



Biotrickling filtration of *n*-butanol vapors: process monitoring using electronic nose and artificial neural network

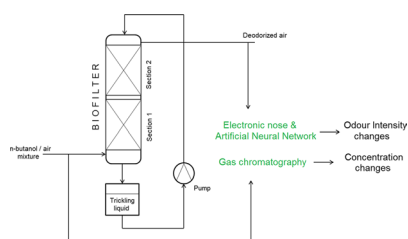
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Abstract

Biotrickling filtration is one of the techniques used to reduce odorants in the air. It is based on the aerobic degradation of pollutants by microorganisms located in the filter bed. The research presents the possibility of using the electronic nose prototype combined with artificial neural network for biofiltration process monitoring in terms of reduction in *n*-butanol concentration and odour intensity of treated air. The study was conducted using two-section biotrickling filter packed with a commercially available mixture of peat and perlite during 42 days with different *n*-butanol inlet concentrations, i.e., 100, 200, 400, and 800 ppm. During the tests, a concentration and odour intensity removal efficiency of around 90% and 20% was obtained, respectively. It has been shown that the highest values were obtained for an inlet *n*-butanol concentration of 200–400 ppm.

Graphic abstract



Keywords Neural network · Sensors · Odoriferous substances · Alcohols

Introduction

Removal of odorous compounds from air, including nitrogen and sulfur compounds as well as volatile organic compounds, is gaining importance due to social awareness and increasingly stringent environmental legislations [1, 2]. Such a trend enforces the development of sustainable and efficient methods of air deodorization. Among them, biological methods of gas treatment are considered to be more effective and economical alternative to conventional methods such as

chemical scrubbing, incineration, absorption, or condensation [3, 4]. This is especially true when large volumes of gases containing low or medium concentrations of pollutants are taken into account.

One of the most common techniques of biological gas treatment is biofiltration. The process of biofiltration consists in passing a polluted stream of gas through a filter bed, inhabited by microorganisms of various species (typically bacteria or fungi) [5]. The pollutants diffuse from the gas phase to the so-called biofilm, formed on the surface of the elements of the filter packing. The compounds adsorbed on the surface or absorbed in the biofilm undergo biodegradation and the air leaves the biofilter cleansed and free of unpleasant odors. The biofiltration process can be carried out in two main types of apparatus, i.e., in a conventional biofilter or in a biotrickling filter. In a conventional biofilter, the contaminated gas is initially humidified in a separate

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chamber, and only then, it is transferred to the biofilter. The packing of such a biofilter is most often made of natural organic materials, e.g., bark, cones, or peat. The construction of a biotrickling filter, on the other hand, enables the process to be carried out in one apparatus. The filter bed, made of natural or synthetic materials, is regularly trickled with liquid enriched with minerals necessary for the growth of microorganisms. The trickling liquid may circulate in a closed system subject to periodic regeneration. Biotrickling filters allow easier control and regulation of the process, including increased process stability, pH and temperature control as well as continuous nutrient supply, compared to conventional biofilters [6–8].

Biotrickling filtration is currently considered as the most promising technology of biological air treatment [9]. This results, among others, from the application of mobile liquid phase, providing means for washing out from the bed the biodegradation intermediates, possibly inhibiting the microbial activity in conventional biofilters as well as outstanding process efficiency and economy, as compared to other types of bioreactors [10–12].

In this paper, a counter-current biotrickling filtration of variable inlet loads of *n*-butanol was investigated. Butanol is a representative of hydrophilic volatile organic compound. However, it is treated in biological systems with rather high purification efficiency, and its removal from air in biotrickling filters has been scarcely investigated [13]. In addition, in this study, a biotrickling filter was packed with a mixture of inert and organic materials, i.e., a mixture of perlite and peat, contrary to majority of experimental studies, where biotrickling filters are packed with inert materials (ceramic elements, polyurethane foam, or lava rock) [14]. Such an approach allows a rapid start-up of the process and

eliminates the step of inoculating the packing materials with microbial species.

The most frequently used techniques to evaluate effectiveness of biofiltration process are gas chromatography techniques. They enable separation and determination of concentrations of individual components of the mixture. In terms of the evaluation of the odour quality of purification, such information is not directly useful. Therefore, electronic noses—devices enabling holistic analysis of gas samples—are increasingly used to assess the effectiveness of biofiltration [15, 16]. The electronic nose system consists of four main components, as presented in Fig. 1.

As pattern recognition system various chemometric algorithms are used, e.g., principal component analysis (PCA), linear discriminant analysis (LDA), support vector machine (SVM), or partial least square (PLS). However, the most valuable method used in the e-nose system in artificial neural network (ANN). Artificial neural networks are now considered the best method of analyzing data from artificial senses, mainly due to the fact that ANNs in their architecture and functioning resemble the nervous system in humans. The simplest, having only one neuron, ANN is called the perceptron. The McCulloch–Pitts neuron, which is a simplified model of the biological nerve cell. The similarity in the construction of both neurons is presented in Fig. 2.

The use of ANN for data analysis is possible only after prior collection of the training data set—examples of inputs along with defined, corresponding output values. Neural network learning process involves changing its internal parameters (weight coefficients and neuron activation thresholds). This is done using the appropriate algorithm, usually learning under supervision. The most frequently used algorithm for this purpose is the back error propagation algorithm

Fig. 1 Electronic nose system description

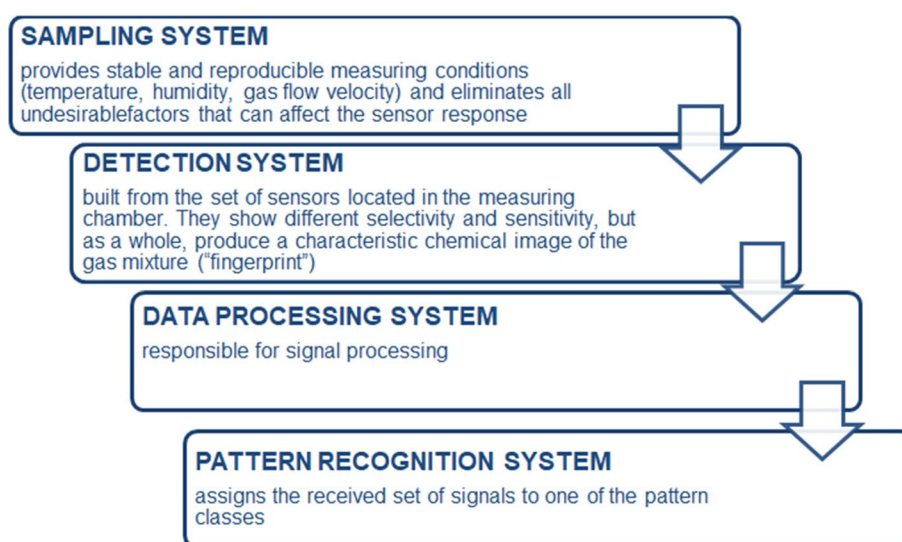


Fig. 2 Similarity of neuron cell and artificial neural network construction

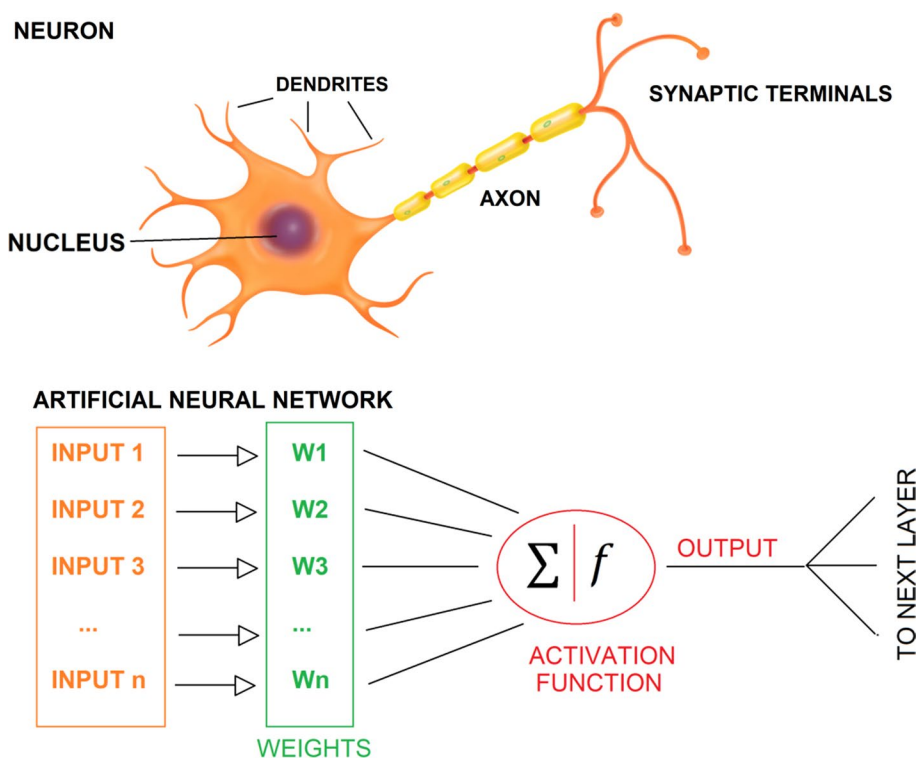
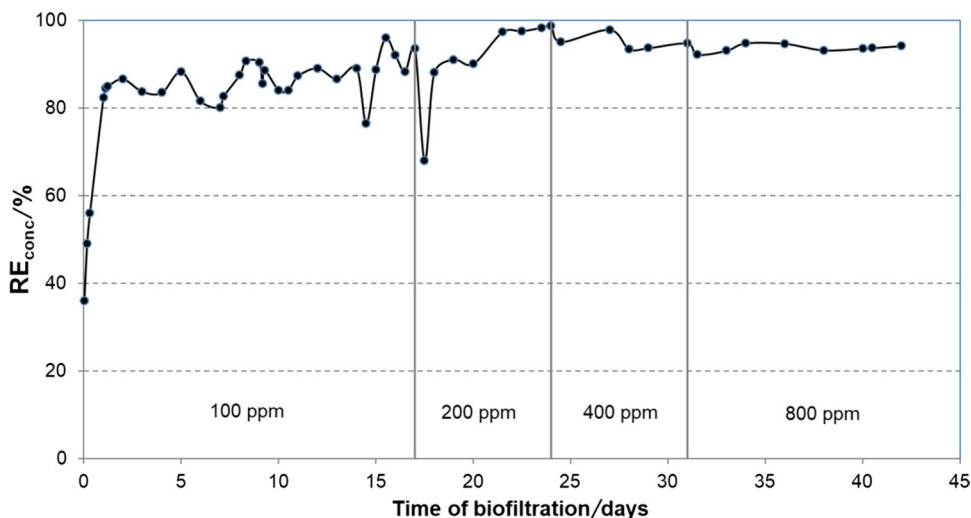


Fig. 3 Concentration removal efficiency (RE_{conc}) changes during the process



[17]. Its operation consists in the modification of weights and threshold values based on training data in such a way as to minimize the error made by the network while performing its assigned tasks for all data included in the training set.

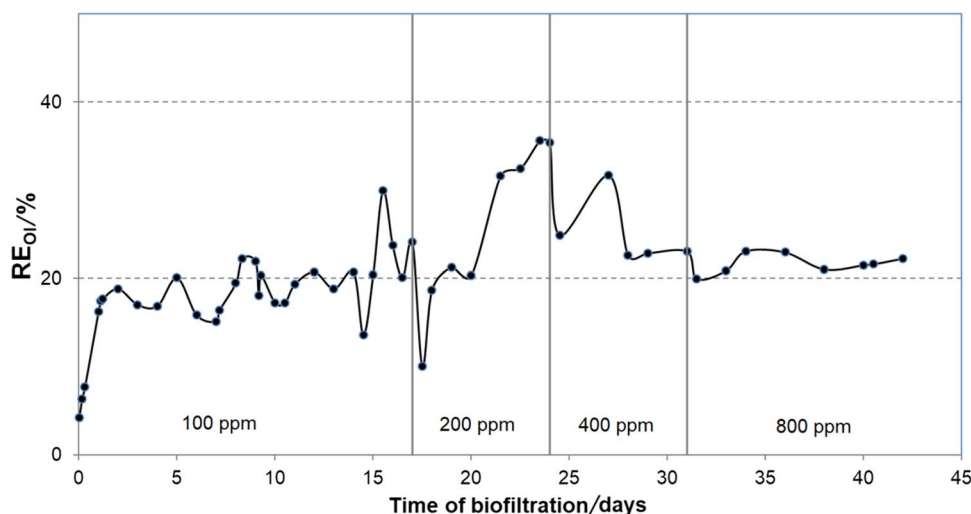
The article presents estimation of the odour intensity of air samples (contaminated with *n*-butanol) undergoing biofiltration process using electronic nose prototype and artificial neural network. The removal efficiency calculated using odour intensity changes was compared with values calculated using changes in *n*-butanol concentration.

Results and discussion

The performance of a peat-perlite-packed biotrickling filter is shown in Fig. 3 (concentration changes) and Fig. 4 (odour intensity changes).

Shortly after introducing the mixture of air and *n*-butanol to the biofilter, concentration removal efficiency of about 37% is obtained and it increases to about 80–85% during the first day of the process. When continuing the process, values of RE_{conc} slightly increase, reaching the values close to 90% within first 15 days of biofiltration.

Fig. 4 Odour intensity removal efficiency (RE_{OI}) changes during the process



In terms of odour intensity for 0–15 and 31–42 days, removal efficiency was close to 20%. The higher values were noticed for inlet concentrations equal to 200 and 400 ppm. Such high values of RE result from two main reasons: first, *n*-butanol is a hydrophilic volatile organic compound and it is easily removed from air streams in biological systems [15, 18]; second, the applied packing of a biotrickling filter is rich in plenty of microbial species which favors biodegradation of air pollutants [19, 20]. In addition, stable values of *n*-butanol removal efficiency indicate that the biological system attains adaptation to the treated gas and microorganisms present in the peat structure consume *n*-butanol as a source of carbon. Thus, the period of 15 days was assumed as a start-up period, i.e., the time period ensuring steady-state conditions in the system [3].

To investigate the influence of an inlet loading on the efficiency of biofiltration, inlet concentrations of *n*-butanol were increased at 17th, 24th, and 31st days of the process duration. This is reflected by a drop in the RE values corresponding to day 17th of biofiltration. However, within next 2 days, the concentration removal efficiency increases, reaching the values in the range 95–99%. Interestingly, further increases in the inlet loading of *n*-butanol (days 24 and 31) only slightly influence the values of RE_{conc} . Such behavior of the system suggests a good adaptation of the microbial species inhabiting the biofilter packing for the treated compound and suggests a stable working conditions for long-term experiments. Obtained results are in accordance with literature data regarding biotrickling filtration of *n*-butanol [21]. It is worth noting that no problems with bed clogging were faced during the investigated period of biofiltration for the set values of EBRT and trickling liquid velocity and frequency (Table 1). The measured pressure drop across the filter packing during the experiment was in the range of about 60–90 mm H_2O .

Table 1 Basic process parameters of a biotrickling filter

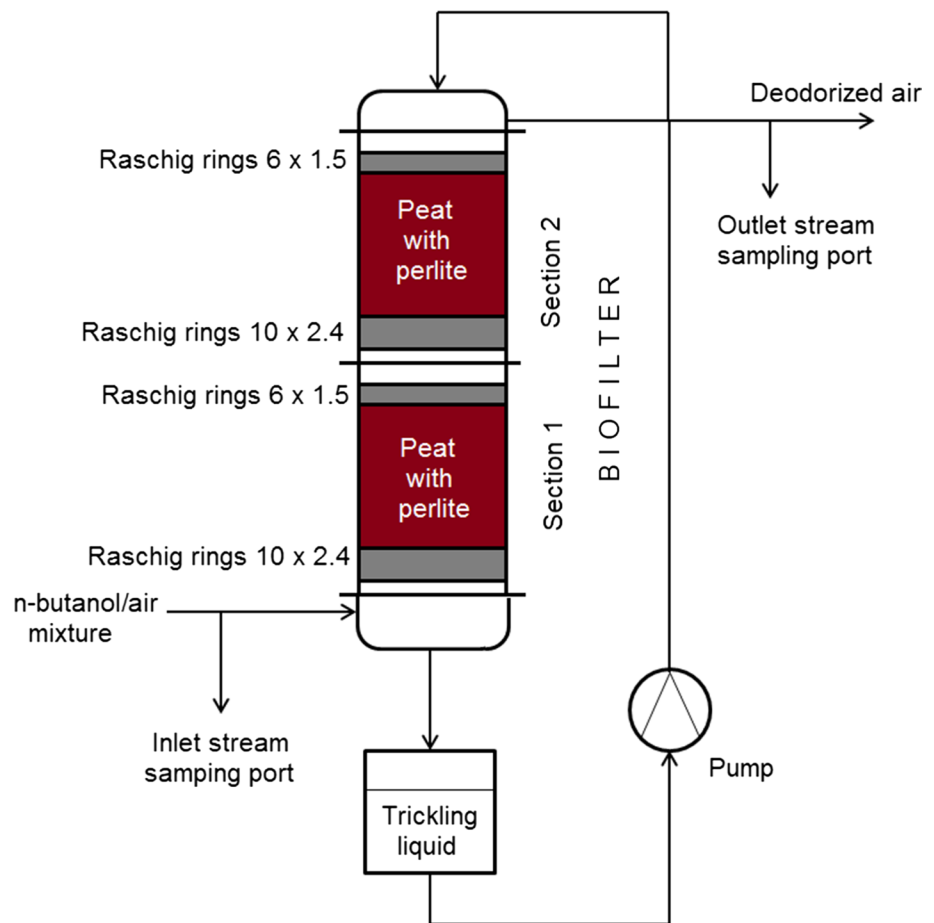
Parameter	Value
Internal diameter of a biofilter	8 cm
Total height of a two-section biofilter	68 cm
Total volume of biofilter packing	2.5 dm ³
Volumetric flow rate of gas phase	2.5 dm ³ min ⁻¹
Empty bed residence time (EBRT)	60 s
Inlet <i>n</i> -butanol concentrations	100, 200, 400, 800 ppm
Height of a Raschig rings layer	
10×2.4 mm	3 cm
6×1.5 mm	2 cm
Volumetric flow rate of trickling liquid	0.2 dm ³ min ⁻¹
Frequency of trickling	1 min 2 h ⁻¹

Conclusion

As a result of the studies, it was found that the electronic nose prototype along with the proposed artificial neural network can be successfully used to estimate the odour intensity of *n*-butanol contaminated air samples undergoing biofiltration process. Presented results indicate that *n*-butanol vapors may be effectively eliminated from air in a biotrickling filter packed with a mixture of peat and perlite. Concentration removal efficiency of *n*-butanol exceeding 90% has been achieved after the period of 20 days of biofiltration. Such RE_{conc} values correspond 20–30% in odour intensity removal. On 23th day of process duration the highest efficiency was observed ($RE_{conc} = 98.6\%$, $RE_{OI} = 35.4\%$). High values of RE were observed for inlet concentrations equal to 200 and 400 ppm. This allows to conclude that for this biofilter the values of these concentrations are optimal.

The choice of *n*-butanol as the target compound is also justified by previous research, dealing with conventional

Fig. 5 Schematic diagram of the biotrickling filter experimental setup



biofiltration of air polluted with *n*-butanol vapors [16]. Thus, the experimental comparison between the process efficiency (conventional biofiltration versus biotrickling filtration) is interesting both from scientific as well as practical points of view. These results indicate that *n*-butanol biofiltration precedes with much higher efficiency in biotrickling filters (95–99%), keeping high removal efficiency even for increased inlet loads, as compared to conventional biofiltration (80–90%).

Experimental

The research was carried out in a two-section biotrickling filter, shown schematically in Fig. 5. The biofilter was made of organic glass and packed with a commercially available mixture of peat and perlite (COMPO SANA, Compo, Germany). Dimensions and process parameters of a biofilter are given in Table 1.

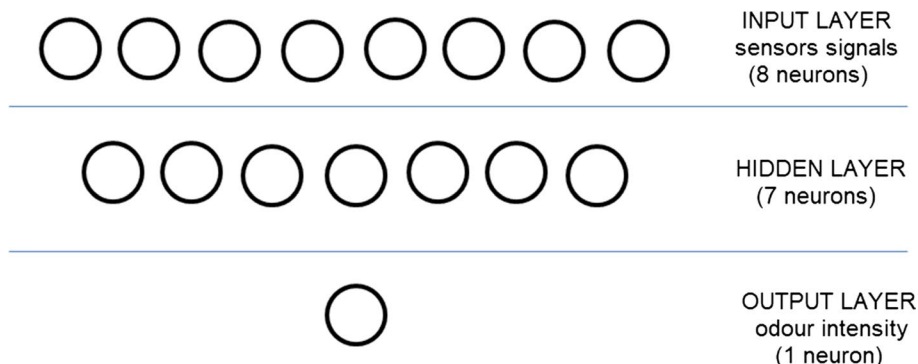
After placing a filter packing bed in the individual sections and having the bioreactor twisted, a flow of a clean air as well as a trickling of a liquid were introduced through a packed bed. The intensity of air and trickling liquid flows

are shown in Table 1. The activation of the biofilter packing lasted 4 days, and as a trickling liquid a buffered peptone water medium was used. After activation of the bed, the flow of *n*-butanol and air mixture was started, further using a trickling liquid as described above. After 4 days of operation of the system, the liquid used was replaced with a fresh medium. The composition of the medium was changed and it is as follows: $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ (15.2 g dm^{-3}), KH_2PO_4 (3 g dm^{-3}), NaCl (0.5 g dm^{-3}), and NH_4Cl (1 g dm^{-3}). A trickling liquid of such a composition was changed every 4 days. During the tests, at least once a day, gas samples were taken from the inlet and outlet streams. The flow of the gas mixture stream at the inlet to the biofilter and the pressure drop on the filter bed were regularly controlled. The temperature of the filter bed was in the range 23–25 °C.

Odour intensity of the air samples was determined using an electronic nose and artificial neural network. Electronic nose analyses were carried out using an e-nose prototype equipped with eight chemical gas sensors: ION Science MiniPID, Figaro FECS44-100, Figaro FECS50-100, Figaro TGS2600, Figaro TGS2602, Figaro TGS2603, Figaro TGS823, Figaro TGS8100. The sensor characteristics are presented in Table 2. The collected samples were sucked

Table 2 Electronic nose sensors characteristics

Sensor model	Sensor type	Detected compounds
ION Science MiniPID	Photoionisation	Aromatic hydrocarbons, isobutylene
Figaro FECS44-100	Electrochemical	Ammonia, amines
Figaro FECS50-100	Electrochemical	Hydrogen sulfide, mercaptanes
Figaro TGS2600	Metal oxide semiconductor	Ethanol, isobutane
Figaro TGS2602	Metal oxide semiconductor	Hydrogen sulfide, toluene, ammonia
Figaro TGS2603	Metal oxide semiconductor	Trimethylamine, methyl mercaptane, ethanol
Figaro TGS823	Metal oxide semiconductor	Benzene, <i>n</i> -hexane, acetone
Figaro TGS8100	Metal oxide semiconductor	Methane, isobutene, ethanol

Fig. 6 Architecture of the artificial neural network**Table 3** VDI 3940 Odour intensity scale

Intensity level	Odour strength
0	No odour
1	Very weak
2	Weak
3	Distinct
4	Strong
5	Very strong
6	Extremely strong

by a membrane pump into the e-nose chamber for 15 s. The sample was then kept in the chamber for 30 s. The purified air was then directed into the chamber for regeneration of the sensors. For data analysis, the maximum signal value of each sensor was used.

The odour intensity of the sample was determined using previously designed Artificial Neural Network (topology: 8-7-1). Architecture of the network is presented in Fig. 6.

Three layer neural network was designed. The ANN learning process was based on the analysis of *n*-butanol samples of known odour intensity using electronic nose and sensory analysis. It was carried out by four persons, selected according to the procedure described in [22]. Each member of the panel was responsible for assigning the appropriate odour intensity value to a given sample using a seven-step scale described in German Standard VDI 3940 (Table 3).

The weights were modified until the error between the measured and predicted values are minimized. RStudio Desktop (v. 1.0.143) software was used as the computational software.

Concentrations of *n*-butanol in the gas samples at the inlet and outlet of the biofilter were determined using a VARIAN CP-3800 gas chromatograph with a flame ionization detector (FID). A 30 m long HP-5MS capillary column was used (inner diameter 0.25 mm, film thickness 0.25 μm). Conditions of chromatographic analysis were as follows: carrier gas: nitrogen; carrier gas flow: 2 $\text{cm}^3 \text{min}^{-1}$; split: 3; oven temperature: 150 $^\circ\text{C}$; injector temperature: 150 $^\circ\text{C}$; FID temperature: 200 $^\circ\text{C}$; total time of a single analysis: 5 min. The analyses were performed in triplicate and the results are presented for mean values.

Removal efficiency (RE) was calculated using change in *n*-butanol concentration and odour intensity between inlet and outlet stream using Eqs. (1) and (2), respectively:

$$\text{RE}_{\text{conc}} = \frac{C_{\text{inlet}} - C_{\text{outlet}}}{C_{\text{inlet}}} \times 100\%, \quad (1)$$

$$\text{RE}_{\text{OI}} = \frac{\text{OI}_{\text{inlet}} - \text{OI}_{\text{outlet}}}{\text{OI}_{\text{inlet}}} \times 100\%. \quad (2)$$

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