

# 1 Towards azeotropic MeOH-MTBE separation using pervaporation chitosan-based 2 deep eutectic solvent membranes

3  
4 Roberto Castro-Muñoz<sup>1,2\*</sup>, Asma Msahel<sup>3,4</sup>, Francesco Galiano<sup>5\*</sup>, Marcin Serocki<sup>6</sup>,  
5 Jacek Ryl<sup>7</sup>, Sofiane Ben Hamouda<sup>3,8</sup>, Amor Hafiane<sup>3</sup>, Grzegorz Boczkaj<sup>1\*</sup>, Alberto  
6 Figoli<sup>5</sup>

7  
8 <sup>1</sup>Gdansk University of Technology, Faculty of Chemistry, Department of Process Engineering and  
9 Chemical Technology, 11/12 Narutowicza St., 80-233, Gdansk, Poland

10 <sup>2</sup>Tecnologico de Monterrey, Campus Toluca. Av. Eduardo Monroy Cárdenas 2000 San Antonio  
11 Buenavista, 50110, Toluca de Lerdo, Mexico.

12 <sup>3</sup>Laboratory of Water Membrane and Environmental Biotechnology (LMBE), CERTE BP 273, 8020  
13 Soliman, Tunisia

14 <sup>4</sup>Department of Chemistry, University of Tunis El-Manar, Farhat Hached University Campus, BP n° 94  
15 Rommana, 1068 Tunis, Tunisia

16 <sup>5</sup>Institute on Membrane Technology, ITM-CNR, Via P. Bucci 17/c, 87036 Arcavacata di Rende (CS), Italy

17 <sup>6</sup>Department of Pharmaceutical Technology and Biochemistry, Faculty of Chemistry, Gdansk University of  
18 Technology, 11/12 Narutowicza St., 80-233, Gdansk, Poland

19 <sup>7</sup>Department of Electrochemistry, Corrosion and Materials Engineering, Faculty of Chemistry, Gdansk  
20 University of Technology, 11/12 Narutowicza St., 80-233, Gdansk, Poland

21 <sup>8</sup>NANOMISENE Laboratory, LR16CRMN01, Centre for Research on Microelectronics and  
22 Nanotechnology (CRMN) of Technopole of Sousse B. P334, 4054 Sahloul Sousse, Tunisia

23  
24 \*E-mail: [food.biotechnology88@gmail.com](mailto:food.biotechnology88@gmail.com) ; [castromr@tec.mx](mailto:castromr@tec.mx) (R. Castro-Munoz)  
25 [grzegorz.boczkaj@pg.edu.pl](mailto:grzegorz.boczkaj@pg.edu.pl) (Grzegorz Boczkaj) ; [f.galiano@itm.cnr.it](mailto:f.galiano@itm.cnr.it) (F. Galiano)

26 -----

27 \*Corresponding Author

28

29 **Abstract**

30 Deep eutectic solvents (DESs) are a new class of solvents that can offset some of the  
31 major drawbacks of common solvents and ionic liquids. When dealing with the  
32 preparation of dense membranes, the use of DESs is still challenging due to their low  
33 compatibility with the polymer phase. In this research, a novel L-proline:sulfolane (molar  
34 ratio 1:2) DES was synthesized and used for the preparation of more sustainable bio-  
35 based membranes using chitosan (CS) as a polymer phase. The compatibility among  
36 both phases (polymer and DESs) and their ability to form homogenous membranes was  
37 preliminary studied. In this regard, scanning electron and confocal microscopies were  
38 used to completely map the structure of the resulting membranes evidencing a complete  
39 homogenous structure. The membranes were also characterized in terms of contact  
40 angle (CA), Fourier transformed infrared spectroscopy (FTIR), mechanical resistance and  
41 swelling degree (uptake). Preliminary pervaporation tests for the separation of a methanol  
42 (MeOH)- methyl *tert*-butyl ether (MTBE) azeotropic mixture were, thus, performed. In this  
43 regard, the addition of DESs provided an enhanced separation efficiency in comparison  
44 to pristine CS membranes. Thanks to the morphology and properties exhibited, the newly  
45 developed membranes can be considered as excellent bio-based candidates to be  
46 explored in other gas selective and solvent oriented membrane operations.

47

48 **Keywords:** Deep eutectic solvents; membrane preparation; chitosan; methanol/MTBE;  
49 pervaporation.

50

51

## 52 **1. Introduction**

53 According to the “*Twelve Principles of Green Chemistry*” established by Anastas and  
54 Warner [1], there is a big need of implementing green materials and processes in the  
55 manufacturing of new products. Therefore, the research community is continuously  
56 exploring the potentialities of new feedstocks to follow such principles aiming at the  
57 preservation of the environment [2]. Today, deep eutectic solvents (DESs), which are a  
58 new class of solvents, have been categorized as “green alternatives” to conventional  
59 solvents for various applications, including metal plating and coatings [3], sustainable  
60 media in organic reactions [4], extraction and separation of biologically active compounds  
61 from natural sources [5], CO<sub>2</sub> capture [6], desulfurization [7], as stationary phases for  
62 chromatography [8], enzymatic biodiesel production [9], chemical and biocatalysis [10],  
63 to mention just a few. DESs overcome most of the major drawbacks of common solvents  
64 and ionic liquids, together with several advantages, such as low toxicity, low cost, easy  
65 handling, biodegradability, biocompatibility and reusability [11]. Typically, DESs are  
66 synthesized by combining hydrogen bond acceptor (HBA), like quaternary ammonium  
67 salt, with hydrogen bond donor (HBD) compounds [3].

68 When dealing with membrane preparation, DESs have been primarily proposed and used  
69 as additives in the manufacture of DES-liquid supported membranes [12,13], with the aim  
70 of enhancing the separation properties of polymeric membranes. The superior



71 performance exhibited by DESs based membranes are generally attributed to a facilitated  
72 molecule transport and adsorption through the functional groups of DESs [14–16]. DESs  
73 have been also explored as pore forming agents in the fabrication of asymmetric  
74 membranes via phase-inversion method [17].

75 To the best of our knowledge, there are no studies employing DESs in the preparation of  
76 non-porous (well-known as dense) membranes. This is because, fundamentally, the  
77 compatibility of DESs and polymer phases, for their complete merging, is still a challenge.  
78 Therefore, in this work, we describe the successful incorporation of a specific DES (L-  
79 proline:sulfolane) into chitosan (CS) membranes surmounting one of the most common  
80 constraints related to the proper dispersion of the DES into the polymer matrix.

81 In order to achieve this goal, a hydrophilic and water-soluble DES, such as L-  
82 proline:sulfolane (molar ratio 1:2), was firstly synthesized. Afterward, its ability to form  
83 dense homogenous membranes was evaluated using CS as a continuous polymer phase.  
84 The cross-linking effect on membrane properties and performance was also evaluated  
85 using glutaraldehyde (GA). To evaluate their miscibility, the complete blending of both  
86 phases (DES and polymer) was studied by mapping the complete membrane structure.  
87 For this purpose, scanning electron microscopy (SEM) and scanning confocal electron  
88 microscopy (SCEM), were employed. Secondly, the generated membranes were also  
89 characterized in terms of contact angle (CA), Fourier transformed infrared spectroscopy  
90 (FTIR), mechanical test and swelling degree (uptake). Preliminary pervaporation tests  
91 towards the methanol (MeOH)- methyl *tert*-butyl ether (MTBE) separation have been  
92 performed in order to prove the applicability of the new synthesised membranes.

93



## 94 **2. Methodologies**

### 95 *2.1. Reactants and materials*

96 L-proline (purity  $\geq 98\%$ , Sigma Aldrich), sulfolane (purity  $\geq 99\%$ , Alfa Aesar), phosphoric  
97 acid (purity  $\geq 85\%$ , POCH S.A.) and GA (grade II, 25 wt%) were acquired and used  
98 without further purification. CS (medium molecular weight) was acquired from Sigma  
99 Aldrich. MeOH (99.8%) and MTBE (99.7%) were also purchased from Sigma-Aldrich (St.  
100 Louis, USA) and used without further purification.

101

### 102 *2.2. DES synthesis*

103 *L-proline:sulfolane*, at a molar ratio (1:2), was synthesized. Basically, 32.4 g of L-proline  
104 (0.282 mol), 67.6 g of sulfolane (0.564mol) and 705 mL of phosphoric acid 1M (0.705  
105 mol) were mixed at 1000 rpm (70°C) until obtaining a transparent solution. Lately, the  
106 excess of water was removed out by means of a rotary vacuum evaporator (Rotavapor  
107 R-300 with a V-300 vacuum pump, BUCHI).

108

### 109 *2.3. Membrane preparation*

110 CS-DES membranes were prepared via dense-film casting method and solvent  
111 evaporation. The CS dope solutions (1.5 wt.%) in acidic water solutions were prepared.  
112 Herein, an acetic acid solution (2wt.% in distilled water) was preliminarily prepared. The  
113 polymer dope solutions were stirred over 24 h (at room temperature). Later, 5 wt.% of  
114 DES (L-proline:sulfolane), with respect to CS concentration, was separately added in the  
115 respective dope solutions. The resulting mixture was stirred for 4 h before applying the  
116 *in-situ* cross-linking procedure. The latter procedure was utilized to ensure the entrapment



117 of the DES into the polymer phase, in which chemical cross-linking with GA was used  
118 [18]. Here, *in situ* cross-linking procedure was carried out by adding 100  $\mu\text{L}$  of GA and  
119 100  $\mu\text{L}$  of HCl (to speed up the reaction) to every dope. This was stirred for 15 min, cast  
120 on clean Petri dishes and then dried at room temperature for 2 days. The final appearance  
121 of membranes was of a homogeneous and transparent film with approximately 25  $\mu\text{m}$  of  
122 thickness. To sum up, the prepared membranes were labelled as follows: pristine CS,  
123 cross-linked CS (xCS), chitosan:L-proline:sulfolane (CS:PRO:SUF) and cross-linked  
124 chitosan: L-proline:sulfolane (xCS:PRO:SUF).

125

## 126 2.4. Membrane characterization

127 2.4.1. *Scanning electron microscopy (SEM)*. The morphological structure of the  
128 membrane surface and cross-section was preliminarily analysed using a SEM instrument  
129 (Hitachi S-3400N, Japan), operating with a tungsten electron source. The secondary  
130 electron detector was used for the analysis, and micrographs were made under 5 kV  
131 accelerating voltage. Before the microanalysis, a 10 nm layer of metallic gold was  
132 sputtered at each sample surface to compensate the low surface conductivity. The  
133 corresponding images were acquired at suitable magnification. In the case of cross-  
134 section analysis, all samples were prepared by cryogenic fracture after immersion in liquid  
135  $\text{N}_2$ .

136 2.4.2. *Scanning confocal electron microscopy (SCEM)*. In order to assess the  
137 homogeneity and dense structure of the resulting membranes, a complete scanning of  
138 the structure was performed using a LSM 800 T-PMT confocal microscope (Carl Zeiss  
139 AG, Oberkochen, Germany) with a CCD camera [19]. Images were acquired and



140 processed with ZEN Blue software.

141 2.4.3. *Fourier transformed infrared spectroscopy (FTIR)*. FTIR was performed on all  
142 membrane formulations described previously using a Nicolet iS10 FTIR (Thermo Fisher  
143 Scientific) spectrometer equipped with a DTGS detector and a Golden Gate diamond ATR  
144 accessory. The spectra were recorded in the 4000–400  $\text{cm}^{-1}$  wave number range at a  
145 resolution of 16  $\text{cm}^{-1}$ .

146 2.4.4. *Water contact angle (CA)*. The water contact angle measurements were performed  
147 using ultrapure water by the method of the sessile drop using a goniometer OCA15 (Data  
148 Physics). The average and standard deviation values were determined for three  
149 measurements.

150 2.4.5. *Mechanical test*. Mechanical properties of pristine CS and xCS:PRO:SUF  
151 membranes were measured using a Zwick/Roell Z2.5 test unit (BTC-FR2.5TN-D09,  
152 Germany). Measurements were performed at room temperature (25 °C) using a  
153 membrane sample of 1 × 5 cm. The samples were extended at the constant elongation  
154 rate of 5 mm  $\text{min}^{-1}$  until their break. Elongation at break, Young's modulus and tensile  
155 strength were, then, determined. Each sample was analyzed at least four times, the  
156 results were expressed as the average and standard deviation. Mechanical tests were  
157 carried out on all the investigated membranes before and after use in PV tests (14.3 wt%  
158 MeOH; 85.7 wt% MTBE).

159 2.4.6. *Uptake*. The degree of swelling (uptake) of the membranes was determined for pure  
160 feed components (MeOH, MTBE), various MeOH-MTBE mixtures (5, 10 25, 50, wt.%  
161 MeOH) as well as azeotropic mixture (14.3% MeOH and 85.7% MTBE). Three small  
162 pieces of membranes (1 × 5 cm) were weighed and immersed in the solvent mixtures at



163 30 °C for 48 h. The wet membrane samples were quickly wiped with tissue paper to  
164 remove the excess free liquid on their surface and immediately weighed with a digital  
165 balance (Gibertini, Crystal 500, Italy, Crystal 500, Gibertini Elettronica srl, Milan, Italy)  
166 with an accuracy of 0.001 g. In general, the uptake was calculated as follows:

167

$$168 \quad \text{Uptake (\%)} = \frac{W_w - W_d}{W_d} \cdot 100 \quad \text{Eq. (1)}$$

169 where  $W_w$  and  $W_d$  correspond to the weight of the wet and dry membranes, respectively.

170

### 171 2.5. Pervaporation tests

172 The PV experiments were performed in a laboratory-scale setup, whose graphical  
173 drawing and details can be found elsewhere [20]. Basically, a mixture (250 mL) of an  
174 azeotropic MeOH-MTBE (14.3–85.7 wt.%, respectively) solution was poured in the  
175 pervaporation cell. The feed operating temperature was varied (at 25, 35, 45 °C) and  
176 controlled with an accuracy of 0.01 °C using a thermo digital circulating bath (Neslab RTE-  
177 201, USA). The vacuum on the permeate side (at 0.05 mbar) was maintained by using a  
178 RV5 two-stage vacuum pump (Edwards, UK).

179 The membrane, having an active area of 9.6 cm<sup>2</sup>, was placed on a porous support within  
180 the membrane cell. The permeated vapour was condensed and collected in a glass trap  
181 placed in a liquid nitrogen condenser. Once reached the steady-state, the permeates  
182 were collected for 5 h of experiment and immediately weighted to determine the total  
183 permeate flux. The permeate flux ( $J$ ) was determined as follows:

184

$$J = \frac{Q}{A \cdot t} \quad \text{Eq. (2)}$$



185 where  $Q$  corresponds to the weight of the permeate (expressed in kg),  $A$  corresponds to  
186 the membrane area ( $m^2$ ) and  $t$  is the operating time (h). The partial flux ( $J_i$ ) for each  
187 component  $i$  was calculated by multiplying its weight fraction ( $y_i$ ) in the collected permeate  
188 sample by the total permeate flux ( $J$ ), as follows:

$$189 \quad J = Y_i \cdot J \quad \text{Eq. (3)}$$

190

191 The separation factor ( $\alpha$ ) was calculated according to the following equation:

192

$$193 \quad \alpha = \frac{y_{MeOH}/y_{MTBE}}{x_{MeOH}/x_{MTBE}} \quad \text{Eq. (4)}$$

194 where  $y$  and  $x$  correspond to the weight fractions of the components in the permeate and  
195 feed, respectively. The permeate composition was determined by an Abbe 60 type direct  
196 reading refractometer (Bellingham + Stanley Ltd., UK) at 25 °C. The  $J$  and  $\alpha$  values were  
197 expressed as the average of more than two runs to ensure the accuracy of the outcomes.

198

### 199 **3. Results and discussion**

#### 200 **3.1. Scanning electron microscopy (SEM).**

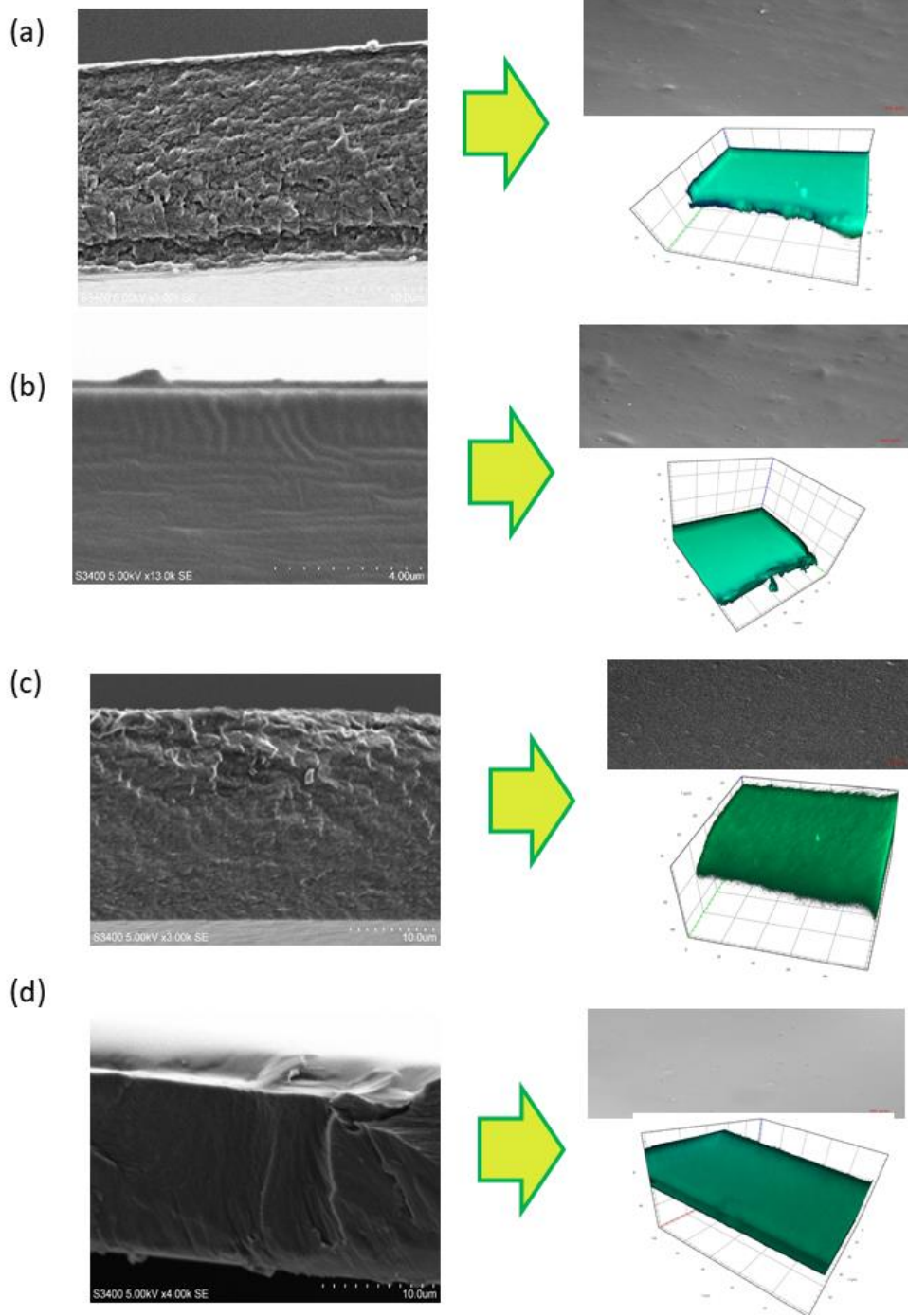
201 In general, all resulting membranes showed a smooth and uniform surface pattern without  
202 signs of plastic deformation, being common for dense polymeric membranes [21].  
203 Particular attention has been devoted to the cross-section analyses of the membranes,  
204 as shown in **Figure 1**. For instance, the pristine CS membrane (Figure 1a) displayed a  
205 crater-like pattern, which is commonly generated during deformation caused by the  
206 freeze-fracture of polymer membranes. Such a pattern has been well documented in



207 pristine [22] and cross-linked CS membranes [18]. Regarding the CS blend with  
208 PRO:SUF (Figure 1c), the membrane exhibited a more homogeneous dense pattern  
209 structure with non-visible pores. Similarly, cross-linked CS-DESs membranes (Figure 1d)  
210 confirmed a dense structure with non-visible defects among the phases and additives  
211 (like DES agent). Particularly, such characteristics can be used as an evidence of the  
212 good miscibility of the hydrophilic PRO:SUF DES in CS, as demonstrated in other  
213 polymer/polyethylene glycol blends [23,24].

214

215



**Figure 1.** SEM cross-section, surface view and 3D view images of the prepared membranes based on CS and DES. (a) pristine CS (CS), (b) cross-linked chitosan (xCS) (c) chitosan:L-proline:sulfolane (CS:PRO:SUF), (d) cross-linked chitosan:L-proline:sulfolane (xCS:PRO:SUF).

216

217

218

219

220

221 In this work, the addition of the DES has evidenced to provide a tighter and smoother  
222 structure in CS membranes. However, in literature it has been reported that DESs are  
223 employed in the fabrication of porous membranes by acting as pore forming agents  
224 [17,25]. In general, the use of pore formers is aimed, in phase-inversion techniques, to  
225 generate large pores and voids in the membrane matrix [26]. The pore formers (such as  
226 polyethylene glycol and polyvinyl pyrrolidone) are generally removed by the membranes  
227 after its formation by washing them with water.

228 In our case, the approach was oriented to physically entrap the selected DES into the  
229 membrane thus exploiting the benefits that it may offer once preserved in the membrane  
230 structure. DESs have been successfully applied for the extraction, separation and  
231 purification of biomolecules due to their ability to form hydrogen bonds via dipole-dipole  
232 and other specific solute-solvent interactions [27,28].

233 The resulting CS-based membranes visually exhibited a dense structure. However, there  
234 is still a possibility that the used DESs can also be supported (or encapsulated) within the  
235 polymeric structure. Therefore, further analysis of the overall membrane structure were  
236 performed. In this case, SCEM was used for mapping the structures of the membranes,  
237 as detailed in **Figure 1** (right side). It is known that polymer blending with other phases  
238 strongly depends on its available chemical functionalities. If functional groups are not  
239 present between phases, a poor interaction and hence low miscibility will be obtained  
240 [29]. This is also fundamental for blending polymers with inorganic phases (e.g.,  
241 nanoparticles), in which the nano-sized materials tend to be chemically functionalized to  
242 reach a good compatibility and contact between polymer and filler interfaces [30].

243 In particular, CS owns a plenty of amino and hydroxyl groups, making it an excellent



244 candidate for polymer blending [31]. Also, in the case of DESs, they may also offer a  
245 series of functional groups which can interact with CS giving to the final membrane  
246 specific properties. At this point, it is likely that this new DES employed (PRO:SUF),  
247 bearing different functional groups (such as amino, hydroxyl, and carbonyl), can interact  
248 with CS. For instance, **Figure 1** shows the surface view of the synthesized membranes  
249 confirming their smooth and homogenous surfaces with absence of pinholes and defects.  
250 Regarding the 3D view, the images were created by stacking together 210 fluorescence  
251 intensity scans of each membrane. Every single scan is a picture representation of a 0.25  
252  $\mu\text{m}$  cross-section of each membrane, each scan is created by merging green and blue  
253 from two fluorescence channels (DAPI and AF488, excitation: 353 and 493 nm, emission  
254 465 and 517 nm, detection 400-580 nm). By scanning the membrane structure, the  
255 complete blending between CS and DES was confirmed, evidencing a dense-like  
256 morphology. Also, there was no evidence of DES phase separation or encapsulation in  
257 the polymer matrix.

258  
259

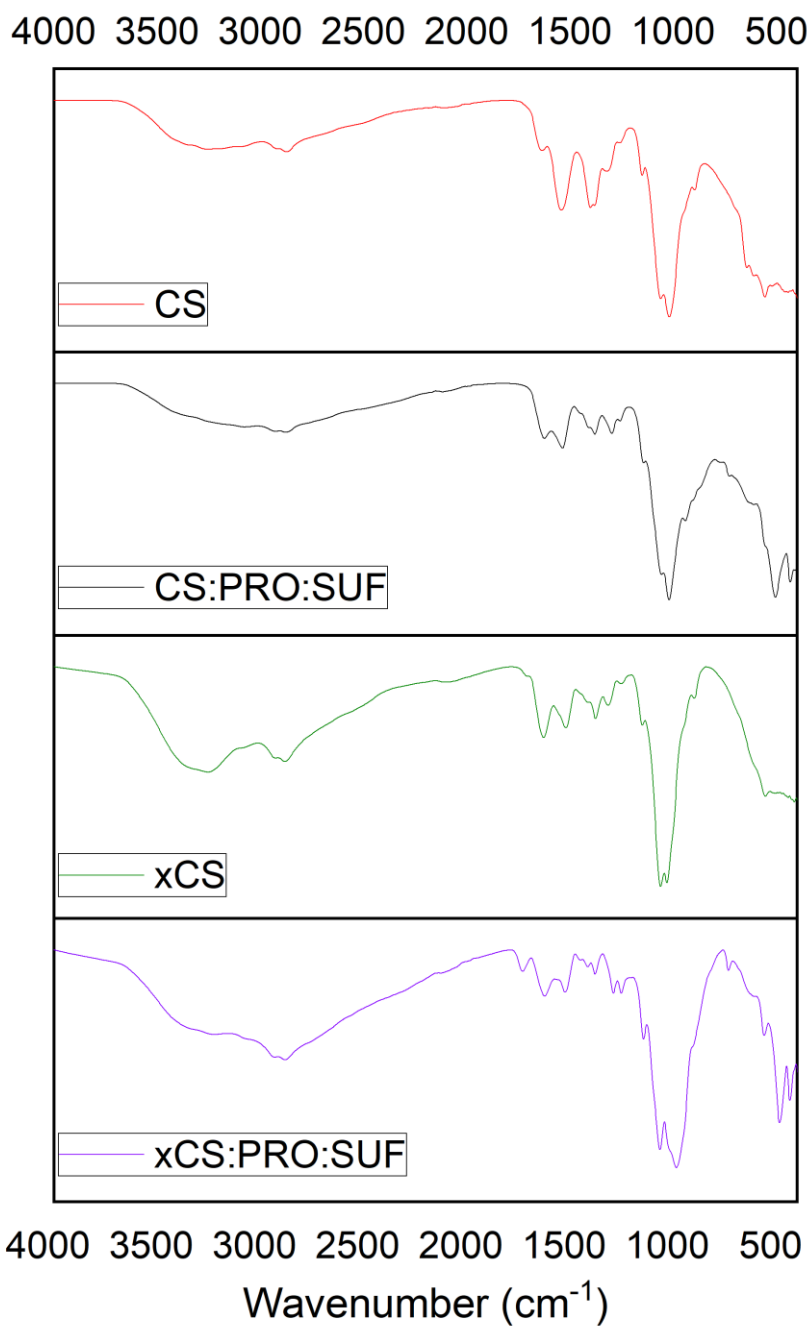
### 260 3.2. *Water contact angle (CA) and Fourier transformed infrared spectroscopy* 261 *(FTIR)*

262 The FTIR results (shown in Figure 2) can be the evidence of the affective blending of the  
263 DES into the CS polymer membrane matrix. All spectra exhibit a strong and broad non-  
264 symmetric band at about  $3400\text{ cm}^{-1}$  that results from overlapping of the O-H and N-H  
265 stretching vibrations of functional groups engaged in hydrogen bonds. The spectrum of  
266 CS shows typical absorption bands at  $1600\text{ cm}^{-1}$  (for C=O stretching in amide group),



267 1550  $\text{cm}^{-1}$  (for N-H bending in no acetylated 2-aminoglucose) and 1560  $\text{cm}^{-1}$  (for N-H  
268 bending in amide group), as reported in literature [32]. Absorption bands at 1100  $\text{cm}^{-1}$ ,  
269 corresponding to an antisymmetric stretching of the C-O-C bridge, 1050  $\text{cm}^{-1}$  and 1000  
270  $\text{cm}^{-1}$ , related to skeletal vibrations implying the C-O stretching, are typical for CS.  
271 Interestingly, the blending of CS with DES provoked a slight shift on such characteristic  
272 polymer bands, giving a proof of the good interaction among the phases and thus  
273 compatibility. For the cross-linked CS-DES membranes with GA, they exhibit an increase  
274 in the absorption between 1600-1650  $\text{cm}^{-1}$  which can be attributed to imine bonds N=C  
275 [33,34]. The stretching at 1540  $\text{cm}^{-1}$ , 1710  $\text{cm}^{-1}$  and 2900  $\text{cm}^{-1}$  corresponds to free-  
276 aldehydic bonds and increased C-H stretch, respectively. Also, the presence of aliphatic  
277 amino groups diminishes as much as the peak 1100  $\text{cm}^{-1}$  does. These shifts may be the  
278 reason of the more hydrophobic nature of the membranes compared to the pristine and  
279 CS-DES blends (non-cross linked) [18], which is in agreement with the CA data.

280



281

282

**Figure 2.** FTIR spectra for pristine and cross-linked CS membranes and their blends with DESs.

283

284

285  
286  
287  
288  
289  
290  
291  
292  
293  
294  
295  
296  
297  
298  
299  
300  
301  
302  
303  
304  
305  
306  
307





An important aspect to address during the incorporation of DESs in membranes deals with the possible effect on the membrane hydrophilicity/hydrophobicity. The pristine CS membrane showed a CA value of 90°, in line with the data reported in literature which place the CA of CS polymer between 84-88° [32,35]. The variability in the hydrophilic/hydrophobic nature of CS strongly depends on the deacetylation degree of the polymer; for instance, a high degree of deacetylation provides highly hydrophilic CS membranes [36], meaning more amine groups available on the CS molecule which can promote the water transport through the membrane matrix [37]. In principle, the hydrophilicity of CS belongs to its hydrophilic groups, such as -OH and -NH<sub>2</sub>; however, such hydrophilicity decreases when cross-linking is applied. For this reason, the CA value reached up to 95° when CS was cross-linked.

The incorporation of the PRO:SUF DES in the CS membrane led to a drastic decrease of the contact angle value (34°). This may suggest that the polar groups provided by the DES (such as amino and carboxyl) have a big impact in enhancing the hydrophilicity of the overall membrane.





308 **Table 1.** CA values and mechanical properties of the pure CS membranes and its  
 309 blends with DES.

Membrane	CA (°)	Image:	Before PV			After PV		
			Young's modulus (N/mm <sup>2</sup> )	Tensile strength (N/mm <sup>2</sup> )	Elongation at break (%)	Young's Modulus (N/mm <sup>2</sup> )	Tensile strength (N/mm <sup>2</sup> )	Elongation at break (%)
CS	90±0.5		415 ± 32	61 ± 9	11 ± 7	632 ± 3	65 ± 11	12 ± 4
xCS	95±0.5		1467 ± 41	40 ± 11	6 ± 2	853 ± 206	45 ± 12	9 ± 2
CS:PRO:SUF	34±2		163 ± 10	38 ± 10	16 ± 4	164 ± 19	41 ± 2	12 ± 1
xCS:PRO:SUF	89±7		226 ± 47	50 ± 13	11 ± 2	205 ± 66	35 ± 13	8 ± 1

310  
 311 Surprisingly, the cross-linking of the CS membrane containing DES (xCS:PRO:SUF  
 312 membrane) led to an increase of the CA value (89°) close to the pristine CS membrane.  
 313 It is worth mentioning that, after a cross-linking protocol, the reaction of GA with primary  
 314 amino groups results in the formation of two Schiff bases involving both aldehyde groups  
 315 of the GA molecule [32,38]. Herein, the decrease of the number of the -NH<sub>2</sub> groups could  
 316 be the responsible of the contact angle increase in the xCS:PRO:SUF membrane.  
 317 As can be seen in **Table 1**, the blending of the DES with CS decreased specific  
 318 mechanical properties, such as Young's modulus and tensile strength (with a decrease

319 of about 61 and 38%, respectively). However, the impact of DES addition on membrane  
320 mechanical properties was observed to be reduced when the cross-linking was adopted.  
321 In this case, the reduction of Young's modulus and tensile strength, in comparison to the  
322 pristine CS membrane, was much reduced (45 and 18%, respectively). DESs can act as  
323 plasticizers into a CS polymer matrix causing a decrease in the intermolecular  
324 interactions. The addition of plasticizers in CS, in fact, conducts to a transition from a  
325 rigid to a softer material with elastic properties [39]. In particular, the loss of tensile  
326 strength is associated to the breakup of the film network provoked by the incorporation of  
327 the additive into the polymer. Similar results have been documented in literature in CS  
328 films when mixed with different additives [40] and natural chlorine chloride/ malonic acid  
329 eutectic mixtures [41]. It has been, in fact, documented that the increase in DES  
330 concentration is responsible of the decrease of Young's modulus and tensile strength in  
331 CS [41]. On the contrary, elongation at break was surprisingly preserved by the DES  
332 addition, which has been associated with the increase in the free volume in the polymer  
333 matrix. According to Jakubowska et al. [41], a possible expansion of a free volume fosters  
334 polymeric chain translation, being worthy in the stabilization of films in the elastic flow  
335 regime. In this case, the cross-linking could have re-established the original free volume  
336 of CS films after DES blending since the elongation at break was comparable to the initial  
337 value of the CS membrane.

338

### 339 3.3. *Pervaporation performance*

#### 340 3.3.1. *Effect of operating temperature on permeation and separation factor.*

341 The PV separation data for all tested membranes is reported in **Table 2**.

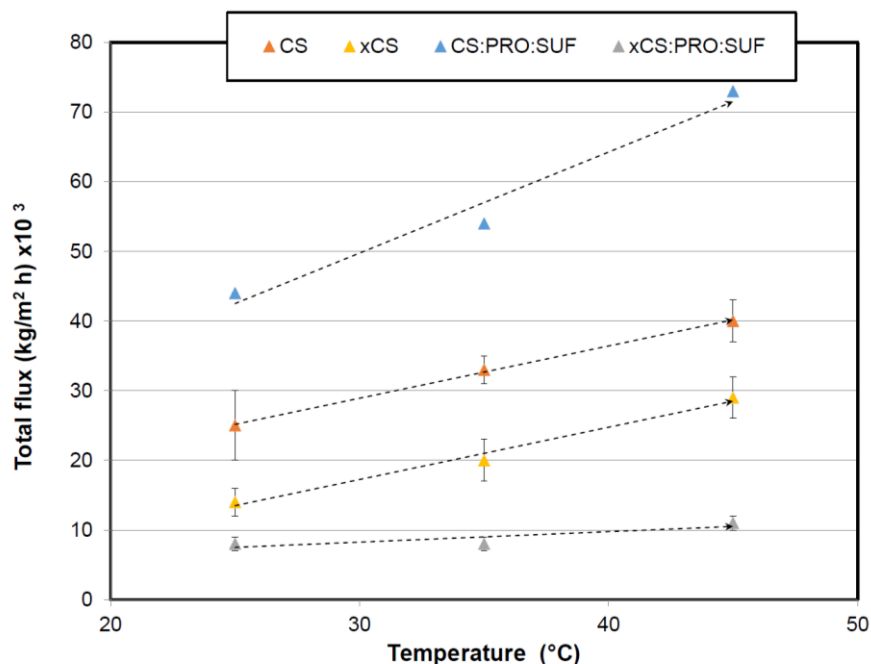


342 **Table 2.** Performance of CS-DES membranes as a function of temperature (feed  
 343 composition: 14.3 wt.% MeOH; 85.7 wt.% MTBE).

Temperature	CS			
	Total flux (kg m <sup>-2</sup> h <sup>-1</sup> ) × 10 <sup>3</sup>	Separation factor (α)	MeOH partial flux (kg m <sup>-2</sup> h <sup>-1</sup> ) × 10 <sup>3</sup>	MTBE partial flux (kg m <sup>-2</sup> h <sup>-1</sup> ) × 10 <sup>3</sup>
25°C	25±5	12.2± 0.11	16.9±3	8.2±2
35°C	33±2	5.6± 0.10	16.4±3	17±3
45°C	40±3	2.7± 0.09	17.4±2	27±4
xCS				
25°C	14±2	28.8± 0.10	11±2	2±0.2
35°C	20±3	27.6± 0.10	16±3	3±0.3
45°C	29±3	24.9± 0.11	24±2	5±0.5
CS:PRO:SUF				
25°C	44±1.5	1.3±0.21	7±0.1	33±1.7
35°C	54±0.0	1.1±0.06	8±5.7	45±0.4
45°C	73±3	1.0±0.029	11±0.8	62±2.8
xCS:PRO:SUF				
25°C	8±2	35.4±0.15	6.8±2	1±1
35°C	8±1	31.5±0.25	7±2	1.4±1
45°C	11±3	28.5±0.22	9.2±1	2.1±1

344  
 345 **Figure 3** graphically illustrates the effect of the feed temperature on the total permeate  
 346 flux, where a permeation increase, as a function of temperature, was observed in pristine

347 CS membrane and its blend with DES. Such a behaviour is a typical trend in polymeric  
 348 membranes since polymer chains are more flexible at higher temperatures fostering the  
 349 sorption ability of the solvent molecules, conducting to an increase in permeation of  
 350 compounds across the intermolecular distances of the polymeric membrane [42].  
 351



352  
 353 **Figure 3.** Effect of feed temperature on total flux (feed composition: 14.3 wt.% MeOH;  
 354 85.7 wt.% MTBE, pressure: 0.05 mbar). The curves are only guides to the eye.

355  
 356 The temperature dependence on permeate flux was determined by means of the  
 357 Arrhenius model, as expressed in Equation 5.

358  
 359 
$$J = J_o \cdot \exp\left(-\frac{E_A}{R \cdot T}\right) \quad \text{Eq. (5)}$$

360 where  $J_o$  corresponds to the pre-exponential factor,  $E_a$  refers to the apparent

361 activation energy for permeation (for the mixture and each compounds)  
362 and  $R \cdot T$  corresponds to the common energy term. Using the logarithms for Eq. 5.  
363  $E_A$  can be calculated in a linear function, which confirms that an Arrhenius relationship  
364 occurs between total fluxes and operating temperature; in other words, an increase in  
365 total permeation takes places by increasing temperature. From **Table 3**, it can be seen  
366 that crosslinked CS membrane display lower  $E_a$  values for methanol (ca. 13.34 kJ/mol)  
367 than MTBE (ca. 15.66 kJ/mol), which gives an input of methanol selectivity of CS.  
368 Interestingly, the incorporation of the hydrophilic DES lowered the  $E_a$  value for  
369 methanol up to 0.49 kJ/mol in xCS:PRO:SUF membrane, while the  $E_a$  value for MTBE  
370 was less affected. At this point, the  $E_a$  decreased substantially towards MeOH  
371 compared with MTBE in the range of 25-45 °C. It is worth mentioning that the PV  
372 process in the operating temperature affects primarily the permeation rate of MeOH,  
373 and does influence minimally the MTBE transport. It is clear that the presence of DES  
374 lowers the energy needed for the molecules to permeate across the membranes and  
375 this is particularly evident for MeOH molecules. This also can be supported by the  
376 hydrophilic nature of the DES, which definitely may display a preference for polar  
377 compounds (like MeOH) [28].

378

379

380

381

382



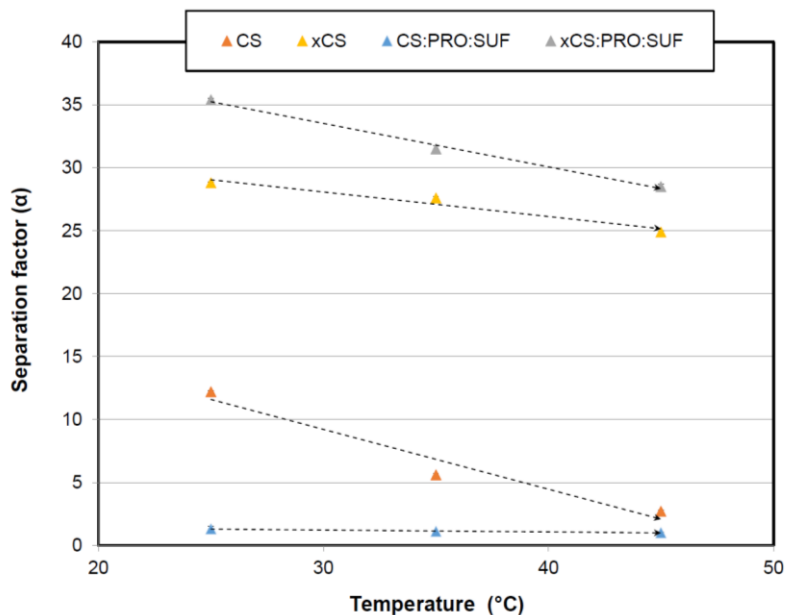
383 **Table 3.** Apparent activation energies for total permeate, MeOH and MTBE partial  
 384 fluxes of the CS-DES membranes.

385

Membrane	Activation energy values (kJ/mol)		
	Total	Methanol	MTBE
CS	8.05	5.12	20.44
xCS	12.46	13.34	15.66
CS:PRO:SUF	8.64	10.79	7.70
xCS:PRO:SUF	5.38	0.49	12.63

386

387 Regarding the selectivity, the separation factor in pristine CS membrane decreases as a  
 388 function of the temperature (see **Figure 4**). Importantly, such selectivity was improved by  
 389 incorporating the DES with the aid of in situ cross-linking, reaching a value up to 35 (at  
 390 25°C). In general, high separation factors and lower permeation rates were, in fact,  
 391 obtained at the lowest temperatures for all membranes. This agrees with the free volume  
 392 theory, which establishes that thermal motion of polymer chains in the amorphous regions  
 393 induces a free volume increase. As temperature increases, the frequency and amplitude  
 394 of the chain jumping (i.e., thermal agitation) increases, and consequently, the free volume  
 395 becomes larger [43]. Even if the kinetic diameters of MeOH and MTBE substantially differ  
 396 (3.6 and 6.2 Å, respectively), the thermal motion of the polymeric chains facilitates also  
 397 the diffusion of larger molecules (like MTBE) across the membrane conducting to a  
 398 decrease in the separation factor. Additionally, the separation factor decrease agrees with  
 399 the fact that activation energy values for MTBE were larger than for MeOH.



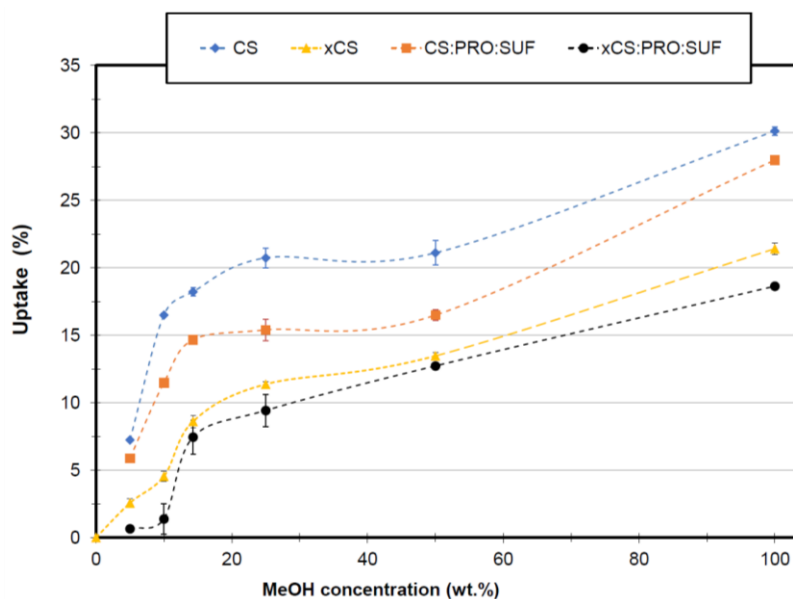
400  
 401 **Figure 4.** Effect of feed temperature on the separation factor (feed composition:  
 402 14.3 wt.% MeOH; 85.7 wt.% MTBE, pressure: 0.05 mbar). The curves are only guides to  
 403 the eye.

404  
 405 It is important to mention that *in situ* cross-linking was required at this specific case. As  
 406 can be seen in **Table 2**, the direct incorporation of DES into CS led to a loss of the  
 407 selective properties of the membrane. To date, it has been documented that DES addition  
 408 tends to increase the free volume in polymer matrices. According to Jakubowska et al.  
 409 [41], a possible expansion of the free volumes fosters the polymeric chain translation,  
 410 which, if from one side can enhance the stabilization of films at elastic flow regime  
 411 (improving the elongation at break properties), from the other side it can lower the  
 412 selective performance of the membrane.

413 Finally, the determination of the uptake properties of the studied membranes is reported  
 414 in **Figure 5**. It can be seen that all membranes display low uptake values, e.g., lower

415 than 7%, for low MeOH concentrations (up to 10 wt.%). However, the increase of MeOH  
416 concentration in the feed mixture, led to a higher swelling of the membranes. The addition  
417 of DES caused a decrease in the solvent uptake ability of the membranes. This may  
418 support Jakubowska's hypothesis [41] that DES provides stabilization of the CS  
419 membranes. Of course, the uptake was substantially suppressed when cross-linking was  
420 applied since such a protocol makes polymer membranes more resistant to the solvent  
421 mixtures due to a higher restriction of polymer chains mobility [44]. It is well known that  
422 PV membranes which are less susceptible to swelling are preferred to guarantee more  
423 stable performance when separating organic/organic feed solutions. To point out, the  
424 membranes also exhibited, to some extent, stable mechanical properties since the  
425 properties did not change strongly after use in PV testing (see **Table 1**).

426



427

428 **Figure 5.** Uptake of CS-DES membranes at different MeOH concentrations (at 30 °C).

429

The curves are only guides to the eye.



430

### 431 3.3.2. Performance comparison of CS-DES membranes with literature

432 As in all membrane-based technologies, it is clear that the PV performance of either  
433 polymer and composite membranes is strongly influenced by several factors, including  
434 membrane properties (such as membrane material, nature, structure) and operating  
435 conditions (such as feed concentration, temperature, driving force, flow rate, etc.) [45].

436 To some extent, the membrane structure depends on the membrane preparation protocol  
437 used [46]. Therefore, since most of the research has been carried out at different  
438 operating conditions, it is challenging to provide a fair comparison of PV data among  
439 different studies [47]. In this work, we tentatively compare the performance of different  
440 membranes (pristine, blend and composites) at close operating conditions, as enlisted in

441 **Table 4.** The main bridle of hydrophilic polymers (like CS) is related to their high swelling  
442 tendency which may limit the mechanical properties and stability of the membranes  
443 prepared with them. If a membrane is swollen, there is a lack of efficient separation  
444 performance due to the polymer chains mobility. Therefore, many efforts have been done  
445 to improve the physicochemical properties of CS, by using several approaches such as  
446 the blending with polymers, agents and inorganic nanomaterials.

447 In this work, the best performance, in terms of selectivity, was obtained for the  
448 xCS:PRO:SUF membrane, which exhibited a separation factor of 35.4 (at 25 °C). Such a  
449 value corresponds to 3-fold higher separation factor compared with pristine CS  
450 membrane; unfortunately, the permeation flux was compromised. When compared with  
451 literature, our xCS:PRO:SUF membranes showed better selectivity than other polymers,  
452 such as modified polyether ether ketone (PEEKWC), poly(vinyl alcohol) (PVA), acrylic



453 acid plasma polymerized poly(3-hydroxybutyrate), and composite membranes (GO-  
 454 polyimide, polyamide filled with Al<sub>2</sub>O<sub>3</sub>) (see **Table 4**). Definitely, the permeation rate could  
 455 be improved by handling the operating temperature but it may affect the selective  
 456 properties.

457 On the contrary, xCS:PRO:SUF membranes did not show a competitive selectivity in  
 458 comparison to cellulose acetate/polyvinyl pyrrolidone (PVP), PVA/cellulose acetate  
 459 blend, cross-linked PVA, cellulose acetate filled with HZSM5 membranes, among other  
 460 composites. At this point, it is confirmed that these membranes (i.e., xCS:PRO:SUF) are  
 461 still limited by their selective-permeable trade-off.

462  
 463 **Table 4.** Comparison of CS-DES membrane performance with some pure polymeric  
 464 and mixed matrix membranes at close MeOH-MTBE azeotropic conditions.

Membrane material	Filler loading:	MeOH Concentration	Operating conditions	J (kg m <sup>-2</sup> h <sup>-1</sup> )	Separation factor (α)	Reference:
xCS:PRO:SUF	-	14.3 wt.% MeOH	25 °C, 0.05 mbar	0.008	35.4	This work
GO-polyimide	4 wt. %	14.3 wt.% MeOH	45 °C, 0.05 mbar	0.091	9.0	(Castro-Muñoz et al., 2019)
PEEKWC	-	15 wt.% MeOH	40 °C, 6.1 mbar	0.068	10	[49]
PVA	-	30 wt.% MeOH	45 °C, 15 mbar	0.900	25	[50]

Poly(lactic acid)	-	15 wt.% MeOH	30 °C, 6 mbar	0.620	5	[51]
Poly(lactic acid)	-	14.3 wt.% MeOH	40 °C, 6.1 mbar	0.090	75	[52]
Cellulose acetate-PVP blend	-	20 wt.% MeOH	45 °C, 3 mbar	0.225	340	[53]
PVA-cellulose acetate blend	-	15 wt.% MeOH	45 °C, 17 mbar	796*	1427	[54]
Acrylic acid plasma polymerized poly(3-hydroxybutyrate)	-	20 wt.% MeOH	45 °C, 1.3 mbar	11*	3	[55]
Cross-linked PVA	-	20 wt.% MeOH	50 °C, 0.4 mbar	0.036	1230	[56]
Cross-linked PAMHEMA	-	11 wt.% MeOH	50 °C, 1.33 mbar	0.140	150	[57]
Polyamide filled with Al <sub>2</sub> O <sub>3</sub>	10 wt.%	50 wt.% MeOH	30 °C	15*	20	[58]
cellulose acetate filled with HZSM5	0.5 wt.%	20 wt.% MeOH	40 °C, 3.3 mbar	4.2*	150	[59]
Sulfonated polyarylethersulfone with cardo filled with [Cu <sub>2</sub> (bdc) <sub>2</sub> (bpy)] <sub>n</sub>	30 wt.%	15 wt.% MeOH	40 °C, 6 mbar	0.28	2300	[60]
cellulose acetate filled with ZnO	10 wt.%	31 wt.% MeOH	40 °C, 5 mbar	2*	400	[61]
Sulfonated polyarylethersulfone with cardo filled with MIL-53(Al)-SO <sub>3</sub> H	15 wt.%	15 wt.% MeOH	40 °C, 6 mbar	0.368	1990	[62]
CS	-	30 wt.% MeOH	50 °C	0.120	7	[63]

465 \* Normalized flux by thickness.



#### 4. Concluding remarks and future perspectives

In this work, dense CS-hydrophilic L-proline:sulfolane membranes have been, for the first time, successfully prepared and characterized. By fully mapping the structure of the membranes, this study demonstrated complete miscibility and embodiment of the proposed novel hydrophilic DESs (PRO:SUF) into the polymer phase. Importantly, since we have used environmentally friendly materials (such as CS, water, organic DES), these membranes can be considered as pioneering work in manufacturing more sustainable dense bio-based membranes.

After preliminarily testing in MeOH-MTBE separation using PV process, this new concept of membranes (xCS:PRO:SUF) presents a 3-fold higher separation efficiency than the pristine CS. As an outlook, the future works can focus on improving the permeation rates of the membranes in order to make them more attractive for other PV separations. In fact, thanks to their ability in forming H-bonding interactions, DESs may promote an enhanced separation of other polar molecules from azeotropic mixtures [27]. Moreover, the novel membranes, due to their dense nature, can be also interesting to be tested in gas separation applications.

Finally, in view of making the entire fabrication process fully sustainable, GA can be replaced with a more benign cross-linker such as genipin.

#### Acknowledgments

The authors gratefully acknowledge the financial support from the National Science Centre, Warsaw, Poland – decision no. UMO-2018/30/E/ST8/00642. Financial support from Polish National Agency for Academic Exchange (NAWA) under Ulam Programme



489 (Agreement No. PPN/U LM/2020/1/00005/U/00001) is gratefully acknowledged. R.  
490 Castro-Muñoz also acknowledges the School of Engineering and Science and the  
491 FEMSA-Biotechnology Center at Tecnológico de Monterrey for their support through the  
492 Bioprocess (0020209113) Focus Group. The authors also acknowledged Emilia Gontarek  
493 for her indispensable support for FTIR and contact angle measurements.

494

#### 495 **Conflict of Interest**

496 The authors declare no conflict of interest.

497

#### 498 **References**

- 499 [1] P.T. Anastas, J.C. Warner, *Green Chemistry: Theory and Practice*, Green Chem.  
500 Theory Pract. Oxford Univ. Press. New York. (1998).
- 501 [2] P.T. Anastas, N. Eghbali, *Green Chemistry: Principles and Practice*, Chem. Soc.  
502 Rev. 39 (2010) 201–312.
- 503 [3] E.L. Smith, A.P. Abbott, K.S. Ryder, *Deep Eutectic Solvents (DESS) and Their*  
504 *Applications*, Chem. Rev. 114 (2014) 11060–11082.  
505 <https://doi.org/10.1021/cr300162p>.
- 506 [4] P. Liu, J.W. Hao, L.P. Mo, Z.H. Zhang, Recent advances in the application of  
507 deep eutectic solvents as sustainable media as well as catalysts in organic  
508 reactions, RSC Adv. 5 (2015) 48675–48704. <https://doi.org/10.1039/c5ra05746a>.
- 509 [5] J. Huang, X. Guo, T. Xu, L. Fan, X. Zhou, S. Wu, Ionic deep eutectic solvents for  
510 the extraction and separation of natural products, J. Chromatogr. A. 1598 (2019)  
511 1–19. <https://doi.org/10.1016/j.chroma.2019.03.046>.



- 512 [6] A.R. Harifi-Mood, F. Mohammadpour, G. Boczkaj, Solvent dependency of carbon  
513 dioxide Henry's constant in aqueous solutions of choline chloride-ethylene glycol  
514 based deep eutectic solvent, *J. Mol. Liq.* 319 (2020) 114173.  
515 <https://doi.org/10.1016/j.molliq.2020.114173>.
- 516 [7] P. Makoś, G. Boczkaj, Deep eutectic solvents based highly efficient extractive  
517 desulfurization of fuels – Eco-friendly approach, *J. Mol. Liq.* 296 (2019) 111916.  
518 <https://doi.org/10.1016/j.molliq.2019.111916>.
- 519 [8] M. Momotko, J. Łuczak, A. Przyjazny, G. Boczkaj, First deep eutectic solvent-  
520 based (DES) stationary phase for gas chromatography and future perspectives for  
521 DES application in separation techniques, *J. Chromatogr. A.* 1635 (2020) 461701.  
522 <https://doi.org/10.1016/j.chroma.2020.461701>.
- 523 [9] F. Merza, A. Fawzy, I. AlNashef, S. Al-Zuhair, H. Taher, Effectiveness of using  
524 deep eutectic solvents as an alternative to conventional solvents in enzymatic  
525 biodiesel production from waste oils, *Energy Reports.* 4 (2018) 77–83.  
526 <https://doi.org/10.1016/j.egyr.2018.01.005>.
- 527 [10] Y.P. Mbous, M. Hayyan, A. Hayyan, W.F. Wong, M.A. Hashim, C.Y. Looi,  
528 Applications of deep eutectic solvents in biotechnology and bioengineering—  
529 Promises and challenges, *Biotechnol. Adv.* 35 (2017) 105–134.  
530 <https://doi.org/10.1016/j.biotechadv.2016.11.006>.
- 531 [11] A.E. Ünlü, A. Arlkaya, S. Takaç, Use of deep eutectic solvents as catalyst: A mini-  
532 review, *Green Process. Synth.* 8 (2019) 355–372. [https://doi.org/10.1515/gps-](https://doi.org/10.1515/gps-2019-0003)  
533 [2019-0003](https://doi.org/10.1515/gps-2019-0003).
- 534 [12] M. Taghizadeh, A. Taghizadeh, V. Vatanpour, M.R. Ganjali, M.R. Saeb, Deep



- 535 eutectic solvents in membrane science and technology: Fundamental,  
536 preparation, application, and future perspective, *Sep. Purif. Technol.* 258 (2021)  
537 118015. <https://doi.org/10.1016/j.seppur.2020.118015>.
- 538 [13] B. Jiang, H. Dou, L. Zhang, B. Wang, Y. Sun, H. Yang, Z. Huang, H. Bi, Novel  
539 supported liquid membranes based on deep eutectic solvents for olefin-paraffin  
540 separation via facilitated transport, *J. Memb. Sci.* 536 (2017) 123–132.  
541 <https://doi.org/10.1016/j.memsci.2017.05.004>.
- 542 [14] Z. Dai, H. Aboukeila, L. Ansaloni, J. Deng, M. Giacinti Baschetti, L. Deng,  
543 Nafion/PEG hybrid membrane for CO<sub>2</sub> separation: Effect of PEG on membrane  
544 micro-structure and performance, *Sep. Purif. Technol.* 214 (2019) 67–77.  
545 <https://doi.org/10.1016/j.seppur.2018.03.062>.
- 546 [15] Z. Dai, L. Ansaloni, J.J. Ryan, R.J. Spontak, L. Deng, Nafion/IL hybrid  
547 membranes with tuned nanostructure for enhanced CO<sub>2</sub> separation: Effects of  
548 ionic liquid and water vapor, *Green Chem.* 20 (2018) 1391–1404.  
549 <https://doi.org/10.1039/c7gc03727a>.
- 550 [16] N.M. Mahmoodi, M. Taghizadeh, A. Taghizadeh, Activated carbon/metal-organic  
551 framework composite as a bio-based novel green adsorbent: Preparation and  
552 mathematical pollutant removal modeling, *J. Mol. Liq.* 277 (2019) 310–322.  
553 <https://doi.org/10.1016/j.molliq.2018.12.050>.
- 554 [17] B. Jiang, N. Zhang, L. Zhang, Y. Sun, Z. Huang, B. Wang, H. Dou, H. Guan,  
555 Enhanced separation performance of PES ultrafiltration membranes by imidazole-  
556 based deep eutectic solvents as novel functional additives, *J. Memb. Sci.* 564  
557 (2018) 247–258. <https://doi.org/10.1016/j.memsci.2018.07.034>.



- 558 [18] P.G. Ingole, N.R. Thakare, K. Kim, H.C. Bajaj, K. Singh, H. Lee, Preparation,  
559 characterization and performance evaluation of separation of alcohol using  
560 crosslinked membrane materials, *New J. Chem.* 37 (2013) 4018–4024.  
561 <https://doi.org/10.1039/c3nj00952a>.
- 562 [19] N. Ptaszyńska, K. Gucwa, K. Olkiewicz, M. Heldt, M. Serocki, A. Stupak, D.  
563 Martynow, D. Dębowski, A. Gitlin-Domagalska, J. Lica, A. Łęgowska, S. Milewski,  
564 K. Rolka, Conjugates of ciprofloxacin and levofloxacin with cell-penetrating  
565 peptide exhibit antifungal activity and mammalian cytotoxicity, *Int. J. Mol. Sci.* 21  
566 (2020) 1–25. <https://doi.org/10.3390/ijms21134696>.
- 567 [20] R. Castro-Muñoz, F. Galiano, V. Fíla, E. Drioli, A. Figoli, Matrimid® 5218 dense  
568 membrane for the separation of azeotropic MeOH- MTBE mixtures by  
569 pervaporation, *Sep. Purif. Technol.* 199 (2018) 27–36.  
570 <https://doi.org/10.1016/j.seppur.2018.01.045>.
- 571 [21] E.P. Favvas, A. Figoli, R. Castro-Muñoz, V. Fíla, X. He, Polymeric membrane  
572 materials for CO<sub>2</sub> separations, 2018. [https://doi.org/10.1016/B978-0-12-813645-](https://doi.org/10.1016/B978-0-12-813645-4.00001-5)  
573 [4.00001-5](https://doi.org/10.1016/B978-0-12-813645-4.00001-5).
- 574 [22] H.G. Premakshi, K. Ramesh, M.Y. Kariduraganavar, Modification of crosslinked  
575 chitosan membrane using NaY zeolite for pervaporation separation of water-  
576 isopropanol mixtures, *Chem. Eng. Res. Des.* 94 (2015) 32–43.  
577 <https://doi.org/10.1016/j.cherd.2014.11.014>.
- 578 [23] M. Loloie, M. Omidkhah, A. Moghadassi, A.E. Amooghin, Preparation and  
579 characterization of Matrimid® 5218 based binary and ternary mixed matrix  
580 membranes for CO<sub>2</sub> separation, *Int. J. Greenh. Gas Control.* 39 (2015) 225–235.





- 581 <https://doi.org/http://dx.doi.org/10.1016/j.ijggc.2015.04.016>.
- 582 [24] R. Castro-Muñoz, V. Fíla, M.Z. Ahmad, Enhancing the CO<sub>2</sub> Separation  
583 Performance of Matrimid 5218 Membranes for CO<sub>2</sub>/CH<sub>4</sub> Binary Mixtures,  
584 Chem. Eng. Technol. (2019) ceat.201800111.  
585 <https://doi.org/10.1002/ceat.201800111>.
- 586 [25] B. Jiang, N. Zhang, B. Wang, N. Yang, Z. Huang, H. Yang, Z. Shu, Deep eutectic  
587 solvent as novel additive for PES membrane with improved performance, Sep.  
588 Purif. Technol. 194 (2018) 239–248. <https://doi.org/10.1016/j.seppur.2017.11.036>.
- 589 [26] F. Russo, R. Castro-Muñoz, F. Galiano, A. Figoli, Unprecedented preparation of  
590 porous Matrimid® 5218 membranes, J. Memb. Sci. (2019).  
591 <https://doi.org/10.1016/j.memsci.2019.05.036>.
- 592 [27] M.K. Hadj-Kali, H.F. Hizaddin, I. Wazeer, L. El blidi, S. Mulyono, M.A. Hashim,  
593 Liquid-liquid separation of azeotropic mixtures of ethanol/alkanes using deep  
594 eutectic solvents: COSMO-RS prediction and experimental validation, Fluid  
595 Phase Equilib. 448 (2017) 105–115. <https://doi.org/10.1016/j.fluid.2017.05.021>.
- 596 [28] A. Pandey, R. Rai, M. Pal, S. Pandey, How polar are choline chloride-based deep  
597 eutectic solvents?, Phys. Chem. Chem. Phys. 16 (2014) 1559–1568.  
598 <https://doi.org/10.1039/c3cp53456a>.
- 599 [29] I.M. Arcana, B. Bundjali, I. Yudistira, B. Jariah, L. Sukria, Study on properties of  
600 polymer blends from polypropylene with polycaprolactone and their  
601 biodegradability, Polym. J. 39 (2007) 1337–1344.  
602 <https://doi.org/10.1295/polymj.PJ2006250>.
- 603 [30] R. Castro-Muñoz, M.Z. Ahmad, V. Fíla, Tuning of Nano-Based Materials for

- 604 Embedding Into Low-Permeability Polyimides for a Featured Gas Separation,  
605 Front. Chem. 7 (2020) 1–14. <https://doi.org/10.3389/fchem.2019.00897>.
- 606 [31] R. Castro-Muñoz, J. González-Valdez, M.Z. Ahmad, High-performance  
607 pervaporation chitosan-based membranes: new insights and perspectives, Rev.  
608 Chem. Eng. (2020) 20190051. <https://doi.org/https://doi.org/10.1515/revce-2019-0051>.
- 609 0051.
- 610 [32] H.S. Tsai, Y.Z. Wang, Properties of hydrophilic chitosan network membranes by  
611 introducing binary crosslink agents, Polym. Bull. 60 (2008) 103–113.  
612 <https://doi.org/10.1007/s00289-007-0846-x>.
- 613 [33] J.Z. Knaul, S.M. Hudson, K.A.M. Creber, Crosslinking of chitosan fibers with  
614 dialdehydes: Proposal of a new reaction mechanism, J. Polym. Sci. Part B Polym.  
615 Phys. 37 (1999) 1079–1094. [https://doi.org/10.1002/\(SICI\)1099-0488\(19990601\)37:11<1079::AID-POLB4>3.0.CO;2-O](https://doi.org/10.1002/(SICI)1099-0488(19990601)37:11<1079::AID-POLB4>3.0.CO;2-O).
- 616 0488(19990601)37:11<1079::AID-POLB4>3.0.CO;2-O.
- 617 [34] R.S. Vieira, M.M. Beppu, Interaction of natural and crosslinked chitosan  
618 membranes with Hg(II) ions, Colloids Surfaces A Physicochem. Eng. Asp. 279  
619 (2006) 196–207. <https://doi.org/10.1016/j.colsurfa.2006.01.026>.
- 620 [35] S.P. Dharupaneedi, R. V. Anjanapura, J.M. Han, T.M. Aminabhavi, Functionalized  
621 graphene sheets embedded in chitosan nanocomposite membranes for ethanol  
622 and isopropanol dehydration via pervaporation, Ind. Eng. Chem. Res. 53 (2014)  
623 14474–14484. <https://doi.org/10.1021/ie502751h>.
- 624 [36] M. Kong, X.G. Chen, K. Xing, H.J. Park, Antimicrobial properties of chitosan and  
625 mode of action: A state of the art review, Int. J. Food Microbiol. 144 (2010) 51–63.  
626 <https://doi.org/10.1016/j.ijfoodmicro.2010.09.012>.



- 627 [37] M. Mathaba, M.O. Daramola, Effect of chitosan's degree of deacetylation on the  
628 performance of pes membrane infused with chitosan during amd treatment,  
629 Membranes (Basel). 10 (2020). <https://doi.org/10.3390/membranes10030052>.
- 630 [38] T.Y. Hsien, G.L. Rorrer, Effects of Acylation and Crosslinking on the Material  
631 Properties and Cadmium Ion Adsorption Capacity of Porous Chitosan Beads,  
632 Sep. Sci. Technol. 30 (1995) 2455–2475.  
633 <https://doi.org/10.1080/01496399508021395>.
- 634 [39] J.R. Rodríguez-Núñez, T.J. Madera-Santana, D.I. Sánchez-Machado, J. López-  
635 Cervantes, H. Soto Valdez, Chitosan/Hydrophilic Plasticizer-Based Films:  
636 Preparation, Physicochemical and Antimicrobial Properties, J. Polym. Environ. 22  
637 (2014) 41–51. <https://doi.org/10.1007/s10924-013-0621-z>.
- 638 [40] Y. Pranoto, S.K. Rakshit, V.M. Salokhe, Enhancing antimicrobial activity of  
639 chitosan films by incorporating garlic oil, potassium sorbate and nisin, LWT - Food  
640 Sci. Technol. 38 (2005) 859–865. <https://doi.org/10.1016/j.lwt.2004.09.014>.
- 641 [41] E. Jakubowska, M. Gierszewska, J. Nowaczyk, E. Olewnik-Kruszkowska,  
642 Physicochemical and storage properties of chitosan-based films plasticized with  
643 deep eutectic solvent, Food Hydrocoll. 108 (2020) 106007.  
644 <https://doi.org/10.1016/j.foodhyd.2020.106007>.
- 645 [42] C. Nagel, K. Günther-Schade, D. Fritsch, T. Strunskus, F. Faupel, Free volume  
646 and transport properties in highly selective polymer membranes, Macromolecules.  
647 35 (2002) 2071–2077. <https://doi.org/10.1021/ma011028d>.
- 648 [43] R. Huang, C. Yeom, Pervaporation separation of aqueous mixtures using  
649 crosslinked poly(vinyl alcohol)(pva). II. Permeation of ethanol-water mixtures, J.



- 650 Memb. Sci. 51 (1990) 273–292.
- 651 [44] Y.L. Xue, J. Huang, C.H. Lau, B. Cao, P. Li, Tailoring the molecular structure of  
652 crosslinked polymers for pervaporation desalination, *Nat. Commun.* 11 (2020)  
653 1461. <https://doi.org/10.1038/s41467-020-15038-w>.
- 654 [45] F. Galiano, F. Falbo, A. Figoli, *Polymeric Pervaporation Membranes: Organic-*  
655 *Organic Separation*, 2016. <https://doi.org/10.1002/9781118831823.ch7>.
- 656 [46] M. Rezakazemi, A. Ebadi Amooghin, M.M. Montazer-Rahmati, A.F. Ismail, T.  
657 Matsuura, State-of-the-art membrane based CO<sub>2</sub> separation using mixed matrix  
658 membranes (MMMs): An overview on current status and future directions, *Prog.*  
659 *Polym. Sci.* (2014). <https://doi.org/10.1016/j.progpolymsci.2014.01.003>.
- 660 [47] A. Figoli, S. Santoro, F. Galiano, A. Basile, Pervaporation membranes:  
661 preparation, characterization, and application, in: A. Basile, A. Figoli, M. Khayet  
662 (Eds.), *Pervaporation, Vap. Permeat. Membr. Distill.*, First edit, Elsevier Ltd.,  
663 Cambridge UK, 2015: pp. 281–304.
- 664 [48] R. Castro-Muñoz, F. Galiano, Ó. de la Iglesia, V. Fíla, C. Téllez, J. Coronas, A.  
665 Figoli, Graphene oxide – Filled polyimide membranes in pervaporative separation  
666 of azeotropic methanol–MTBE mixtures, *Sep. Purif. Technol.* 224 (2019) 265–  
667 272. <https://doi.org/10.1016/j.seppur.2019.05.034>.
- 668 [49] S. Zereshki, A. Figoli, S.S. Madaeni, S. Simone, M. Esmailinezhad, E. Drioli,  
669 Pervaporation separation of MeOH/MTBE mixtures with modified PEEK  
670 membrane: Effect of operating conditions, *J. Memb. Sci.* 371 (2011) 1–9.  
671 <https://doi.org/10.1016/j.memsci.2010.11.068>.
- 672 [50] M. Peivasti, A. Madandar, T. Mohammadi, Effect of operating conditions on



- 673 pervaporation of methanol / methyl tert -butyl ether mixtures, 47 (2008) 1069–  
674 1074. <https://doi.org/10.1016/j.cep.2007.08.005>.
- 675 [51] S. Zereshki, A. Figoli, S.S. Madaeni, S. Simone, E. Drioli, Pervaporation  
676 Separation of Methanol / Methyl tert -Butyl Ether with Poly ( lactic acid )  
677 Membranes, J. Appl. Polym. Sci. 118 (2010) 1364–1371.  
678 <https://doi.org/10.1002/app>.
- 679 [52] F. Galiano, A.H. Ghanim, K.T. Rashid, T. Marino, S. Simone, Q.F. Alsahy, A.  
680 Figoli, Preparation and characterization of green polylactic acid (PLA) membranes  
681 for organic/organic separation by pervaporation, Clean Technol. Environ. Policy.  
682 21 (2019) 109–120. <https://doi.org/10.1007/s10098-018-1621-4>.
- 683 [53] H. Wu, X. Fang, X. Zhang, Z. Jiang, B. Li, X. Ma, Cellulose acetate – poly ( N-  
684 vinyl-2-pyrrolidone ) blend membrane for pervaporation separation of methanol /  
685 MTBE mixtures, 64 (2008) 183–191. <https://doi.org/10.1016/j.seppur.2008.09.013>.
- 686 [54] K. Zhou, Q. Gen, G.L. Han, A.M. Zhu, Q. Lin, Pervaporation of water – ethanol  
687 and methanol – MTBE mixtures using poly ( vinyl alcohol )/ cellulose acetate  
688 blended membranes, J. Memb. Sci. 448 (2013) 93–101.  
689 <https://doi.org/10.1016/j.memsci.2013.08.005>.
- 690 [55] M. Villegas, A.I. Romero, M.L. Parentis, E.F.C. Vidaurre, J.C. Gottifredi, Chemical  
691 Engineering Research and Design Acrylic acid plasma polymerized poly ( 3-  
692 hydroxybutyrate ) membranes for methanol / MTBE separation by pervaporation,  
693 Chem. Eng. Res. Des. 109 (2016) 234–248.  
694 <https://doi.org/10.1016/j.cherd.2016.01.018>.
- 695 [56] J. Rhim, Y. Kim, Pervaporation Separation of MTBE – Methanol Mixtures Using

- 696 Cross-linked PVA Membranes, (1999) 1699–1707.
- 697 [57] S. Ray, S.K. Ray, Synthesis of highly methanol selective membranes for  
698 separation of methyl tertiary butyl ether ( MTBE )– methanol mixtures by  
699 pervaporation, 278 (2006) 279–289.  
700 <https://doi.org/10.1016/j.memsci.2005.11.011>.
- 701 [58] R. Kopec, M. Meller, W. Kujawski, J. Kujawa, Polyamide-6 based pervaporation  
702 membranes for organic-organic separation, Sep. Purif. Technol. 110 (2013) 63–  
703 73. <https://doi.org/10.1016/j.seppur.2013.03.007>.
- 704 [59] X. Ma, C. Hu, R. Guo, X. Fang, H. Wu, Z. Jiang, HZSM5-filled cellulose acetate  
705 membranes for pervaporation separation of methanol/MTBE mixtures, Sep. Purif.  
706 Technol. 59 (2008) 34–42. <https://doi.org/10.1016/j.seppur.2007.05.023>.
- 707 [60] G.L. Han, K. Zhou, A.N. Lai, Q.G. Zhang, A.M. Zhu, Q. Lin, [ Cu<sub>2</sub> ( bdc )<sub>2</sub> ( bpy  
708 ) ] n / SPES-C mixed matrix membranes for separation of methanol / methyl tert-  
709 butyl ether mixtures, J. Memb. Sci. 454 (2014) 36–43.  
710 <https://doi.org/10.1016/j.memsci.2013.11.049>.
- 711 [61] Y. Wang, L. Yang, G. Luo, Y. Dai, Preparation of cellulose acetate membrane  
712 filled with metal oxide particles for the pervaporation separation of  
713 methanol/methyl tert-butyl ether mixtures, Chem. Eng. J. 146 (2009) 6–10.  
714 <https://doi.org/10.1016/j.cej.2008.05.009>.
- 715 [62] G.L. Han, Z. Chen, L.F. Cai, Y.H. Zhang, J.F. Tian, H.H. Ma, S.M. Fang, Post-  
716 synthetic MIL-53(Al)-SO<sub>3</sub> H incorporated sulfonated polyarylethersulfone with  
717 cardo (SPES-C) membranes for separating methanol and methyl tert-butyl ether  
718 mixture, Sep. Purif. Technol. 220 (2019) 268–275.



719 <https://doi.org/10.1016/j.seppur.2019.03.065>.  
720 [63] M.G. Mohd Nawawi, Z. Zamrud, Z. Idham, O. Hassan, N.M. Sakri, Blended  
721 Chitosan and Polyvinyl Alcohol Membrane for Pervaporation Operating  
722 Parameters, J. Teknol. (Sciences Eng. 65 (2013) 39–43).  
723