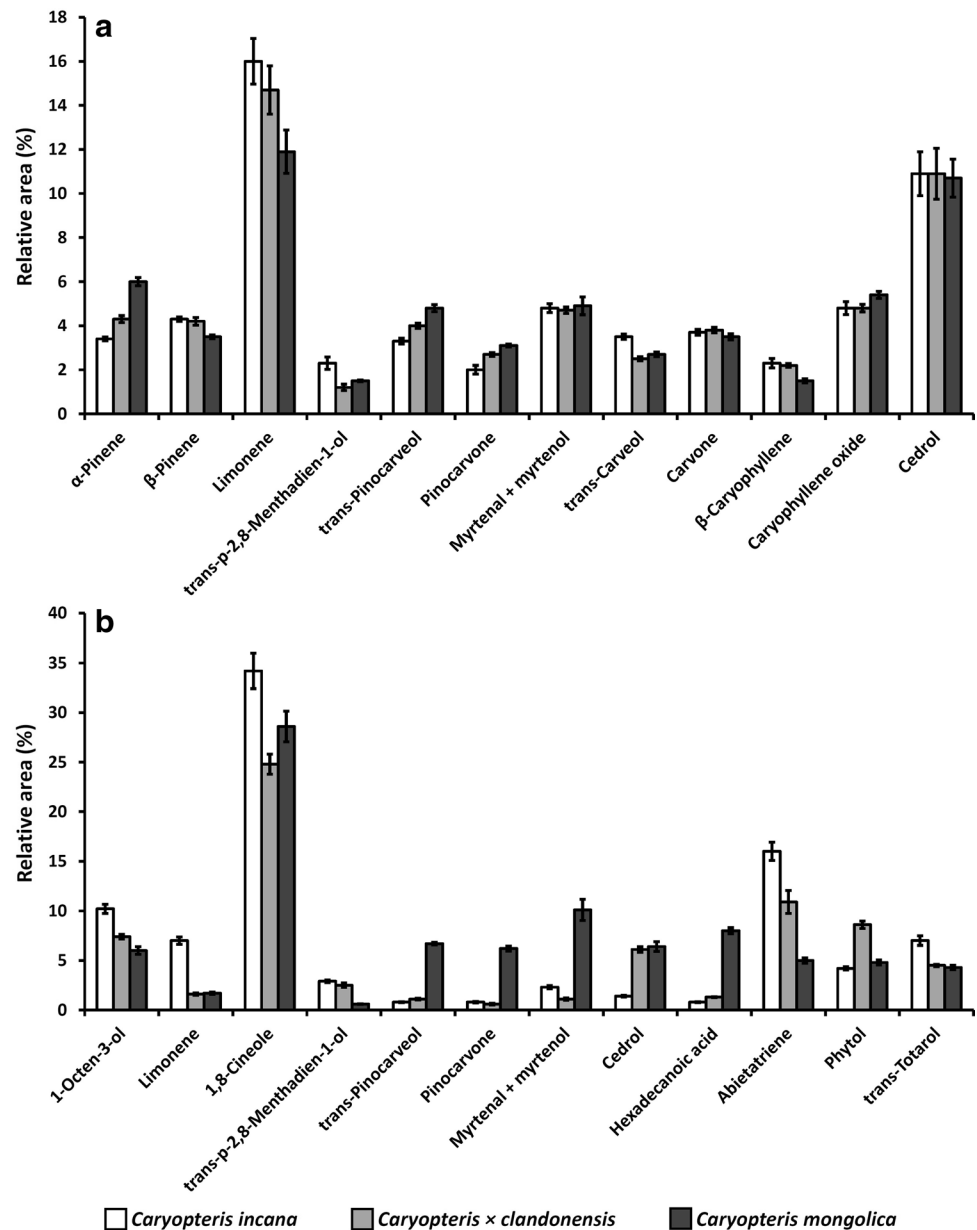
essential oil obtained from in vitro shoot cultures contained abietatriene, *trans*-totarol and 1-octene-3-ol (Table 3), which were absent (or present in trace amounts) in the herbs, but characteristic for in vivo roots of the investigated plants.

As compared to the aerial parts, in vivo roots of the respective *Caryopteris* plants were characterized with higher essential oil content (Fig. 3) which also had a much simpler composition (on average, 19 compounds were identified—Table 3). There were also significant, qualitative differences in the composition of volatile fraction present in the roots, as compared to the herbs (Figs. 4a, 5a; Table 3). In all analyzed species, the dominating constituent of the root oil was 3,5-bis(1,1-dimethyl)-phenol (12.9–26.2 %). Other major compounds included abietatriene (4.5–13.5 %), heneicosane (8.4–14.1 %) and tricosane (5.1–12.4 %) (Fig. 5a; Table 3), which accumulated only in trace amounts in aerial parts of the examined species. To the best of our knowledge, this is the first report

concerning the composition of root-derived *Caryopteris* volatile oil.

Adventitious roots synthesized large quantities of volatile oils with a composition similar to those accumulated in roots of the intact plants. However, linalool appeared as a new constituent (0.9–7.6 %), whereas the production of 3,5-bis(1,1-dimethyl)phenol, the main component of the essential oil from in vivo roots, was slowed down (3.5–6.0 %) (Fig. 5b; Table 3). Regardless of the species, the most abundant element of the adventitious root-derived oil was 1-octen-3-ol (19.7–31.5 %), present only in small quantities in the roots of soil-grown plants except for in vivo roots of *Caryopteris × clandonensis*, whose oil contained nearly 8 % of the above compound. In addition, adventitious roots accumulated significant amounts of abietane diterpenoids; especially, adventitious root cultures of *Caryopteris × clandonensis* were shown to be a rich source of abietatriene and *trans*-totarol (21.6 and 29.2 %, respectively) (Fig. 5b).

**Fig. 4** Percentage content of major constituents of hydrodistilled essential oils obtained from herbs (a) and in vitro shoots (b) of various *Caryopteris* plants. Values the mean  $\pm$  SD of three replicates

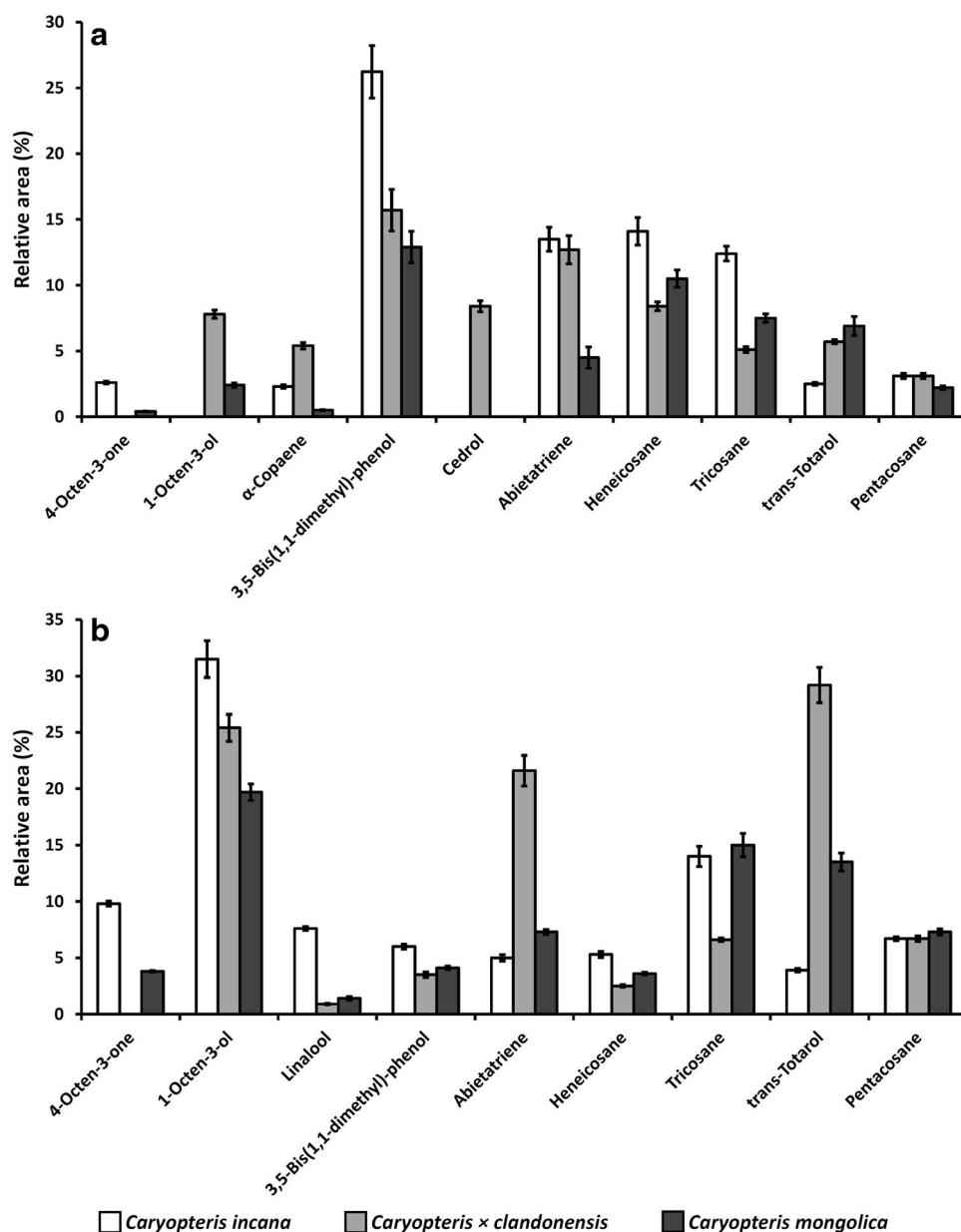


The conducted experiments demonstrated that in vitro cultures of *Caryopteris* accumulated high levels of medicinally relevant compounds absent in the intact plants, such as 1,8-cineole, abietatriene and totarol. The first of these constituents, abundant in in vitro shoots of the investigated plants, is a potent anti-bacterial agent (Bosnić et al. 2006) and acetylcholinesterase inhibitor (Aazza et al. 2011). The major volatile constituent of adventitious roots (1-octen-3-ol) found practical application as an ingredient of mosquito-repellent devices (Burfield and Reekie 2005) and as a flavoring agent in food industry (Maggi et al. 2009). However, perhaps the most valuable constituents of *Caryopteris* de novo roots are abietane-type diterpenoids, previously shown to exhibit significant cytotoxic activity against human leukemia HL<sub>60</sub> cells (Gao and Han 1997). Moreover,

abietatriene was reported to display a strong gastroprotective effect, comparable to lansoprazole (Areche 2007), whereas totarol demonstrated antifungal, antimalarial and antifibrotic activities (Lee et al. 2008; Tacon et al. 2012). The latter is also a well-known antimicrobial agent against Gram-positive bacteria, used in cosmetics such as toothpastes, mouthwashes and acne preparations (Varvaresou et al. 2009). Given the high essential oil yield of *Caryopteris* adventitious roots, as well as the suitability of this type of biomass for industrial production of plant secondary metabolites (Murthy et al. 2008; Baque et al. 2012), the established root cultures can be considered a promising, alternative source of *Caryopteris* oil. Thus, further scale-up experiments in bioreactors are required to evaluate essential oil productivity of the investigated cultures.



**Fig. 5** Percentage content of major constituents of hydrodistilled essential oils obtained from in vivo (a) and adventitious roots (b) of various *Caryopteris* plants. Values the mean  $\pm$  SD of 3 replicates



**Author contribution statement** M. Luczkiewicz, A. Jesionek and A. Kokotkiewicz conducted the in vitro research, analyzed the data and wrote the manuscript. P. Migas isolated the volatile fractions from the *Caryopteris* plant material. M. Mardarowicz, A. Szreniawa-Sztajnert and B. Zabiegala carried out the analysis of essential oils with GC–MS. A. Bucinski checked and corrected the manuscript. All the authors read and approved the manuscript in its final form.

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**Conflict of interest** The authors declare that they have no conflict of interest.

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