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## Valorization of waste cabbage leaves by postharvest photochemical treatments monitored with a non-destructive fluorescence-based sensor

--Manuscript Draft--

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<b>Abstract:</b>	<p>The biosynthesis of polyphenolic compounds in cabbage waste, outer green leaves of white head cabbage (<i>Brassica oleracea</i> L. var. <i>capitata</i> subvar. <i>alba</i>), was stimulated by postharvest irradiation with UVB lamps or sunlight. Both treatments boosted the content of kaempferol and quercetin glycosides, especially in the basal leaf zone, as determined by the HPLC analysis of leaf extracts and by a non-destructive optical sensor.</p> <p>The destructive analysis of samples irradiated by the sun for 6 days at the end of October 2015 in Skierniewice (Poland) showed an increase of leaf flavonols by 82% with respect to controls. The treatment by a broadband UVB fluorescent lamp, with irradiance of 0.38 W m<sup>-2</sup> in the 290-315 nm range (and 0.59 W m<sup>-2</sup> in the UVA region) for 12 hours per day at 17 °C along with a white light of about 20 μmol m<sup>-2</sup> s<sup>-1</sup>, produced a flavonols increase of 58% with respect to controls.</p> <p>The kinetics of flavonols accumulation in response to the photochemical treatments was monitored with the FLAV non-destructive index. The initial FLAV rate under the sun was proportional to the daily radiation doses with a better correlation for the sun global irradiance (R<sup>2</sup>=0.973), followed by the UVA (R<sup>2</sup>=0.965) and UVB (R<sup>2</sup>=0.899) irradiance.</p> <p>The sunlight turned out to be more efficient than the UVB lamp in increasing the flavonols level of waste leaves, because of a significant role played by UVA and visible solar radiation in the regulation of the flavonoid accumulation in cabbage. The FLAV index increase induced on the adaxial leaf side was accompanied by a lower but still significant FLAV increase on the unirradiated abaxial side, likely due to a systemic signaling by mean of the long-distance movement of macromolecules.</p> <p>Our present investigation provides useful data for the optimization of postharvest photochemical protocols of cabbage waste valorization. It can represent a novel and alternative tool of vegetable waste management for the recovery of beneficial phytochemicals.</p>
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She has decades of experience with the effects of elicitor treatments, including UV radiation, on crop plants, and is one of the leading experts in this topic.  
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Among other activities, he studies the plant responses to the light (mainly UVB, but also UVA and PAR), in particular the synthesis of phenolic compounds both under natural conditions and under artificial irradiation. He is a member of the UV4Plant association which is focused on the interaction between UV radiation and plants.  
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**Response to Reviewers:**

The manuscript entitled “Valorization of waste cabbage leaves by postharvest photochemical treatments monitored with a non-destructive fluorescence-based sensor”  
by Kowalski et al.

has been revised according to the Editor and the Reviewers comments and requests.

We carefully addressed them and hope that now the manuscript could be suitable for publication in the Journal of Photochemistry and Photobiology B: Biology.

The changes/additions are highlighted in the manuscript text.

Reviewer #1

In general, the subject of the work is worthy of investigation and follows current trends to establish which wavelength and intensity of light are more appropriate for inducing valorisation of by-products. There are a number of issues to be addressed, as outlined below, which make the manuscript not yet ready for publication.

-In the material and methods the authors must report which is the % of UV-A wavelength emitted from the broadlamps used in this experiment because the results obtained and reported as a consequence of UV-B irradiation could be derived also from this component.

*Reply:*

*The % of UVA (60%) and UVB /40%) to the total UV have been reported in the text (Lines 198-200).*

*Of course, the presence of UVA radiation may contribute to the treatment effects. This has been highlighted in the text. All over the manuscript, care has been taken to specify that broadband UVB lamp (containing UVA) and not specific UVB radiation was used in the treatments of leaves.*

-In the fig 1 the authors have to explain why at time 0 of treatments the FLAV index of UVAB and CTBAB relating to apical part of the leaves are so different also if not statistically significant

*Reply:*

*It is widely accepted that the FLAV index of leaves is very sensitive to sunlight exposure. In our study we used the cabbage leaves closest to the cabbage head. For those, the most sunlight exposed leaf part is the apical one. Since the leaves treated by the UVB lamp were chosen randomly, the high variability found in the FLAV index on the apical part, especially AB side, are likely due to rather different exposure of the part to sunlight. To note that the apical part of some cabbage leaves could assume a different orientation, more or less folded, exposing to sunlight the AD or the AB side. This explanation has been add in the text. (see Lines 292-293)*

- In my opinion to improve the quality of the manuscript it should be better to insert a chromatogram of HPLC analysis to individuate the profiling of flavonols detected

*A new figure (Fig. 2 in the revised version) reporting chromatograms of leaf extracts of apical and basal portion of cabbage leaves stored with and without UVB treatment is included.*

-The authors have to explain the discrepancy between Fig 3 A and B. In fig 3A as the irradiance decreases, an increase in FLAV index is observed in the basal part of the leaves while in fig 3B the increase of the FLAV index in both AD and AB occurs in conditions of constant brightness. Indeed during the time period considered a reduction of UV-B and UV-A radiations was observed.

*Reply:*

*Comparing the day-by-day levels of FLAV and global solar radiation is not easy since we do not know precisely what could be the lapse time between the radiation stimulus, the synthesis of flavonols and the localization of flavonols on the leaf surface. And if the stimulation of the flavonol accumulation is based on a threshold for irradiance. We can observe that during the first four days of the end of September experiment, global solar irradiance decreased from 10.85 to 7.5 MJ m<sup>-2</sup> day<sup>-1</sup>, while at the end of Oct, global solar irradiance varied less between 4 and 5.4 MJ m<sup>-2</sup>*



*day-1. Therefore, even if global irradiance decreased in September, it likely remained above the threshold to synthesize new compounds.*

*We believe that comparing irradiance (both UV or global) integrated over a time period and the FLAV accumulation rate, as reported in Table 1, is more correct and informative on the FLAV-radiation correlation.*

Reviewer #2

Reviewer #2: Artur Kowalski and co-workers prepare a manuscript describing the effects of postharvest photochemical treatments on waste cabbage leaves by as monitored by HPLC and by a non-destructive fluorescent method. I am not in favor of publication because of several reasons:

1. The manuscript is purely observational. Amount of flavonols are compared with or without irradiation and there is not any tentative description of the mechanisms involved. I understand the study could be published in JPPB if there is mechanistic information, explaining how and why light treatment postharvest will increase levels of polyphenols. The sole information that the irradiation can increase levels of phenols is not new.

*Reply:*

*We thank the Reviewer for the request to introduce the indication about possible mechanisms of photochemical induction of leaf flavonols.*

*We added a new section “3.5 Mechanisms of photochemically-induced flavonols accumulation in waste leaves” reporting mechanisms of action and referring to a schematic representation of them in the a figure (Fig. 6).*

*We also highlighted the effect of the irradiation treatments in enhancing the flavonols content in the abaxial non-irradiated side. This is a quite new information in the field.*

*A novel aspect of the manuscript is also the comparison between two methods of photochemical treatments. And it provides quantitative parameters to optimize the treatment for an industrial level of application.*

2. While the HPLC method is quantitative the indirect fluorescence decrease may be caused by a variety of factors, not only by the increase of polyphenols content. Though I agree that the fluorescence quenching method is valuable as a screening tool, I do not recognize its benefits as a quantitative method for the present publication.

*Reply:*

*The chlorophyll fluorescence excitation filtering method is a widely accepted method to non-destructively quantify both flavonols and anthocyanins on leaf or fruit surfaces. It is based on the ratio between far-red chlorophyll fluorescence detected at two excitation wavelengths, one absorbed by the compound under detection and the other as reference. Therefore, it is very sensitive to the absorbance of leaf/fruit surface at a specific wavelength.*

*It has been applied by our group and others on several species finding good correlations between the optical indices and the actual amount of leaf/fruit compounds determined destructively.*

*We are very confident with that.*

*See for example:*

*Agati et al. J. Agric. Food Chem. 64 (2016) 85–94*

*Agati et al. Functional Plant Biol 33 (2008) 77-84*

*Rybarczyk-Plonska et al. Postharvest Biol. Technol. 116 (2016) 105–114*

*Zivcak et al. Planta 245 (2017) 1225-1229*

*Hagen et al. Postharvest Biol Tech 41 (2006) 156-163*

*Pinelli et al. Food Chem 244 (2018) 213-223*

*The use in the present study of the optical sensor to follow the leaf increase of flavonols during irradiation was fundamental to measure the rates of accumulation and compare different treatments. It allowed to follow in time the very same leaf, no possible with destructive analyses, limiting the biological variability and giving more precise evaluation of the effects.*



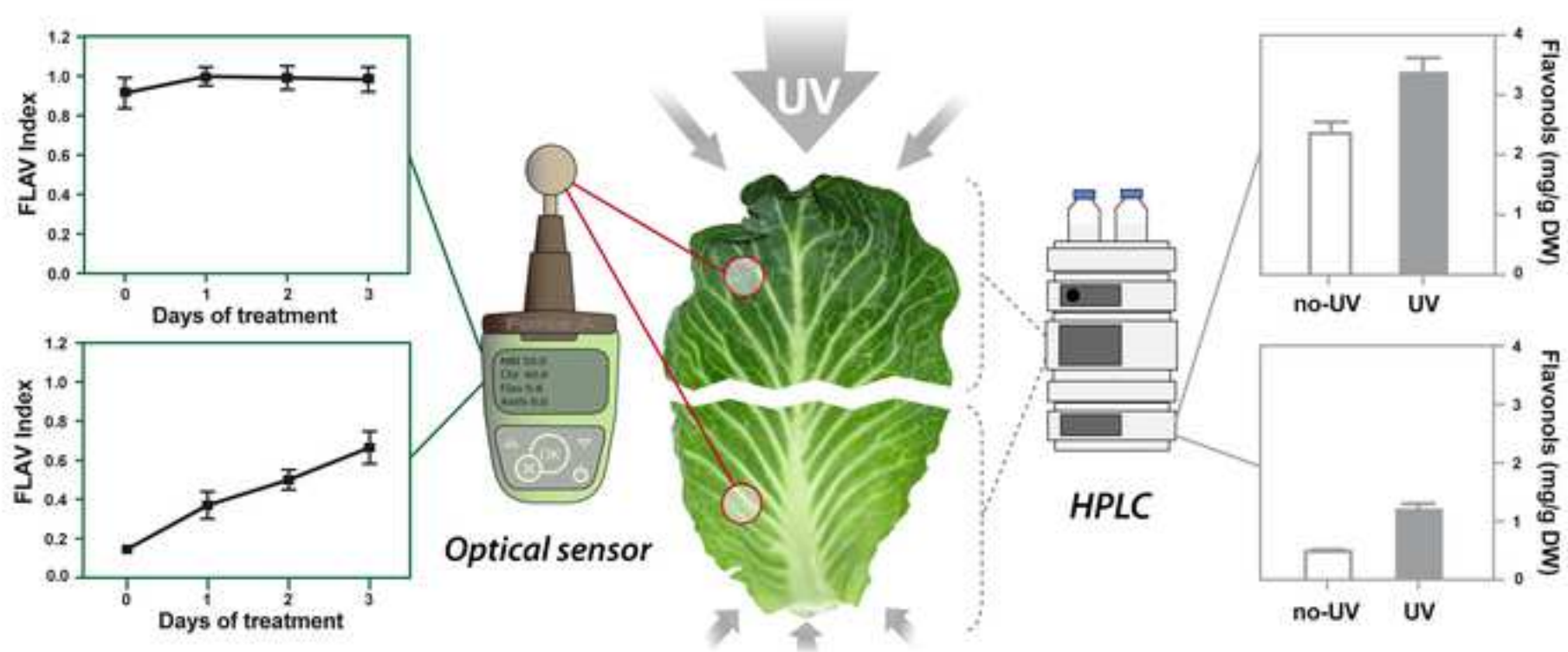
3. The results revealing the significant role for solar UVA and visible radiation in the regulation flavonoids could be better exploited. Why is this the case? Can the authors provide a mechanistic explanation?

*Reply:*

*These mechanisms are reported in the added 3.5 section.*

- Sunlight and UVB postharvest irradiation increase flavonols in waste cabbage leaves
- Non-destructive fluorescence sensors control bioactive compounds in cabbage waste
- Sunlight is an efficient and sustainable valorisation tool of cabbage waste leaves
- Accumulation of flavonols in the basal leaf parts is proportional to irradiance
- Signalling from irradiated leaf sides induces flavonols on unirradiated leaf sides





**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

## Author contributions

Artur Kowalski: Investigation. Giovanni Agati: Conceptualization, Methodology, Writing-Original draft preparation. Maria Grzegorzewska: Investigation, Data curation. Ryszard Kosson: Methodology. Barbara Kuznierewicz: Investigation, Data curation, Visualization. Tomasz Chmiel: Data curation, Validation. Agnieszka Bartoszek: Writing- Reviewing and Editing, Lorenza Tuccio: Investigation. Daniele Grifoni: Formal analysis. Ingunn M. Vågen: Conceptualization, Writing- Reviewing and Editing. Stanislaw Kaniszewski: Funding acquisition, Supervision.

1 Valorization of waste cabbage leaves by postharvest photochemical treatments  
2 monitored with a non-destructive fluorescence-based sensor

3

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26 **ABSTRACT**

27 The biosynthesis of polyphenolic compounds in cabbage waste, outer green leaves of white head  
28 cabbage (*Brassica oleracea* L. var. *capitata* subvar. *alba*), was stimulated by postharvest irradiation  
29 with UVB lamps or sunlight.

30 Both treatments boosted the content of kaempferol and quercetin glycosides, especially in the basal  
31 leaf zone, as determined by the HPLC analysis of leaf extracts and by a non-destructive optical  
32 sensor.

33 The destructive analysis of samples irradiated by the sun for 6 days at the end of October 2015 in  
34 Skierniewice (Poland) showed an increase of leaf flavonols by 82% with respect to controls. The  
35 ~~UVB~~-treatment by a broadband UVB fluorescent lamp, with irradiance of  $0.38 \text{ W m}^{-2}$  in the 290-  
36 315 nm range (and  $0.59 \text{ W m}^{-2}$  in the UVA region) for 12 hours per day at  $17 \text{ }^\circ\text{C}$  along with a white  
37 light of about  $20 \mu\text{mol m}^{-2} \text{ s}^{-1}$ , produced a flavonols increase of 58% with respect to controls.

38 The kinetics of flavonols accumulation in response to the photochemical treatments was monitored  
39 with the FLAV non-destructive index. The initial FLAV rate under the sun was proportional to the  
40 daily radiation doses with a better correlation for the sun global irradiance ( $R^2=0.973$ ), followed by  
41 the UVA ( $R^2=0.965$ ) and UVB ( $R^2=0.899$ ) irradiance.

42 The sunlight turned out to be more efficient than the UVB lamp in increasing the flavonols level of  
43 waste leaves, because of a significant role played by UVA and visible solar radiation in the  
44 regulation of the flavonoid accumulation in cabbage. The FLAV index increase induced on the  
45 adaxial leaf side was accompanied by a lower but still significant FLAV increase on the  
46 unirradiated abaxial side, likely due to a systemic signaling by mean of the long-distance movement  
47 of macromolecules.

8 Our present investigation provides useful data for the optimization of postharvest photochemical  
9 protocols of cabbage waste valorization. It can represent a novel and alternative tool of vegetable  
0 waste management for the recovery of beneficial phytochemicals.

1



52 Keywords:

53 *Brassica oleracea* L.; chlorophyll fluorescence; flavonols; non-destructive sensors; postharvest  
54 irradiation; vegetable waste valorisation.

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57 Abbreviations:

58 AB, Abaxial; AD, Adaxial; Chl, Chlorophyll; ChlF, Chlorophyll Fluorescence; DW, Dry Weight;

59 FLAV, Flavonols index; ROS, Reactive Oxygen Species; UVBE, Ultraviolet Biologically Effective

60 radiation; UVBE<sub>ERYTH</sub>, Ultraviolet Biologically Effective radiation for Erythema induction;

61 UVBE<sub>GPD</sub>, Ultraviolet Biologically Effective radiation for Generalized Plant Damage; UVBE<sub>flav</sub>,

62 Ultraviolet Biologically Effective radiation for flavonols induction.

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## 66 **1. Introduction**

67 Within the growing interest of a sustainable utilization of agri-food waste, significant care is now

68 dedicated to the recovery of bioactive compounds [1]. This represents a worthwhile alternative to

69 the most common utilization of vegetable and fruit wastes as bio-fuel, livestock feeds or as organic  
70 fertilizers.

71 The improvement of green-chemistry technologies permits now to extract high-value products from  
72 wastes and by-products of the agri-food sector [2].

73 This approach can generate benefits for the pharmaceutical, cosmetics as well as food industries as  
4 users of the bioactive natural compounds recovered from waste for the production of nutraceuticals  
5 and functional foods.

6 Moreover, in this way the negative environmental impact due to greenhouse gas emission from  
7 landfill vegetable and fruit residues would be reduced [3].



78 Fruit and vegetable wastes are rich in beneficial for humans and industrially valuable compounds as  
79 recently reviewed [4]. Dietary fibers and plant secondary metabolites can be exploited for their  
80 health properties such as antioxidant, anti-obesity, anti-inflammatory as well as cancer and  
81 cardiovascular disease preventive [1].

82 Among different vegetable wastes, those deriving from Brassicaceae are of particular interest,  
83 because of their significant content of beneficial phytochemicals [5] and large amount of waste  
84 material. In broccoli (*Brassica oleracea* L. var. *italica*) for example, up to 90% of the above ground  
85 plant parts become waste [6]. The non-edible parts of cauliflower (*Brassica oleracea* L. var.  
86 *botrytis*), outer leaves, stems and pods, account for about 36% of the total biomass. Similarly, in  
87 white cabbage (*Brassica oleracea* L. var. *capitata* subvar. *alba*), the percentage of discarded leaves  
88 can reach about 35% of the whole plant [7]. Considering that the world production of cabbage for  
89 2018 was estimated around 70 million of tons [8], cabbage waste can represent about 24 million  
90 tons worldwide.

91 The amount of bioactive compounds contained in the inedible part of broccoli was found to be  
92 superior to that of the edible florets [9,10] and comparable with other vegetables currently used as  
93 food [6].

94 The outer leaves of white cabbage were characterized by a significantly greater nutritional value  
95 compared to marketable heads, inclusive of higher content of total flavonoids, soluble polyphenols,  
96 ascorbic acid and dry matter [7].

97 Wasted leaves are source of dietary antioxidant fiber powder as obtained by recently improved  
98 processing [11]. White cabbage leaf powder was also proposed for the preparation of cakes with  
99 particular qualitative characteristics [12]. Cabbage and cauliflower are likewise reported to be  
0 important sources of isothiocyanates, produced upon hydrolysis of glucosinolates, which are  
1 associated with chemopreventive properties [13,14].

2 The chromatographic LC/MS analysis of aqueous methanolic leaf extracts revealed that flavonols  
3 were the major group of phenolic compounds in cabbage waste. Fourteen quercetin and kaempferol



104 glycosides were detected and characterized [15]. Analogously, nineteen flavonoid glycosides  
105 mainly derived from kaempferol and quercetin were found in cauliflower waste [16].

106 These compounds are of special interest because of their anti-inflammatory and anti-carcinogenic  
107 properties [17,18].

108 There is now large evidence that low-dosage UVB radiation has a stimulatory effect on the plant  
109 secondary metabolism, determining the accumulation of phenolic compounds, in contrast to the  
110 distress damage induced by high-dosage UV [19].

111 Such non-destructive treatments were thus suggested to improve the nutraceutical value of edible  
112 products ensuring high content of health-promoting compounds, both during cultivation as well as  
113 in postharvest [20]. Besides, UVB radiation was found to be effective in increasing the total phenol  
114 content also in broccoli waste [10].

115 Harbaum-Piayda et al.[21] observed a significant increase in the content of quercetin-tri-glycoside  
116 and hydroxycinnamic acids in cabbage head leaves after postharvest low-dose UVB broadband  
117 treatments and low temperature (4 °C) storage.

118 Quantitative determination of bioactive compounds in plant material is usually achieved by high-  
119 performance liquid chromatography and spectrophotometric analysis of solvent extracts of samples.

120 These methods are costly and time consuming, especially for industrial applications, and may  
121 produce artifacts due to compound instability and loss of material. Lately developed optical  
122 methods, instead, allow frequent and rapid monitoring of phytochemicals. The flavonoid content of  
123 leaves can be assessed in situ by non-destructive indirect methods developed within the past 20  
124 years and based on the detection of chlorophyll (Chl) fluorescence [22].

125 The attenuation of the Chl fluorescence signals of leaves excited in the UV spectral range is  
6 proportional to the concentration of the epidermis-located flavonoids and therefore can be used as a  
7 proxy of their leaf content. Even if the method is limited to the detection of epidermal particles, it  
8 can represent a fast useful tool for the rapid screening of the presence of bioactive compounds in  
9 plant material.





130 The technique was previously applied to leaves and fruit skins of different plant species, even  
131 directly in the field by using portable sensors, as reviewed by Julkunen-Tiitto et al. [23]. In  
132 Brassicaceae, non-destructive fluorescence-based methods were previously used to estimate the  
133 flavonol content in curly kale (*Brassica oleracea* L. var. *acephala*) [24], broccoli flower buds  
134 [25,26] and white cabbage [15]. This last study showed that the content of flavonols in cabbage  
135 leaves was following a decreasing gradient from the apical to the basal part, due to the reduced (or  
136 absent) exposure of the basal leaf portion to sunlight.

137 In this work, we exposed waste cabbage leaves to postharvest irradiation by both sunlight and UVB  
138 artificial broadband lamps in order to heighten their phenolic content. Beside wet chemistry, the  
139 effect of the treatments was evaluated by non-destructive optical sensors allowing the monitoring  
140 with time of the leaf flavonol accumulation on the very same leaf material. The study provided  
141 basic data for the development of postharvest photochemical protocols dedicated to cabbage waste  
142 valorization.

143

## 144 **2. Materials & Methods**

### 145 *2.1. Plant material*

146 The white cabbage (*Brassica oleracea* L. var. *capitata* subvar. *alba*), cultivar Typhoon F1, plants  
147 were cultivated in the experimental fields of the Research Institute of Horticulture in Skierniewice,  
148 central Poland.

149 Transplants of cabbage were produced in greenhouse. Seeds were sown to multi-cell trays “Vefi” a’  
150 96 filled with prepared peat substrate “Select” from Klasmann-Deilman. Mean air temperature was  
151 maintained between 20 and 23°C (day) and 17 and 18°C (night) during the course of transplant  
2 production. Transplants at the stage of 5-6 leaves were transplanted to the field on a sandy-loam  
3 soil at a plant distance of 0.6 m and 0.5 m within and between rows, respectively, corresponding to  
4 3.3 plants m<sup>-2</sup>.



155 Nitrogen fertilization was applied at a rate of 200 kg N ha<sup>-1</sup>. The phosphorus and potassium  
156 fertilization was applied according to the results of soil tests by bringing up the soil fertility to 80  
157 mg P and 200 mg K per liter of soil. Irrigation was applied when soil moisture tension at the depth  
158 of 30 cm reached 0.04 MPa.

159 Cabbage heads harvest occurred on 20<sup>st</sup> October 2015 and 21<sup>st</sup> November 2016.

160

## 161 2.2. Non-destructive measurements of leaves

162 The non-destructive determination of leaf flavonols was performed by using the Dualex Scientific+  
163 optical sensor (Force-A, Orsay, France) previously described by Goulas et al. [27]. It is a leaf-clip  
164 active device that uses LED radiation sources at 375 and 650 nm to excite chlorophyll fluorescence  
165 (ChlF) and photodiode detectors to indirectly estimate the in vivo absorbance of the leaf epidermis  
166 by using the ChlF screening method [22].

167 Flavonols present in the epidermis of leaves and absorbing at 375 nm attenuate the incident  
168 radiation, before this can reach the first chlorophyll layer present in the mesophyll. Therefore, the  
169 intensity of the ChlF induced by this radiation (ChlF<sub>UV</sub>) will be inversely proportional to the  
170 epidermal flavonols concentration. Using a red light excitation, not attenuated by flavonols, a ChlF  
171 signal (ChlF<sub>R</sub>) independent of the flavonols concentration is obtained. This signal is used as a  
172 reference. By comparing the ChlF signals from the two different excitations, the index of flavonols  
173 can be calculated (in accordance with the Beer–Lambert law) as the logarithm of the ratio between  
174 the ChlF under red light and that under UV radiation:

$$175 \text{FLAV} = \log(\text{ChlF}_R/\text{ChlF}_{UV}) \quad (1)$$

176 It is worth to remark that by definition the sensor limits the measurement to the leaf epidermis.

7 It was used to measure both the adaxial and abaxial sides of the cabbage leaves on a spot of about  
8 0.5 cm diameter. FLAV values were expressed as Dualex units.

9 The sensor provides also an index of Chl assessed as difference in the leaf light transmission  
0 between 710 and 850 nm.

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### 2.3. UVB lamp treatment of detached leaves

Cabbage outer leaves were collected on October 27<sup>th</sup> 2016. Immediately after harvest, the detached cabbage leaves were kept in a storage room at the temperature of 15-17 °C and 70% relative humidity.

Part of them (12 leaves) were treated daily by a TL 100W/12 RS UVB Broadband Philips fluorescent lamp wrapped with cellulose acetate to remove residual UVC radiation.

The leaves were positioned with their adaxial surface facing the UVB lamp at a distance of about 140 cm and received an irradiance between 0.34 and 0.43 W m<sup>-2</sup> in the 290-315 nm band.

The UVB radiation was measured by the OPHIR PD300-UV (Ophir Optronics Solutions Ltd, Jerusalem, Israel) power meter set at 313 nm and previously calibrated with a portable spectroradiometer (model SR9910-PC; Macam Photometrics Ltd., Livingstone, UK), on a Q-Panel (Q-Panel, Cleveland, OH) UVB lamp having a spectrum very close to that of the used lamp.

Twelve other leaves were positioned within the same storage room, outside the UVB lamp irradiation area, covered by the LEE 226 (Lee Filters, Andover, UK) plastic film that filter out all the UV radiation. These leaves were used as control.

The emission spectrum of a UVB lamp filtered by the cellulose acetate film and the transmittance spectrum of the LEE 226 film are reported in Fig. S1 of the online Supplementary data. From that, the percentage of UVB and UVA to the total UV was calculated to be about 40% and 60%, respectively.

The UVB lamp treatment consisted of a 12 hours period of irradiation and 12 hours of dark carried out for three consecutive days. During the irradiation period a supplementary sodium lamp was switched on.

The PPFD of the sodium lamp was measured by the SunScan sensor (Delta-T Devices, Cambridge, England) and was 19.7 μmol m<sup>-2</sup> s<sup>-1</sup> at the samples level. On the controls under the LEE filter it was 14 μmol m<sup>-2</sup> s<sup>-1</sup>.



207 All the leaves were measured by the Dualex sensor, described above, at two zones, apical and basal,  
208 on three points, adaxial and abaxial. The average of the three punctual measurements was  
209 calculated. Dualex measurements were taken before starting the UVB treatment and after each day  
210 of treatment.

211 At the end of the 3-days treatment, each leaf was divided into three parts, central part was discarded  
212 while apical and basal ones were collected and frozen in liquid N and then freeze-dried for  
213 successive extraction and analysis of phenolics by HPLC.

214

#### 215 *2.4. Sunlight irradiation of waste cabbage leaves*

216 In a first trial, the cabbage leaves closest to the cabbage heads were harvested on Sep 22<sup>nd</sup> 2015 and  
217 left on the ground, with the adaxial side up, under solar irradiation until Sep 30<sup>th</sup>. During this  
218 period, the detached leaves were monitored by the Dualex sensor (Fig. S2 of Supplementary data).

219 In a second trial, the leaves were detached from the plants on Oct 27<sup>th</sup> 2015 and irradiated by  
220 sunlight in the field for 6 days. On Nov 2<sup>nd</sup> 2015, after the Dualex sensor measurements, 9 of the  
221 treated leaves were cut transversally into three parts, the central part was discarded while the apical  
222 and basal ones were then freeze-dried. Three more leaves were collected and treated the same on  
223 Oct 27<sup>th</sup> 2015 just before the irradiation period, and used as control.

224 Freeze-dried samples were extracted and analyzed for their content of polyphenols. In a third trial,  
225 cabbage heads were harvested on Oct 20<sup>th</sup> 2015 in a way to maintain the leaves closest to the heads  
226 attached to the stalk (Fig. S2 of Supplementary data). Twenty-four plants were used. On each plant,  
227 the most exposed leaf was monitored over time by measuring the apical and basal adaxial parts by  
228 the Dualex sensor up to 21 days, till Nov 10<sup>th</sup>.

9 During the trials, the average air temperature was ranging between 11.9 and 13.1 °C at the end of  
0 September and between 10.7 and 11.1 °C at the end of October. Daily average relative humidity  
1 was between 70% and 76% in September and between 83% and 97% in October. The daily mean  
2 global sun irradiance ranged between 0.86 MJ m<sup>-2</sup> (Nov 10<sup>th</sup>) and 10.85 MJ m<sup>-2</sup> (Sep 22<sup>nd</sup>).



233 The daily average values of UVB and UVA were obtained from AC SAF offline surface UV  
234 product database derived from the measurements of the operational polar orbiting Metop and  
235 NOAA satellites with a spatial resolution of 0.5°. Together with solar UVB and UVA, AC SAF  
236 makes also available the daily Ultraviolet Biologically Effective radiation (UVBE) calculated  
237 according to action spectra such as that for Erythema (UVBE<sub>ERYTH</sub>, [28]) and that for the  
238 Generalized Plant Damage (UVBE<sub>GPD</sub>, [29]). Unfortunately, AC SAF does not provide the solar  
239 UVBE daily dose calculated according to the ~~using the~~ biological weighting function for the  
240 induction of flavonols in *Mesembryanthemum crystallinum* (UVBE<sub>flav</sub>) obtained by modelling the  
241 plant response to different cut-off filtered radiation between 280 and 360 nm [30] and considered  
242 more appropriate to evaluate the flavonol response in this study.

243 The STAR (System for Transfer of Atmospheric Radiation) model [31] was used to simulate clear  
244 sky solar spectral radiation at 30 min-time step intervals for the experimental site [32]. The  
245 resulting solar emission spectrum on Nov 1<sup>st</sup> 2015 under clear sky conditions is reported in Fig. S1.  
246 From the sun spectra available during the September-November 2015 period, the daily UVBE<sub>GPD</sub>  
247 and UVBE<sub>flav</sub> were calculated and their interrelationship under clear sky was defined. This was then  
248 used to estimate the solar UVBE<sub>flav</sub> over the experimental period from the UVBE<sub>GPD</sub> values  
249 obtained by AC SAF, in order to take into account the possible cloud attenuation.

250 The daily average values of UVB, UVA and UVBE<sub>flav</sub> doses are reported in Fig. S3 of online  
251 Supplementary data.

### 252

### 253 *2.5. Chromatographic determination of leaf flavonols*

254 The method based on RP-HPLC with UV and MS detections ~~was~~ used for the quantitative and  
5 qualitative determination of flavonols in freeze-dried cabbage leaves after acid hydrolysis of parent  
6 glycosides ~~is~~ as described elsewhere [15]. The freeze-dried cabbage powders (60 mg) were extracted  
7 with the use of 1 mL of methanol/water (7:3, v/v). The extracts were sonicated (10 min),  
8 centrifuged (3000 rpm, 15 min) and clear supernatants collected. The extraction procedure was

259 repeated with a new portion of solvent (1 mL). The supernatants from two repeated extractions were  
260 combined. The portions of final extracts (1 mL) were used for acidic hydrolysis (90 °C for 120 min)  
261 by hydrochloric acid (3.233 M, 0.59 mL). The obtained hydrolysates were filtered through a 0.45  
262 µm Millex HV filter (Merck Millipore, Warsaw, Poland) and immediately injected (20 µL) to  
263 HPLC-DAD-ESI-MS (1200 series with 6130 Quadrupole, Agilent Technologies, Santa Clara, CA).  
264 Phenomenex Kinetex XB-C18 100A column (150 mm x 4.6 mm, 5 µm) was used for  
265 chromatographic separation. Gradient elution was carried out using a mobile phase consisting of  
266 0.1% formic acid in water (solvent A) and 0.1% formic acid in methanol (solvent B). The HPLC  
267 system was operated at flow-rate of 1 mL/min. The elution gradient profile used was 10-100% B in  
268 30 min, followed by 5 min 100% B. The column was allowed to equilibrate between the injections  
269 for 5 min with the initial composition of mobile phase. Absorbance spectra were recorded between  
270 190 and 700 nm, while the chromatograms were monitored at 350 nm. MS parameters were as  
271 follows: capillary voltage, 3000 V; fragmentor, 120 V; drying gas temperature, 350 °C; gas flow  
272 (N<sub>2</sub>), 12 L/min; nebulizer pressure, 35 psig. The instrument was operated both in positive and  
273 negative ion modes, scanning from m/z 100 to 1200. The peaks were tentatively identified by  
274 comparison of UV and MS spectra with those for standards and literature data.

275 Total flavonols content in dry weight of cabbage leaves was calculated as a sum of aglycones and  
276 flavonol monoglycosides remaining after hydrolysis. The quercetin and kaempferol derivatives  
277 were quantitated based on external standards of respectively quercetin and kaempferol, respectively  
278 (Sigma Chemical Co., St. Louis, MO).

279

## 280 2.6. Data analysis

1 Statistical analysis and curve fitting were carried out with the SigmaPlot for Windows Version 12.5  
2 software (Systat Software, Inc., San Jose, CA). Mean values of data underwent t-test or One Way  
3 Repeated Measures ANOVA and were compared by the All Pairwise Multiple Comparison Holm-  
4 Sidak Test. P-values <0.05 were considered statistically significant.



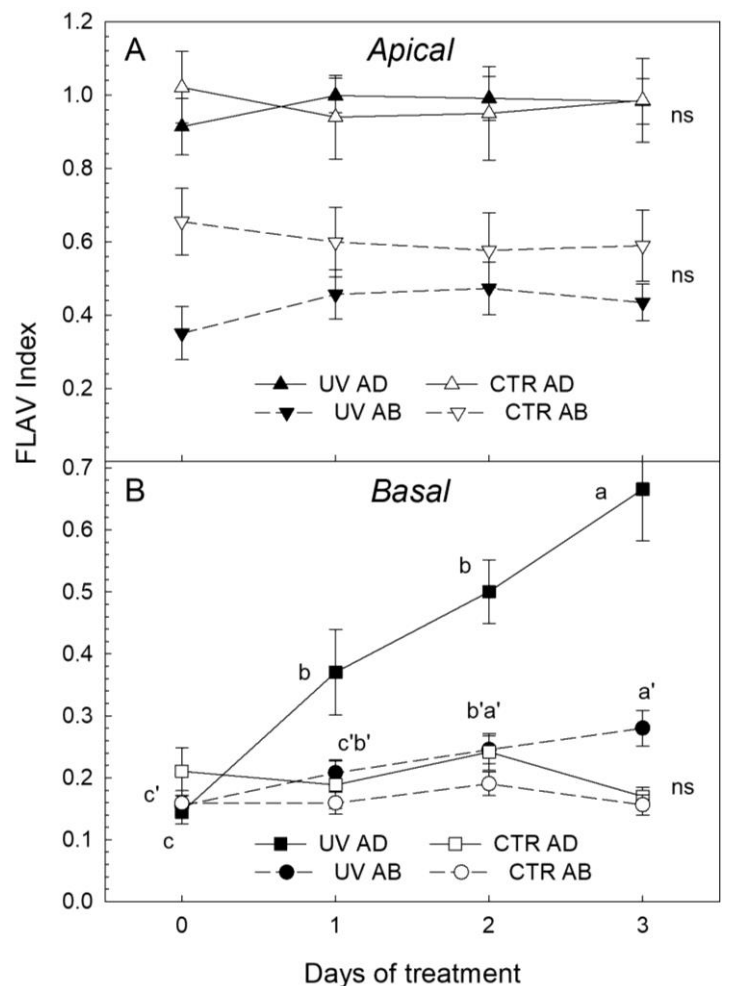
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### 286 3. Results and Discussion

#### 287 3.1 UVB lamp irradiation of waste cabbage leaves

288 In Figure 1, the flavonol FLAV index of harvested cabbage leaves monitored before and during  
289 storage with or without the UVB lamp treatment is reported. At harvest, the adaxial (AD) apical  
290 part of leaves showed FLAV values almost double that the abaxial (AB) apical part indicating that  
291 AD sides were more exposed to sunlight than AB sides (Fig. 1A).

292 The higher variability of FLAV observed on the AB leaf apical parts than the AD ones before the  
293 UVB lamp treatment, was likely due to their more or less folded orientation towards the sun.



7  
8 Figure 1. Kinetics of the FLAV index (mean  $\pm$  SE, n=12) measured during storage of detached  
9 cabbage leaves with (closed symbols) or without (open symbols) the UVB lamp treatment (12 h d<sup>-1</sup>)  
0 on the apical (A) and basal (B) leaf portion. In B, values of UV AD (adaxial leaf side) and UV AB

311 (abaxial leaf side) followed by a different letter and primed letter, respectively, are significantly  
312 different ( $P < 0.05$ ) according to the Holm-Sidak Test, ns = not significant.

313

314 Yet, on the basal part of leaves the lowest values of FLAV (around 0.15-0.2) were recorded (Fig.  
315 1B). The difference in the FLAV index between apical and basal leaf zones is in agreement with the  
316 cabbage leaf longitudinal gradient of indices observed before [15].

317 During storage, no significant changes in FLAV were observed on the apical part of both UVB  
318 treated and untreated leaves ( $P > 0.5$ , one way Repeated Measures ANOVA). Instead, a marked  
319 increase in FLAV ( $P < 0.001$ ) was found on the basal AD sides of leaves (Fig. 1B). The increase in  
320 FLAV with UVB energy dose was linear with a rate of 0.0102 Dualex units per  $\text{kJ m}^{-2}$  (considering  
321 an average daily UVB dose of  $16.6 \text{ kJ m}^{-2}$ ).

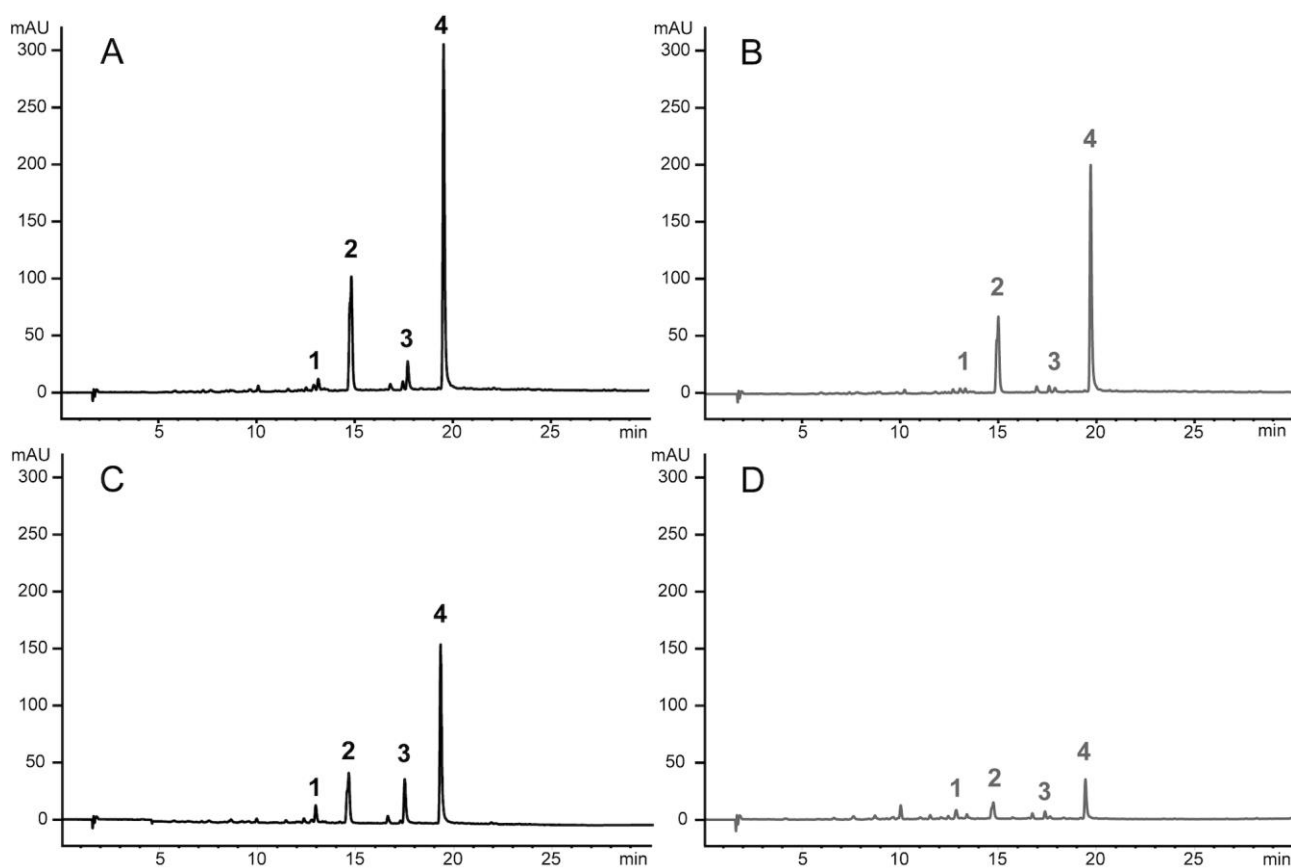
322 A slighter but still significant ( $P < 0.001$ ) increase in FLAV with UVB treatment was also observed  
323 on the basal AB side of treated leaves, while flavonols content in controls did not change with  
324 storage on either leaf sides (Fig. 1B). A possible explanation for the enhancement of FLAV in the  
325 AB non-irradiated leaf surface in given below in the 3.5 section.

326 The HPLC analysis of leaf methanolic extracts confirmed that flavonols were the major group of  
327 phenolic compounds in cabbage leaves, as previously observed [15].

328 The chromatograms of apical and basal leaf extracts recorded at 350 nm shown in Figure 2 indicate  
329 the flavonols profile found in the UV-treated and control samples.

330



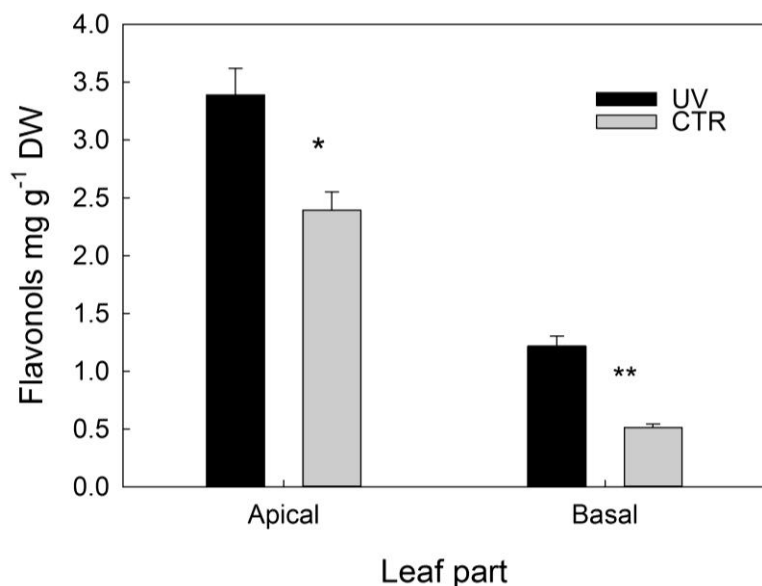


331  
332  
333  
334 Figure 2. High performance liquid chromatography profiles registered at 350 nm of the flavonols  
335 liberated by acid hydrolysis from crude extracts of apical (A, B) and basal (C, D) portion of  
336 cabbage leaves stored with (A, C) and without (B, D) the UVB treatment. The four flavonols were  
337 identified as 1 quercetin glycoside, 2 kaempferol glycoside, 3 quercetin, and 4 kaempferol.

338  
339 The identity of compounds remaining after acid hydrolysis of parent flavonols was confirmed based  
340 on MS fragmentation pattern characterized by major molecular ion peak, that in the case of positive  
341 ionization occurs as  $[M+H]^+$  and in negative ionization as  $[M-H]^-$ . The presence of two flavonol  
2 aglycons: quercetin (303 Da  $[M+H]^+$ , 301 Da  $[M-H]^-$ ) and kaempferol (287 Da  $[M+H]^+$ , 285 Da  
3  $[M-H]^-$ ) and two flavonol monoglycosides: quercetin glycoside (465 Da  $[M+H]^+$ , 463 Da  $[M-H]^-$ )  
4 and kaempferol glycoside (449 Da  $[M+H]^+$ , 447 Da  $[M-H]^-$ ) was observed in cabbage hydrolysates.

345 The total content of flavonols in dry weight of cabbage leaves was calculated as a sum of these four  
346 compounds and with the use of calibration curves generated for standards of quercetin and  
347 kaempferol.

348 The flavonol content determined by the destructive (chromatographic) analysis in samples stored  
349 for three days with or without the UVB lamp radiation is reported in Fig. 32. It showed that the  
350 UVB-treatment increased the flavonol content compared to controls on both leaf zones, by 42% and  
351 136% on the apical and basal leaf parts, respectively. On average, the flavonol content of cabbage  
352 leaves after 3-days of UVB lamp irradiation (at about 0.38 W/m<sup>2</sup>) rose to 4.61 mg g<sup>-1</sup> DW, and to  
353 2.91 mg g<sup>-1</sup> DW in untreated leaves.



354  
355 Figure 32. Leaf flavonols concentration (mean  $\pm$  SE, n=12) as determined by HPLC on extracts  
356 from the aApical and bBasal parts of detached leaves stored for three days with (black bars, UV) or  
357 without (gray bars, CTR) the 12 h d<sup>-1</sup> UVB lamp treatment. The t-test gave a significant difference  
358 of \*P=0.002, \*\*P<0.001.

9  
0 While the results obtained by the HPLC analysis for the basal leaf parts agree with those observed  
1 by the Dualex sensor, the FLAV index failed to reveal the increase in the leaf flavonol content on  
2 the apical leaf part though it could be measured by HPLC following the leaves destruction.

363 This discrepancy suggests that the UVB [lamp](#)-induced accumulation of flavonols is not limited to  
364 the leaf outer layers but occurs also in the mesophyll. The amount of flavonols localized in the  
365 palisade and spongy tissues cannot be measured, by definition, by the Dualex, while they are  
366 quantified by the destructive analysis of the whole leaf extracts.

367 It is noteworthy that the FLAV index detected an increase in flavonols on the AB epidermis of basal  
368 leaf parts even if these were not directly exposed to the UV radiation. It is evident that a UV-  
369 induced stress signaling occurs through all the leaf tissue layers.

370 Postharvest UV irradiation effects on different plant materials were previously reported. The UVB  
371 [broadband lamp](#) treatment of tomato fruits at the dose of about  $6 \text{ kJm}^{-2}$  per day ( $1.69 \text{ Wm}^{-2}$ ) for 10  
372 or 18 days, depending on the ripening stage at harvest, increased the level of ascorbic acid,  
373 carotenoids [33] and polyphenols [34]. As little as 3 min of  $0.43 \text{ Wm}^{-2}$  of UVBE<sub>GPD</sub> was sufficient  
374 to significantly increase the level of secondary metabolites in the flavedo of postharvest lemons  
375 [35].

376 Even a short UVB [lamp](#) irradiation of 5 min for 3 days inside a refrigerator induced the increment  
377 of flavonoids in fresh detached leaves of spinach, radish and parsley during the consecutive 3 days  
378 of storage [36]. This was related to the expression enhancement by UVB of all the genes involved  
379 in the flavonoid biosynthesis that reached its maximum at 12 h after irradiation. A similar delayed  
380 response to low-dosage UVB [lamp](#) pre-harvest treatment in increasing phenolic compounds during  
381 storage was observed in *Ocimum basilicum* [37].

382 In the *Brassica* species, an increase of flavonols in broccoli (*Brassica oleracea* L. var. *italica*)  
383 flower buds irradiated postharvest with a combination of visible light ( $19 \mu\text{mol m}^{-2} \text{ s}^{-1}$ ) and low-  
384 dose UVB ( $0.23 \text{ W m}^{-2}$ ) [broadband lamp](#) was observed by both destructive and non-destructive  
5 tools [26]. While Harbaum-Piayda et al. (2016) reported the induction by [a](#) low-dosage UVB [lamp](#)  
6 of quercetin-tri-glycoside in white cabbage heads.

7 Our results showed that the increase of flavonols induced by the postharvest UVB [lamp](#) treatment  
8 on the basal part, not exposed to sun, of waste leaves was  $0.7 \text{ mg/g DW}$ , somewhat higher than that



389 observed by Harbaum-Piayda et al. [21] in cabbage head leaves by a similar treatment. The  
390 difference can be due to the higher storage temperature (17 °C vs 4°C) used in the present study  
391 and/or the lack of supplemental white light in the previous study. In fact, maintaining the light/dark  
392 cycle during postharvest storage of Brassicaceae is important to preserve quality and  
393 phytochemicals [38].

394 Recently, a study related to the present one with postharvest UVB (1.4 W m<sup>-2</sup>) broadband lamp in  
395 combination with white light treatments of leaf waste fractions from industrial trimming of white  
396 cabbage has showed an increased flavonol level of about 0.5 mg g<sup>-1</sup> DW compared to untreated  
397 controls (Seljåsen and Vågen 2020, personal communication). Treatment with UVB alone resulted  
398 in the formation of just half the amount of flavonols compared to treatments where UVB was  
399 combined with white light. Interestingly, increasing the UVB dose (by 1.7 time) in combination  
400 with white light did not enhance the flavonol concentration.

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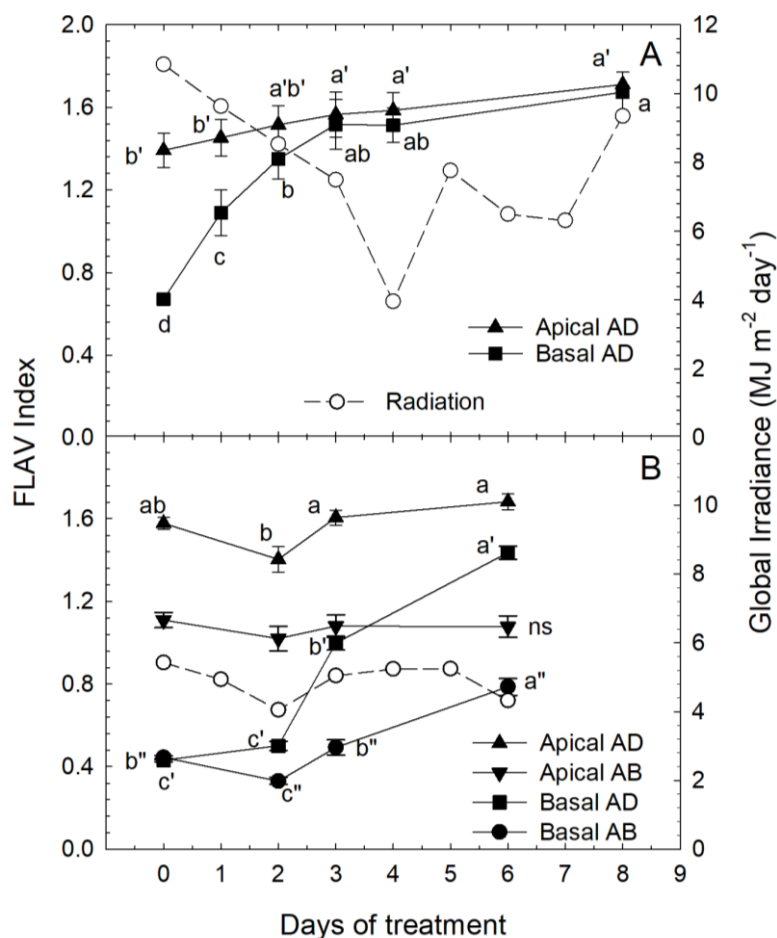
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412 Figure 43. Evolution of the mean ( $\pm$ SE) FLAV index in detached cabbage leaves exposed to  
 413 sunlight from Sep 22 to Sep 30 (A, each point  $n=4$ ) and from Oct 27 to Nov 2, 2015 (B, each point  
 414  $n=13-24$ ). For each leaf part/side, values followed by a different letter are significantly different  
 415 ( $P<0.05$ ) according to the Holm-Sidak Test, ns = not significant.

416

417 All the above observations are explained consistent with by the UV-induced increase of the activity  
 418 of phenylalanine ammonia-lyase and of other enzymes participating in the flavonoid synthesis such  
 419 as chalcone synthase, chalcone isomerase and dihydroflavonol-4-reductase, as explained later  
 0 (section 3.5). This process must take place in post-harvest the same way it occurs during pre-harvest  
 1 UV treatments of plants [39].

2

3

### 424 3.2 Sun-exposure of waste cabbage leaves

425 Figure 43 reports the effect of sun irradiation on the FLAV index of detached cabbage leaves during  
426 two different periods, end of Sep (Fig. 43A) and end of Oct (Fig. 43B). In both cases, a large  
427 increase of flavonols in the basal leaf part was observed, while the apical part of leaves was only  
428 slightly affected by the irradiation. This result was expected since before harvest the apical parts of  
429 leaves were already acclimated to sunlight while the basal parts were sun-screened by the closest  
430 other leaves and heads.

431 As observed in the UVB lamp experiment, the FLAV index significantly increased even in the AB  
432 epidermis of basal leaf parts not directly exposed to sunlight, evoking again the occurrence of a  
433 stress signaling through the leaf tissues (see section 3.5).

434 The trial was repeated on leaves maintained attached to the plant after removing the cabbage heads  
435 to increase leaf irradiance (Fig. S2 of Supplementary data). In Fig. 54, the temporal average change  
436 of the leaf FLAV index from the start of exposure up to 3 weeks is reported. The flavonols content  
437 in the apical leaf part slightly decreased at 6 days of treatment but recovered the initial value at 10  
438 days after treatment and maintained it for the next 11 days. On the leaf basal part, a continuous  
439 increase of flavonols was observed. This observation may suggest a saturation of plant capability of  
440 secondary polyphenolic metabolites biosynthesis in the sun exposed apical part of the leaves.

441 The effect of the different solar irradiation treatments was evaluated by comparing the initial rate of  
442 FLAV enhancement on the AD basal leaf parts. The highest slope of the linear phase of the FLAV  
443 evolution was observed for detached leaves irradiated at the end of September (0.28 Dualex units  
444 day<sup>-1</sup>), while at the end of October, still on detached leaves, it was 0.18 Dualex units day<sup>-1</sup>. The  
445 slowest in field treatment effect was recorded on the attached leaves at the end of October (0.09  
6 Dualex units day<sup>-1</sup>). These differences are consistent with the difference in the average global, UVA  
7 and UVB irradiance observed during the relative periods: 22-25 Sep, 20-26 Oct and 27 Oct-2 Nov,  
8 2015, as reported in Table 1. Moreover, attached leaves may have been oriented in a way to reduce  
9 full exposure to sunlight.



450 A linear direct regression between the FLAV accumulation rate and any of the daily radiation doses  
 451 was observed, however, the correlation was better for the sun global irradiance ( $R^2=0.973$ ),  
 452 followed by the UVA irradiance ( $R^2=0.965$ ). The less accurate correlation ( $R^2= 0.899$ ) was found  
 453 for the UVB irradiance.  
 454 Accordingly, the FLAV rate correlated to  $UVBE_{flav}$  similarly to UVA and better than to UVB doses  
 455 (Table 1). This is because the action spectrum used in calculating  $UVBE_{flav}$  attributes a biological  
 456 effect, in inducing the synthesis of flavonoids, not only to the UVB, but also to the UVA component  
 457 of solar radiation.

Table 1. Average daily sun radiation doses and initial rate of FLAV enhancement on the AD basal leaf parts.

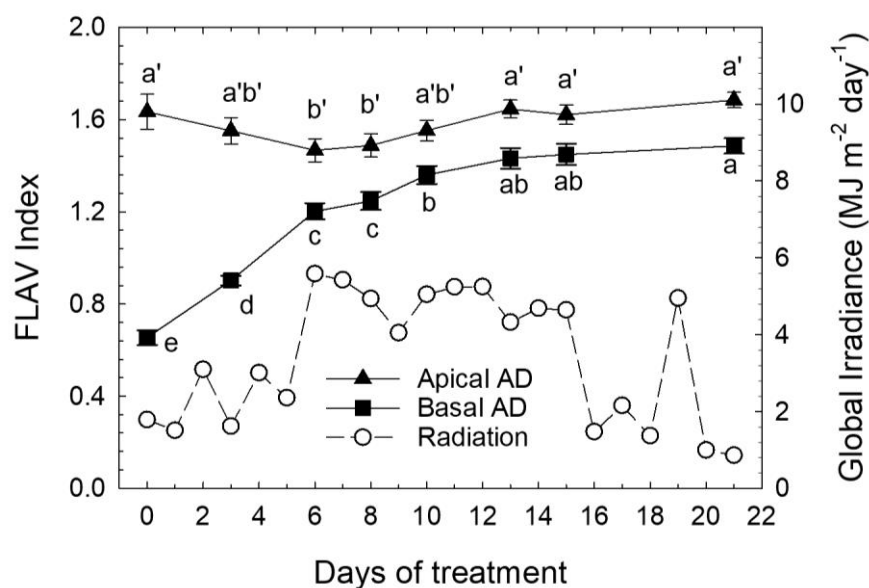
2015 periods	Global MJ m <sup>-2</sup> day <sup>-1</sup>	UVA kJ m <sup>-2</sup> day <sup>-1</sup>	UVB kJ m <sup>-2</sup> day <sup>-1</sup>	$UVBE_{flav}$ kJ m <sup>-2</sup> day <sup>-1</sup>	FLAV rate Dualet units day <sup>-1</sup>
22-25 Sep	9.1	646	10.7	5.05	0.28
20-26 Oct	2.7	215	2.4	1.20	0.09
27 Oct - 2 Nov	4.8	347	3.9	2.18	0.18
$R^{2,a}$	0.973	0.965	0.899	0.94	

<sup>a</sup>Coefficient of determination for the regression between the FLAV rate and the radiation dose of each column.

458

459 Our results indicate that detached leaves maintain the mechanisms to activate the biosynthesis of  
 460 secondary metabolites in response to the sunlight irradiation, as it occurs in intact plants. The main  
 461 difference in this effect between attached and detached leaves was likely due to the different sun  
 462 irradiance of treatments that determined the rate of the process. At longer irradiation times, in fact,  
 463 the extent of flavonol accumulation in both attached and detached leaves was similar (compare  
 464 Figs. [54](#) and [43](#)).

5



466

467 Figure 54. Evolution from Oct 20<sup>th</sup> 2015 of the mean ( $\pm$ SE) FLAV index in cabbage adaxial leaves  
 468 attached to the plant exposed to sunlight after head removal. Each point is the average of 24  
 469 different samples. For Basal and Apical samples, values followed by a different letter are  
 470 significantly different ( $P < 0.05$ ) according to the Holm-Sidak Test.

471

472 The destructive HPLC analysis confirmed the effect of solar irradiation in increasing the flavonol  
 473 content of waste cabbage leaves estimated by the Dualex sensor, mainly in the basal part (Table 2).  
 474 The mean flavonol content in the apical part of irradiated detached leaves was 1.4 times higher than  
 475 that of non-irradiated leaves. In the basal part, flavonol levels increased 4.1 times after irradiation.

476 On average, the total flavonol content of cabbage leaves after 6-days of sun irradiation (at the end of  
 477 October) increased to 5.032 mg g<sup>-1</sup> DW with respect to 2.776 mg g<sup>-1</sup> DW of untreated leaves. This  
 478 effect was markedly similar to that obtained by exposing leaves to the artificial UVB lamp radiation  
 479 under storage (Fig. 32).

0 When cabbage is harvested in the field, a number of leaves surrounding the cabbage head will  
 1 normally be left intact on the plant. A practical inference of our study is that these leaves, when left  
 2 in the field for a time, will accumulate flavonols, and will represent a valuable valorized source of  
 3 nutraceuticals.



Table 2. Leaf flavonols content (mean  $\pm$  SE), mg/g DW, as determined by HPLC on extracts from the Apical and Basal parts of detached leaves exposed for 6 days to sunlight from Oct 27 to Nov 2, 2015 (n=9). Values are compared with those determined on just harvested not exposed leaves (n=3).

Leaf part	Postharvest sun-exposure (6-days)		At harvest
Apical	3.27 $\pm$ 0.14	*	2.34 $\pm$ 0.09
Basal	1.75 $\pm$ 0.07	**	0.42 $\pm$ 0.04

485 The t-test gave a significant difference between exposed and not exposed leaves, apical \*(P=0.004)  
 486 and basal \*\*(P<0.001) values.  
 487

488

### 489 3.3 Comparison of treatments

490 The in field sun-induced increase rate in leaf FLAV under postharvest at the end of Oct 2015 on  
 491 detached leaves was similar to that induced by the UVB [lamp](#) radiation under storage, 0.18 versus  
 492 0.17 Dualex units day<sup>-1</sup> on the AD leaf basal zones. However, the proper comparison of the  
 493 efficiency of the two photochemical treatments must be fulfilled considering the intensity and  
 494 spectral emission of the irradiation sources.

495 The daily doses of the UV lamp were 16.6 kJ m<sup>-2</sup> and 25 kJ m<sup>-2</sup>, for UVB and UVA, respectively,  
 496 that is about 1.6 times higher and 9 times lower than the maximal UVB and the minimal UVA  
 497 amount of radiation, respectively, seen by the cabbage leaves during the sun treatments (see Table  
 498 1). The emission spectrum of the UV lamp was, of course, much different from that of the sun,  
 499 especially late in the season at the latitude of the experimental site (Fig. S1 of the Supplementary  
 500 material). [The UVA/UVB ratio was 1.5 and around 89 for the UVB lamp and the Oct sun radiation,](#)  
 501 [respectively.](#) Therefore, their effectiveness in producing leaf flavonols should be evaluated by

2 weighting the emission spectra by the action spectrum for the synthesis of flavonols in plants. For  
 3 this, we used the UVBE<sub>flav</sub> biological weighting function defined by Idbah et al. [30] for the  
 4 production of mesembryanthin, the main flavonol in *M. crystallinum*. The resulting UVBE<sub>flav</sub> daily  
 5 dose for the UV lamp was 9.12 kJ m<sup>-2</sup>, that is 80% higher than the maximal UVBE<sub>flav</sub> dose received



506 by the leaves during the sun-treated trials (5.05 kJ m<sup>-2</sup>, Table 1). According to the linear regression  
507 between the FLAV rate and the sun UVBE<sub>flav</sub> (Table 1), we expected a FLAV rate induced by the  
508 UVB lamp about 2.8 folds higher than the 0.17 Dualex units day<sup>-1</sup> actually observed. This  
509 discrepancy can be explained by the fact that the biological weighting function used does not  
510 properly take into account the role of solar UVA and visible radiation in the synthesis of the  
511 specific flavonols of cabbage. Yet, it has been shown that UVB radiation is not a prerequisite for  
512 the biosynthesis of flavonoids [22,40], and that blue light can be an inducer of quercetin and  
513 kaempferol compounds even moreas effective thanas UVB radiation [41].

514 Our results, then, revealed the significant role for solar UVA and visible radiation in the regulation  
515 of the accumulation of flavonoids in cabbage leaves.

516 Although the sun photochemical treatment of waste leaves is largely dependent on the weather  
517 condition at cabbage harvest time, it could be considered superior to the UVB lamp treatment,  
518 because of its larger efficiency and sustainability.

519

#### 520 3.4 Effect of leaf detaching on polyphenols

521 Additional stresses faced by waste leaves as cutting and desiccation [42] may have induced a  
522 further raise of polyphenols over that caused by UV and sunlight irradiation alone.

523 Wounding of fresh produce may elicit an increase in antioxidant capacity and phenolic compounds,  
524 however, this effect is markedly species-dependent [43]. In the case of short-term storage of  
525 broccoli, some contradictory data are reported into the literature. Total flavonoid content of five  
526 broccoli cultivars were observed to decrease during three days of storage of florets at 20°C in  
527 polyethylene bags [44]. While, Starzyńska et al. [45] found that short-term storage of broccoli heads  
8 at room temperature caused the significant increase of antioxidant activity and the accumulation of  
9 phenylpropanoids and flavonoids. The accumulation of phenolic compound during storage can  
0 also depend on the day time of broccoli harvest [46].



531 In white cabbage, cutting produced the increase of PAL-activity in detached leaves, but it had no  
532 significant effect on the total soluble phenolics during two days of storage at 15 °C [43].  
533 No change in flavonols was observed in white cabbage heads stored for 4 months at 1-2°C and at  
534 80-85% of relative humidity [47].  
535 The actual amount of phenolic compounds measured in plant material results from the balance  
536 between their synthesis and utilization rates. In this equilibrium, a fundamental role is played by the  
537 plant primary antioxidants, i.e., antioxidant enzymes and low molecular-weight compounds, such as  
538 ascorbic acid and glutathione, that constitute the first line of protection against oxidative stress.  
539 Generally, the primary antioxidants are able to detoxify the reactive oxygen species (ROS)  
540 produced under mild to moderate stresses, but they may result not enough to face severe or multiple  
541 stresses [48].  
542 The consequent depletion of antioxidant enzymes can activate the biosynthesis of secondary  
543 antioxidants, including flavonoids [49]. Indeed, it was observed that vegetables with lower or higher  
544 levels of ascorbic acid were associated to an increase or reduction, respectively, of phenolic content  
545 after wounding [43].  
546 Accordingly, we can expect that in white cabbage detached leaves studied here, the rates of  
547 phenolic synthesis and decrease due to mechanical injuries would be similar and that the increase of  
548 the flavonols production was induced when the irradiation stress was added.  
549 Monitoring the leaf chlorophyll content can be considered a good indicator of early postharvest  
550 senescence [43,45]. In the studied cabbage samples, Chl evolution on the very same leaves was  
551 controlled non-destructively by the Dualex sensor, as reported in Table 3.  
552 From the previously defined calibration curve for the Dualex sensor [15], the Chl concentration at  
3 harvest for the different trials was ranging between 81-113 and 15-19  $\mu\text{g cm}^{-2}$  for the apical and  
4 basal parts of leaves, respectively.  
5 The index of Chl of the basal leaf parts showed no significant changes during the irradiation  
6 treatments (Table 3). A small decrease in the apical leaf part Chl was only observed between time 0



557 and 3 days of sunlight irradiation at the end of September 2015 (P=0.024). A significant decrease of  
 558 Chl was instead found in the apical part of leaves stored with or without the UVB lamp radiation in  
 559 the 2016 experiment.

560

Table 3. Leaf Chl content (Dualex units) estimated non-destructively by the optical sensor on cabbage leaves under the different irradiation treatments.

Days of treatment	Sun-exposure				Storage Nov 7-10, 2016			
	Sep 22-30, 2015		Oct 27 – Nov 2, 2015		Under UVB		Without UVB	
	leaf part		leaf part		leaf part		leaf part	
	Apical	Basal	Apical	Basal	Apical	Basal	Apical	Basal
0	66.1±1.8a	14.8±2.6	61.5±1.3	14.8±1.3	54.5±2.0a	16.4±1.8	58.9±1.9a	17.7±1.4
1	59.8±1.9ab	13.0±1.5			56.6±2.1a	17.6±1.6	57.0±1.9ab	16.6±1.7
2	61.3±1.0ab	14.4±1.9	57.5±2.0	16.9±1.4	46.4±2.3b	13.9±1.0	49.8±2.8bc	18.4±1.2
3	57.7±0.8b	11.9±1.1	59.4±1.2	16.2±1.2	37.9±2.4c	16.7±1.4	47.2±2.7c	15.6±1.2
4	60.0±1.4ab	12.8±1.4						
6			57.4±1.1	12.4±0.9				
8	59.8±2.3ab	18.8±2.6						
P*	0.032	0.207	0.126	0.072	<0.001	0.324	0.002	0.495

561 \*Probability of the ANOVA test. For each column, values followed by a different letter are significantly  
 562 different (P<0.05) according to the Holm-Sidak Test.

563

564 These results suggest that there is not a direct correlation between the early postharvest senescence  
 565 induced by leaf detaching and dehydration and the increase of the FLAV index (compare Figs 1, ~~4~~  
 566 and Table 3), confirming the main role of irradiation in determining the accumulation of flavonols  
 567 compounds. On the other hand, during the 2015 open-field trials on detached leaves the average air  
 568 temperature was ranging between 11.9 and 13.1 °C at the end of September and between 10.7 and  
 569 11.1 °C at the end of October, while relative humidity was always higher than 70%. Therefore,  
 570 under these conditions leaf desiccation could have been rather limited, with a minimal effect on the  
 1 production of secondary metabolites.

2

3

4

### 3.5 Mechanisms of photochemically-induced flavonols accumulation in waste leaves

A schematic representation of the mechanisms involved in the photo-induced accumulation of flavonols in the basal part of cabbage leaves is reported in Fig.6.

It is widely known that plants respond to stressful UV radiation activating protective mechanisms that combine the accumulation of UV-absorbing compounds in the outer epidermal layers and the enhancement of leaf antioxidants.

In the UVB range, the plant response is mediated by the UVR8 (UV RESISTANCE LOCUS 8) photoreceptor that absorbs radiation by mean of a tryptophan chromophore and then interacts with the CONSTITUTIVE PHOTOMORPHOGENIC 1 (CROP1) regulator, leading to the change of gene expression [50]. Besides controlling plant development, this process regulates the leaf accumulation of UV-B-absorbing flavonols.

The UVB-induced expression of genes encoding the key enzymes of the flavonol biosynthesis included PAL, phenylalanine ammonium lyase; C4H, cinnamate-4-hydroxylase; 4CL, 4-coumarate:CoA ligase; CHS, chalcone synthase; CHI, chalcone isomerase; F3H, flavanone 3-hydroxylase; F3'H, flavonoid 3'-hydroxylase; FLS, flavonol synthase and UGT, UDP-dependent glycosyltransferase [50]. This process leads to a significant increase of flavonols in the adaxial epidermis.

UV-induced gene expression could be transferred to deeper leaf tissues, not directly illuminated, by the long-distance movement of microRNAs via the vasculature of the plant [51].

This transmitted signaling can be responsible for the production of a certain amount of flavonols in the abaxial epidermis, as shown by the FLAV index in Figs 1B and 4B.

A similar effect was observed by Bidel et al. [52] when the AD sides of *Centella asiatica* leaves cultivated in a growth chamber were exposed to UVB lamp or full sun radiation. They also observed an increase of flavonols in the mesophyll, by selective leaf tissue extractions, and a FLAV increase in the half leaf adjacent to the irradiated one and protected by an UVB filter [52].



600 Our and these evidences suggest that a systemic translocation of macromolecules [53] could  
601 determine the synthesis of flavonols into the deeper leaf tissues starting from the adaxial UVB  
602 initiated signaling. A direct effect of the radiation penetrating through the palisade and spongy cell  
603 layers is unlikely since the diffusion of both UVB and UVA wavelengths is limited to the first few  
604 tenths of micron of leaf [54].

605 Definition of the mechanisms of the phenylpropanoid biosynthetic pathway activation by solar  
606 radiation is more complex due to the presence of multispectral band effects.

607 The UVA radiation induction of leaf flavonoids is less studied than that of UVB radiation. UVA  
608 effects has been properly reviewed by Verdaguer et al. [55], highlighting contrasting changes and  
609 large variability according to the species, compound specificity, doses of radiation and co-presence  
610 of UVB radiation.

611 In field sun spectral band-exclusion experiments have clearly shown the positive contribution of  
612 UVA and blue radiation in inducing leaf flavonols accumulation [41,56,57].

613 Both UVA and blue radiation are involved in the expression of flavonoid biosynthesis genes and  
614 phenolic accumulation in leaves mediated by the cryptochromes (CRYs) photoreceptors [41,58].

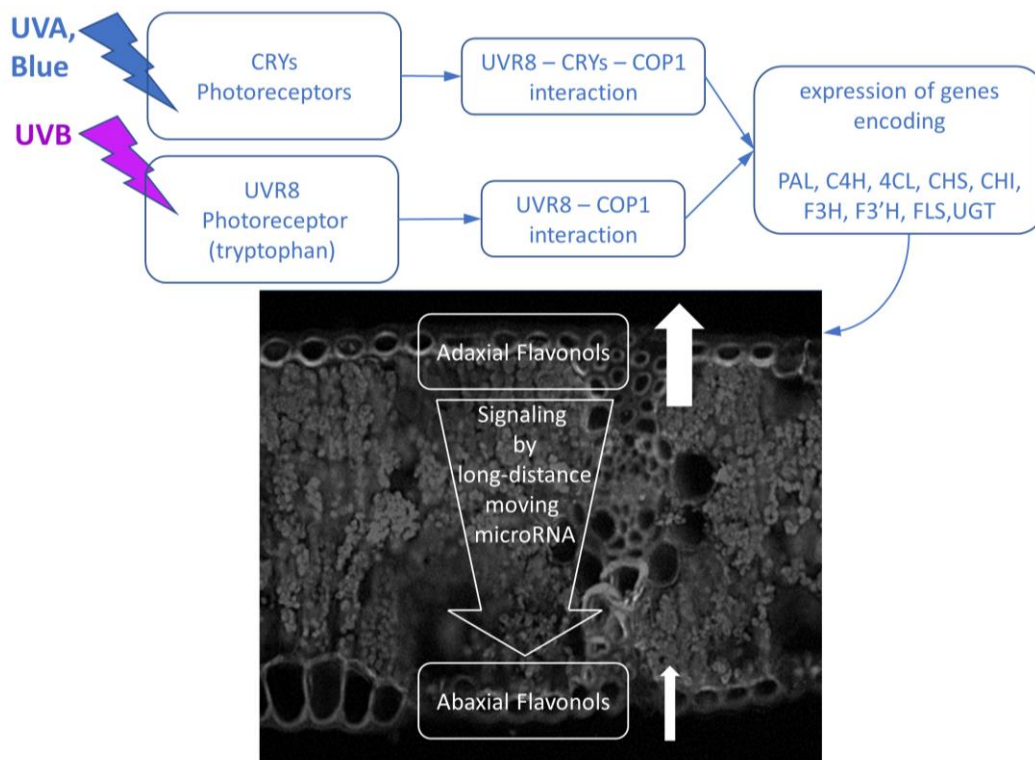
615 This is likely to occur through the interaction between CRYs and UVR8, favored by partially  
616 overlapped absorption spectra, and their binding to COP1 [40].

617 Considering also that the UVA/UVB and Blue/UVB ratios of solar radiation under our experimental  
618 conditions, accounting for about 89 and 260, respectively, were much larger than those in the UVB  
619 lamp (UVA/UVB = 1.5 and Blue/UVB = 0.48), the above proposed mechanism can explain the  
620 higher efficiency of the sun photochemical treatment with respect to the UVB lamp in enhancing  
621 flavonols in waste cabbage leaves.

2 Additionally, because of the large number of evidences in favor to the antioxidant role of  
3 dihydroxy-B-ring substituted flavonoids (quercetin, luteolin) [59], it is possible that their  
4 biosynthesis be regulated by excess light-induced oxidative damage, irrespective of the proportion  
5 of solar wavelengths reaching the leaf [60].



626 Since the irradiation treatment had similar effects on stalk-attached and detached leaves, we can  
 627 assume that the processes depicted in Fig. 6 occur both in on-plant leaves and postharvest. Indeed,  
 628 flavonols were increased by an UVB lamp treatment in detached Ginkgo biloba leaves [61]. Yet,  
 629 the UVB-induced rise of the CHS and PAL enzyme activity was observed in detached basil leaves  
 630 [62] and harvested carrot roots [63], respectively.



632  
 633  
 634 Figure 6. Schematic representation of the photochemically-induced mechanism of flavonols

635 accumulation in the waste cabbage leaves. Abbreviations: UVR8: UV RESISTANCE LOCUS 8;

636 CROP1: CONSTITUTIVE PHOTOMORPHOGENIC 1; CRYs: Cryptochromes; PAL:

7 phenylalanine ammonium lyase; C4H: cinnamate-4-hydroxylase; 4CL: 4-coumarate:CoA ligase;

8 CHS: chalcone synthase; CHI: chalcone isomerase; F3H: flavanone 3-hydroxylase; F3'H: flavonoid

9 3'-hydroxylase; FLS: flavonol synthase; UGT: UDP-dependent glycosyltransferase. The size of the

0 white arrows indicate the extent of the increase in flavonols: higher in the AD and lower in the AB

641 leaf sides. The image refers to a cross section of a wheat leaf acquired by an epifluorescence  
642 microscope.

#### 645 4. Conclusions

646 The present work indicated that exposing the left-over cabbage leaves after harvest of heads to  
647 sunlight in the field may be proposed as the sustainable method to improve the content of flavonols.

648 Treatments of waste leaves by sunlight or by using an artificial source of UV (40% UVB and 60%  
649 UVA) radiation gave similar results regarding the increase in the total flavonols content.

650 The sunlight exposure at the end of October in Skierniewice (Poland) yielded an increase of 82% in  
651 total leaf flavonols, resulting in a significant valorization of cabbage waste. Certainly, the sun  
652 photochemical treatment markedly depend on the weather condition at harvest time, nevertheless it  
653 has been proved to be more efficient than the UVB lamp treatment, because of the additional  
654 contribution from UVA and blue light, and preferred for its sustainability.

655 Once the protocols of the treatment will be optimized for its application at an industrial level, we  
656 believe that this method can be useful as a novel and alternative management of vegetable waste for  
657 the recovery of beneficial phytochemicals.

658 Our study also confirmed the usefulness of a non-destructive device for a rapid control of flavonols  
659 accumulation during the treatments. It was fundamental to measure the rates of accumulation and  
660 compare different treatments on the very same leaf, no possible with destructive analyses, limiting  
661 the biological variability and giving more precise evaluation of the effects.

662 A significant result of the optical monitoring consisted in proving the induction of the flavonols  
3 synthesis in the abaxial unirradiated leaf tissue due to a signaling effect by systemic translocation of  
4 macromolecules originating from the adaxial irradiated leaf side.

5 Measuring the evolution of the FLAV index on the very same leaf samples allowed to evaluate the  
6 correlation between flavonols increase and irradiance.





667 The sensor can be also used for the optical sampling of leaf waste to select that with the highest  
668 content in bioactive compounds.

669

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687

## 688 **Conflict Of Interest**

9 The authors declare no conflict of interest.

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## 1 **Appendix A. Supplementary Data**

2 Supplementary data to this article can be find online at....

694 **References**

- 695 [1] S. Ben-Othman, I. Jõudu, R. Bhat, Bioactives from agri-food wastes: Present insights and  
696 future challenges, *Molecules*. 25 (2020) 1–34. <https://doi.org/10.3390/molecules25030510>.
- 697 [2] A. Saini, P.S. Panesar, M.B. Bera, Valorization of fruits and vegetables waste through green  
698 extraction of bioactive compounds and their nanoemulsions-based delivery system,  
699 *Bioresour. Bioprocess*. 6 (2019). <https://doi.org/10.1186/s40643-019-0261-9>.
- 700 [3] S. Parameshwari, Impact of food waste and its effect on environment, *Int. J. Food Sci. Nutr*.  
701 2 (2017) 184–187. <https://doi.org/2017>.
- 702 [4] N. Jiménez-Moreno, I. Esparza, F. Bimbela, L.M. Gandía, C. Ancín-Azpilicueta,  
703 Valorization of selected fruit and vegetable wastes as bioactive compounds: Opportunities  
704 and challenges, *Crit. Rev. Environ. Sci. Technol*. 50 (2020) 2061–2108.  
705 <https://doi.org/10.1080/10643389.2019.1694819>.
- 706 [5] S. Neugart, S. Baldermann, F.S. Hanschen, R. Klopsch, M. Wiesner-Reinhold, M. Schreiner,  
707 The intrinsic quality of brassicaceous vegetables: How secondary plant metabolites are  
708 affected by genetic, environmental, and agronomic factors, *Sci. Hortic. (Amsterdam)*. 233  
709 (2018) 460–478. <https://doi.org/10.1016/j.scienta.2017.12.038>.
- 710 [6] E. Berndtsson, R. Andersson, E. Johansson, M.E. Olsson, Side streams of broccoli leaves: A  
711 climate smart and healthy food ingredient, *Int. J. Environ. Res. Public Health*. 17 (2020).  
712 <https://doi.org/10.3390/ijerph17072406>.
- 713 [7] R. Kosson, K. Felczyński, J. Szwejda-Grzybowska, M. Grzegorzewska, L. Tuccio, G. Agati,  
714 S. Kaniszewski, Nutritive value of marketable heads and outer leaves of white head cabbage  
5 cultivated at different nitrogen rates, *Acta Agric. Scand. Sect. B Soil Plant Sci*. 67 (2017)  
6 524–533. <https://doi.org/10.1080/09064710.2017.1308006>.
- 7 [8] FAOSTAT, <Http://Www.Fao.Org/Faostat/En/>, Access: 27.08.2020. (2020).
- 8 [9] M. Liu, L. Zhang, S.L. Ser, J.R. Cumming, K.M. Ku, Comparative phytonutrient analysis of

- 719 broccoli by-products: The potentials for broccoli by-product utilization, *Molecules*. 23  
720 (2018). <https://doi.org/10.3390/molecules23040900>.
- 721 [10] A.C. Formica-Oliveira, G.B. Martínez-Hernández, V. Díaz-López, F. Artés, F. Artés-  
722 Hernández, Use of postharvest UV-B and UV-C radiation treatments to revalorize broccoli  
723 byproducts and edible florets, *Innov. Food Sci. Emerg. Technol.* 43 (2017) 77–83.  
724 <https://doi.org/10.1016/j.ifset.2017.07.036>.
- 725 [11] Y. Tanongkankit, N. Chiewchan, S. Devahastin, Evolution of antioxidants in dietary fiber  
726 powder produced from white cabbage outer leaves: effects of blanching and drying methods,  
727 *J. Food Sci. Technol.* 52 (2015) 2280–2287. <https://doi.org/10.1007/s13197-013-1203-8>.
- 728 [12] T. Prokopov, Z. Goranova, M. Baeva, A. Slavov, C.M. Galanakis, Effects of powder from  
729 white cabbage outer leaves on sponge cake quality, *Int. Agrophysics*. 29 (2015) 493–500.  
730 <https://doi.org/10.1515/intag-2015-0055>.
- 731 [13] D. Kołodziejki, A. Piekarska, F.S. Hanschen, T. Pilipczuk, F. Tietz, B. Kuszniereicz, A.  
732 Bartoszek, Relationship between conversion rate of glucosinolates to isothiocyanates/indoles  
733 and genotoxicity of individual parts of Brassica vegetables, *Eur. Food Res. Technol.* 245  
734 (2019) 383–400. <https://doi.org/10.1007/s00217-018-3170-9>.
- 735 [14] T. Amofa-Diatuo, D.M. Anang, F.J. Barba, B.K. Tiwari, Development of new apple  
736 beverages rich in isothiocyanates by using extracts obtained from ultrasound-treated  
737 cauliflower by-products: Evaluation of physical properties and consumer acceptance, *J. Food*  
738 *Compos. Anal.* 61 (2017) 73–81. <https://doi.org/10.1016/j.jfca.2016.10.001>.
- 739 [15] G. Agati, L. Tuccio, B. Kuszniereicz, T. Chmiel, A. Bartoszek, A. Kowalski, M.  
740 Grzegorzewska, R. Kosson, S. Kaniszewski, Nondestructive Optical Sensing of Flavonols  
1 and Chlorophyll in White Head Cabbage (*Brassica oleracea* L. var. capitata subvar. alba)  
2 Grown under Different Nitrogen Regimens, *J. Agric. Food Chem.* 64 (2016) 85–94.  
3 <https://doi.org/10.1021/acs.jafc.5b04962>.
- 4 [16] G.B. Gonzales, K. Raes, S. Coelus, K. Struijs, G. Smaghe, J. Van Camp, Ultra(high)-

- 745 pressure liquid chromatography-electrospray ionization-time-of-flight-ion mobility-high  
746 definition mass spectrometry for the rapid identification and structural characterization of  
747 flavonoid glycosides from cauliflower waste, *J. Chromatogr. A.* 1323 (2014) 39–48.  
748 <https://doi.org/10.1016/j.chroma.2013.10.077>.
- 749 [17] A.Y. Chen, Y.C. Chen, A review of the dietary flavonoid, kaempferol on human health and  
750 cancer chemoprevention, *Food Chem.* 138 (2013) 2099–2107.  
751 <https://doi.org/10.1016/j.foodchem.2012.11.139>.
- 752 [18] M.-H. Pan, C.-S. Lai, C.-T. Ho, Anti-inflammatory activity of natural dietary flavonoids,  
753 *Food Funct.* 1 (2010) 15–31. <https://doi.org/10.1039/C0FO00103A>.
- 754 [19] M.A. Schreiner, M., Mewis, I., Neugart, S., Zrenner, R., Glaab, J., Wiesner, M., Jansen, UV-  
755 B elicitation of secondary plant metabolites, in: M. Kneissl, J. Rass (Eds.), *III-Nitride Ultrav.*  
756 *Emit.*, Springer International Publishing, Cham, Switzerland, 2016: pp. 387–414.
- 757 [20] M. Schreiner, S. Huyskens-Keil, Phytochemicals in fruit and vegetables: health promotion  
758 and postharvest elicitors, *CRC. Crit. Rev. Plant Sci.* 25 (2006) 267--278.
- 759 [21] B. Harbaum-Piayda, K. Palani, K. Schwarz, Influence of postharvest UV-B treatment and  
760 fermentation on secondary plant compounds in white cabbage leaves, *Food Chem.* 197  
761 (2016) 47–56. <https://doi.org/10.1016/j.foodchem.2015.10.065>.
- 762 [22] G. Agati, Z.G. Cerovic, P. Pinelli, M. Tattini, Light-induced accumulation of ortho-  
763 dihydroxylated flavonoids as non-destructively monitored by chlorophyll fluorescence  
764 excitation techniques, *Environ. Exp. Bot.* 73 (2011) 3–9.  
765 <https://doi.org/10.1016/j.envexpbot.2010.10.002>.
- 766 [23] R. Julkunen-Tiitto, N. Nenadis, S. Neugart, M. Robson, G. Agati, J. Vepsäläinen, G. Zipoli,  
7 L. Nybakken, B. Winkler, M.A.K. Jansen, Assessing the response of plant flavonoids to UV  
8 radiation: an overview of appropriate techniques, *Phytochem. Rev.* 14 (2015) 273–297.  
9 <https://doi.org/10.1007/s11101-014-9362-4>.
- 0 [24] S.F. Hagen, G.I.A. Borge, K.A. Solhaug, G.B. Bengtsson, Effect of cold storage and harvest



- 771 date on bioactive compounds in curly kale (*Brassica oleracea* L. var. *acephala*), *Postharvest*  
772 *Biol. Technol.* 51 (2009) 36–42. <https://doi.org/10.1016/j.postharvbio.2008.04.001>.
- 773 [25] G.B. Bengtsson, R. Schöner, E. Lombardo, J. Schöner, G.I.A. Borge, W. Bilger, Chlorophyll  
774 fluorescence for non-destructive measurement of flavonoids in broccoli, *Postharvest Biol.*  
775 *Technol.* 39 (2006) 291–298. <https://doi.org/10.1016/j.postharvbio.2005.11.003>.
- 776 [26] A. Rybarczyk-Plonska, A.B. Wold, G.B. Bengtsson, G.I.A. Borge, M.K. Hansen, S.F.  
777 Hagen, Flavonols in broccoli (*Brassica oleracea* L. var. *italica*) flower buds as affected by  
778 postharvest temperature and radiation treatments, *Postharvest Biol. Technol.* 116 (2016)  
779 105–114. <https://doi.org/10.1016/j.postharvbio.2015.12.023>.
- 780 [27] Y. Goulas, Z.G. Cerovic, A. Cartelat, I. Moya, Dualex: A new instrument for field  
781 measurements of epidermal ultraviolet absorbance by chlorophyll fluorescence, *Appl. Opt.*  
782 43 (2004) 4488–4496. <https://doi.org/10.1364/AO.43.004488>.
- 783 [28] A.F. McKinlay, Reference action spectrum for ultraviolet induced erythema in human skin,  
784 *CIE-Journal.* (1987).
- 785 [29] M.M. Caldwell, Solar UV irradiation and the growth and development of higher plants, in:  
786 *Photophysiology*, 1971. <https://doi.org/10.1016/b978-0-12-282606-1.50010-6>.
- 787 [30] M. Ibdah, A. Krins, H.K. Seidlitz, W. Heller, D. Strack, T. Vogt, Spectral dependence of  
788 flavonol and betacyanin accumulation in *Mesembryanthemum crystallinum* under enhanced  
789 ultraviolet radiation, *Plant, Cell Environ.* 25 (2002) 1145–1154.  
790 <https://doi.org/10.1046/j.1365-3040.2002.00895.x>.
- 791 [31] A. Ruggaber, R. Dlugi, T. Nakajima, Modelling radiation quantities and photolysis  
792 frequencies in the troposphere, *J. Atmos. Chem.* 18 (1994) 171–210.  
3 <https://doi.org/10.1007/BF00696813>.
- 4 [32] D. Grifoni, G. Zipoli, F. Sabatini, G. Messeri, L. Bacci, Action spectra affect variability of  
5 the climatology of biologically effective ultraviolet radiation on cloud-free days, *Radiat. Prot.*  
6 *Dosimetry.* 157 (2013) 491–498. <https://doi.org/10.1093/rpd/nct178>.

- 797 [33] A. Castagna, E. Chiavaro, C. Dall'Asta, M. Rinaldi, G. Galaverna, A. Ranieri, Effect of  
798 postharvest UV-B irradiation on nutraceutical quality and physical properties of tomato  
799 fruits, *Food Chem.* 137 (2013) 151–158. <https://doi.org/10.1016/j.foodchem.2012.09.095>.
- 800 [34] A. Castagna, C. Dall'Asta, E. Chiavaro, G. Galaverna, A. Ranieri, Effect of post-harvest UV-  
801 B irradiation on polyphenol profile and antioxidant activity in flesh and peel of tomato fruits,  
802 *Food Bioprocess Technol.* 7 (2014) 2241–2250. <https://doi.org/10.1007/s11947-013-1214-5>.
- 803 [35] V.E. Ruiz, R. Interdonato, L. Cerioni, P. Albornoz, J. Ramallo, F.E. Prado, M. Hilal, V.A.  
804 Rapisarda, Short-term UV-B exposure induces metabolic and anatomical changes in peel of  
805 harvested lemons contributing in fruit protection against green mold, *J. Photochem.*  
806 *Photobiol. B Biol.* 159 (2016) 59–65. <https://doi.org/10.1016/j.jphotobiol.2016.03.016>.
- 807 [36] K. Kanazawa, T. Hashimoto, S. Yoshida, P. Sungwon, S. Fukuda, Short photoirradiation  
808 induces flavonoid synthesis and increases its production in postharvest vegetables, *J. Agric.*  
809 *Food Chem.* 60 (2012) 4359–4368. <https://doi.org/10.1021/jf300107s>.
- 810 [37] L.B.D.S. Nascimento, C. Brunetti, G. Agati, C. Lo Iacono, C. Detti, E. Giordani, F. Ferrini,  
811 A. Gori, Short-term pre-harvest UV-B supplement enhances the polyphenol content and  
812 antioxidant capacity of ocimum basilicum leaves during storage, *Plants.* 9 (2020) 1–19.  
813 <https://doi.org/10.3390/plants9060797>.
- 814 [38] J.D. Liu, D. Goodspeed, Z. Sheng, B. Li, Y. Yang, D.J. Kliebenstein, J. Braam, Keeping the  
815 rhythm: Light/dark cycles during postharvest storage preserve the tissue integrity and  
816 nutritional content of leafy plants, *BMC Plant Biol.* 15 (2015) 1–9.  
817 <https://doi.org/10.1186/s12870-015-0474-9>.
- 818 [39] S. Neugart, M. Schreiner, UVB and UVA as eustressors in horticultural and agricultural  
9 crops, *Sci. Hortic. (Amsterdam).* 234 (2018) 370–381.  
0 <https://doi.org/10.1016/j.scienta.2018.02.021>.
- 1 [40] N. Rai, S. Neugart, Y. Yan, F. Wang, S.M. Siipola, A. V. Lindfors, J.B. Winkler, A. Albert,  
2 M. Brosché, T. Lehto, L.O. Morales, P.J. Aphalo, How do cryptochromes and UVR8 interact

- 823 in natural and simulated sunlight?, *J. Exp. Bot.* 70 (2019) 4975–4990.  
824 <https://doi.org/10.1093/jxb/erz236>.
- 825 [41] S.M. Siipola, T. Kotilainen, N. Sipari, L.O. Morales, A. V. Lindfors, T.M. Robson, P.J.  
826 Aphalo, Epidermal UV-A absorbance and whole-leaf flavonoid composition in pea respond  
827 more to solar blue light than to solar UV radiation, *Plant, Cell Environ.* 38 (2015) 941–952.  
828 <https://doi.org/10.1111/pce.12403>.
- 829 [42] P.M.A. Toivonen, D.M. Hodges, Abiotic Stress in Harvested Fruits and Vegetables, in: A.K.  
830 Shanker, B. Venkateswarlu (Eds.), *Abiotic Stress Plants-Mechanisms Adapt.*, IntechOpen,  
831 Rijka, Croatia, 2011: pp. 39–58. <https://doi.org/10.1016/j.colsurfa.2011.12.014>.
- 832 [43] F.L. Reyes, J. Emilio Villarreal, L. Cisneros-Zevallos, The increase in antioxidant capacity  
833 after wounding depends on the type of fruit or vegetable tissue, *Food Chem.* 101 (2007)  
834 1254–1262. <https://doi.org/10.1016/j.foodchem.2006.03.032>.
- 835 [44] Z. Balouchi, G. Peyvast, M. Ghasemnezhad, M. Saadatian, Changes of antioxidant  
836 compounds of broccoli ( *Brassica oleracea L . var . Italica* ) during storage at low and high  
837 temperatures, *South West. J. Hortic. Biol. Environ.* 2 (2011) 193–212.
- 838 [45] A. Starzyńska, M. Leja, A. Mareczek, Physiological changes in the antioxidant system of  
839 broccoli flower buds senescing during short-term storage, related to temperature and  
840 packaging, *Plant Sci.* 165 (2003) 1387–1395. <https://doi.org/10.1016/j.plantsci.2003.07.004>.
- 841 [46] J.H. Hasperué, A.R. Chaves, G.A. Martínez, End of day harvest delays postharvest  
842 senescence of broccoli florets, *Postharvest Biol. Technol.* 59 (2011) 64–70.  
843 <https://doi.org/10.1016/j.postharvbio.2010.08.005>.
- 844 [47] M. Leja, I. Kamińska, A. Kołton, Phenolic compounds as the major antioxidants in red  
5 cabbage, *Folia Hortic.* 22 (2010) 19–24. <https://doi.org/10.2478/fhort-2013-0146>.
- 6 [48] C. Brunetti, L. Guidi, F. Sebastiani, M. Tattini, Isoprenoids and phenylpropanoids are key  
7 components of the antioxidant defense system of plants facing severe excess light stress,  
8 *Environ. Exp. Bot.* 119 (2015) 54–62. <https://doi.org/10.1016/j.envexpbot.2015.04.007>.

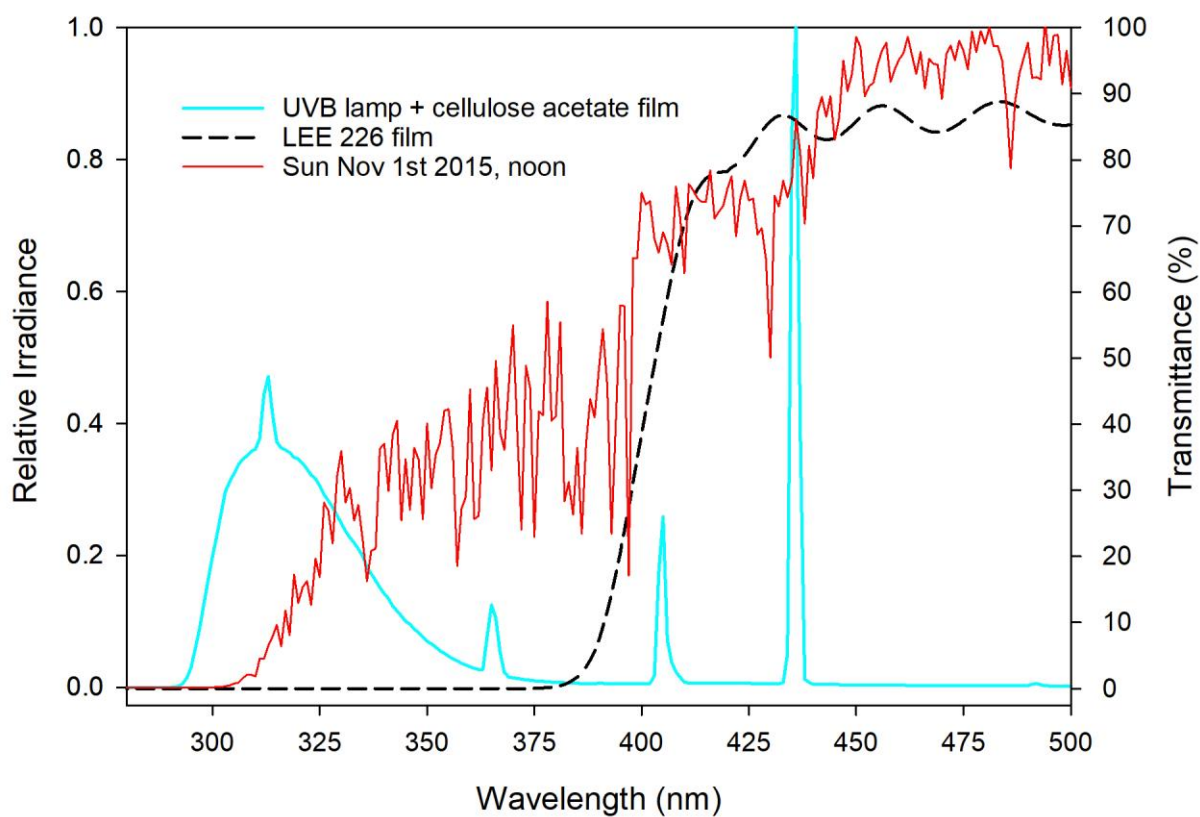


- 849 [49] A. Fini, C. Brunetti, M. Di Ferdinando, F. Ferrini, M. Tattini, Stress-induced flavonoid  
850 biosynthesis and the antioxidant machinery of plants, *Plant Signal. Behav.* 6 (2011) 709–711.  
851 <https://doi.org/10.4161/psb.6.5.15069>.
- 852 [50] K. Tilbrook, A.B. Arongaus, M. Binkert, M. Heijde, R. Yin, R. Ulm, The UVR8 UV-B  
853 photoreceptor: perception, signaling and response, *Arab. B.* 11 (2013) e0164.  
854 <https://doi.org/10.1199/tab.0164>.
- 855 [51] D.H. Chitwood, M.C.P. Timmermans, Small RNAs are on the move, *Nature.* 467 (2010)  
856 415–419. <https://doi.org/10.1038/nature09351>.
- 857 [52] L.P.R. Bidel, G. Chomicki, F. Bonini, L. Mondolot, J. Soulé, M. Coumans, P. La Fisca, Y.  
858 Baissac, V. Petit, A. Loiseau, Z.G. Cerovic, K.S. Gould, C. Jay-Allemand, Dynamics of  
859 flavonol accumulation in leaf tissues under different UV-B regimes in *Centella asiatica*  
860 (*Apiaceae*), *Planta.* 242 (2015) 545–559. <https://doi.org/10.1007/s00425-015-2291-7>.
- 861 [53] E. Marín-González, P. Suárez-López, “And yet it moves”: Cell-to-cell and long-distance  
862 signaling by plant microRNAs, *Plant Sci.* 196 (2012) 18–30.  
863 <https://doi.org/10.1016/j.plantsci.2012.07.009>.
- 864 [54] C.M. Ålenius, T.C. Vogelmann, J.F. Bornman, A three- dimensional representation of the  
865 relationship between penetration of u.v.- B radiation and u.v.- screening pigments in leaves  
866 of *Brassica napus*, *New Phytol.* 131 (1995) 297–302. <https://doi.org/10.1111/j.1469-8137.1995.tb03065.x>.
- 867  
868 [55] D. Verdaguer, M.A.K. Jansen, L. Llorens, L.O. Morales, S. Neugart, UV-A radiation effects  
869 on higher plants: Exploring the known unknown, *Plant Sci.* 255 (2017) 72–81.  
870 <https://doi.org/10.1016/j.plantsci.2016.11.014>.
- 1 [56] D. Grifoni, G. Agati, F. Bussotti, M. Michelozzi, M. Pollastrini, G. Zipoli, Different  
2 responses of *Arbutus unedo* and *Vitis vinifera* leaves to UV filtration and subsequent  
3 exposure to solar radiation, *Environ. Exp. Bot.* 128 (2016) 1–10.  
4 <https://doi.org/10.1016/j.envexpbot.2016.03.012>.





- 875 [57] C.A. Kolb, M.A. Kaser, J. Kopecky, G. Zotz, M. Riederer, E.E. Pfundel, Effects of natural  
876 intensities of visible and ultraviolet radiation on, *Plant Physiol.* 127 (2001) 863–875.  
877 <https://doi.org/10.1104/pp.010373.research>.
- 878 [58] L.O. Morales, R. Tegelberg, M. Brosché, M. Keinänen, A. Lindfors, P.J. Aphalo, Effects of  
879 solar UV-A and UV-B radiation on gene expression and phenolic accumulation in *Betula*  
880 *pendula* leaves, *Tree Physiol.* 30 (2010) 923–934. <https://doi.org/10.1093/treephys/tpq051>.
- 881 [59] G. Agati, C. Brunetti, A. Fini, A. Gori, L. Guidi, M. Landi, F. Sebastiani, M. Tattini, Are  
882 flavonoids effective antioxidants in plants? Twenty years of our investigation, *Antioxidants.*  
883 9 (2020) 1–17. <https://doi.org/10.3390/antiox9111098>.
- 884 [60] G. Agati, M. Tattini, Multiple functional roles of flavonoids in photoprotection, *New Phytol.*  
885 186 (2010) 786–793. <https://doi.org/10.1111/j.1469-8137.2010.03269.x>.
- 886 [61] M. Sun, X. Gu, H. Fu, L. Zhang, R. Chen, L. Cui, L. Zheng, D. Zhang, J. Tian, Change of  
887 secondary metabolites in leaves of *Ginkgo biloba* L. in response to UV-B induction, *Innov.*  
888 *Food Sci. Emerg. Technol.* 11 (2010) 672–676. <https://doi.org/10.1016/j.ifset.2010.08.006>.
- 889 [62] A. Ghasemzadeh, S. Ashkani, A. Baghdadi, A. Pazoki, H.Z.E. Jaafar, A. Rahmat,  
890 Improvement in flavonoids and phenolic acids production and pharmaceutical quality of  
891 sweet basil (*Ocimum basilicum* L.) by ultraviolet-B irradiation, *Molecules.* 21 (2016).  
892 <https://doi.org/10.3390/molecules21091203>.
- 893 [63] R.J. Avena-Bustillos, W.X. Du, R. Woods, D. Olson, A.P. Breksa, T.H. Mchugh, Ultraviolet-  
894 B light treatment increases antioxidant capacity of carrot products, *J. Sci. Food Agric.* 92  
895 (2012) 2341–2348. <https://doi.org/10.1002/jsfa.5635>.



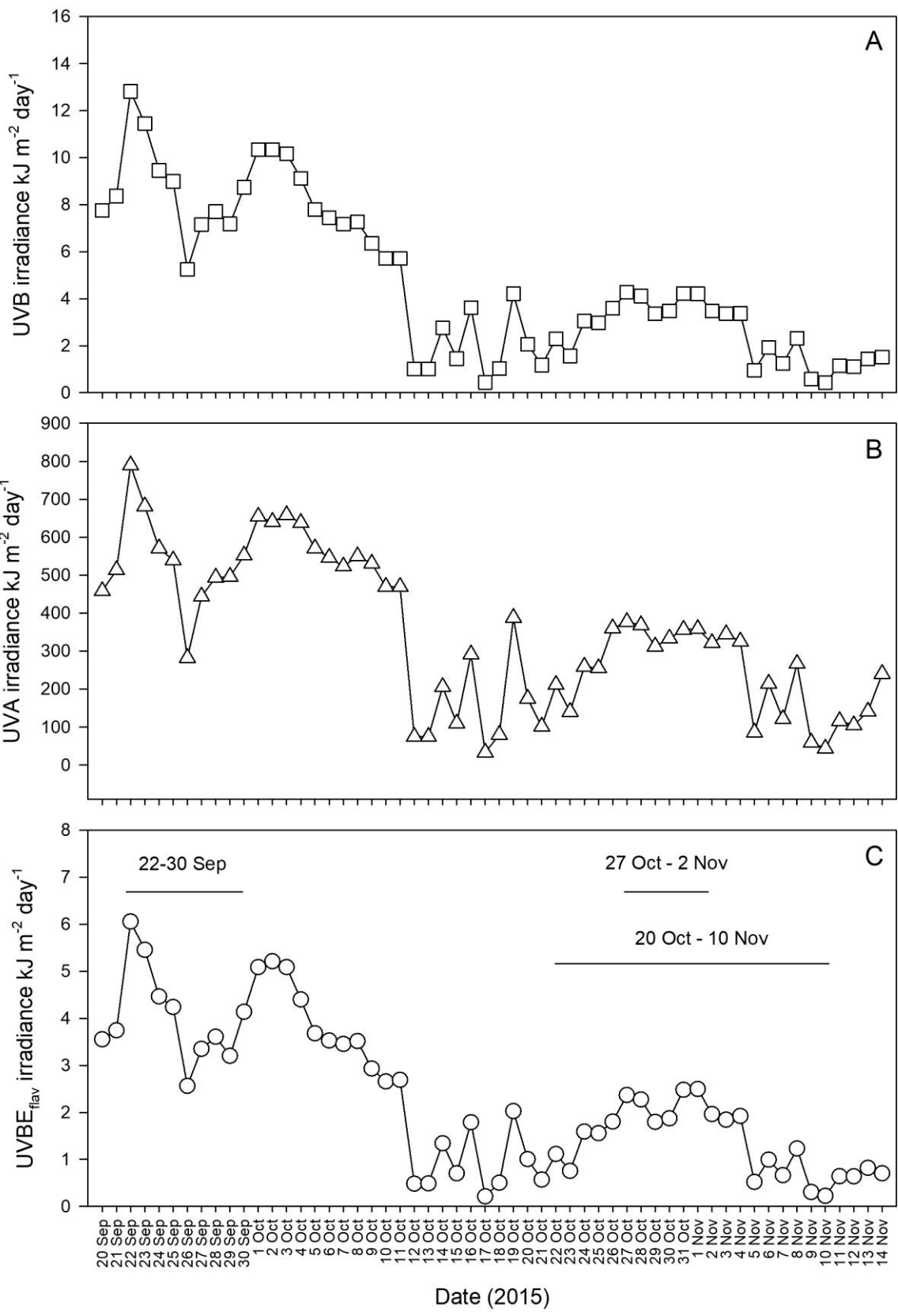
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Figure S1. Emission spectrum of a UVB lamp filtered by the cellulose acetate film and transmittance spectrum of the LEE 226 film. The solar emission spectrum recorded on Nov 1<sup>st</sup> 2015 is also reported. Emission spectra are normalized to their maximum.



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Figure S2. Postharvest sunlight exposure of cabbage leaves, attached to the stalk (top) or detached (bottom), and monitored by the Dualex sensor on the leaf apical and basal parts (Sep-Nov, 2015).



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Figure S3. Average daily solar UVB (A) and UVA (B) irradiance and the biologically effective UVBE<sub>flav</sub> dose calculated using the flavonol induction in *M. crystallinum* weighting function during the Sep-Nov 2015 period.