



Research article

Growth inhibition of cultured cancer cells by *Ribes nigrum* leaf extract

Mikayel Ginovyan¹, Agnieszka Bartoszek², Izabela Koss-Mikolajczyk², Barbara Kuszniereicz², Pierre Andreoletti³, Mustapha Cherkaoui-Malki³ and Naira Sahakyan^{1,*}

¹ Department of Biochemistry, Microbiology & Biotechnology; Research Institute of Biology Yerevan State University, 1 A. Manoogian Str., 0025 Yerevan, Armenia

² Department of Food Chemistry, Technology and Biotechnology, Faculty of Chemistry, Gdańsk University of Technology, Narutowicza 11/12, 80–233, Gdańsk, Poland

³ Laboratoire Bio-PeroxiL EA7270, University of Bourgogne Franche-Comté, 21000 Dijon, France

* **Correspondence:** Email: sahakyannaira@ysu.am; Tel: +37460710547.

Abstract: The present article includes data on the possible selective cytotoxic effect of extract of *Ribes nigrum* L. growing at high Armenian landscape. For this purpose, different non-cancer (microglial BV-2 wild type (*Wt*), acyl-CoA oxidase 1 (ACOX1) deficient (*Acox1*^{-/-}) and cancer (human colon adenocarcinoma HT29 and human breast cancer MCF7) cell lines were applied. *R. nigrum* leaf ethanol extract showed a growth inhibition effect towards HT29 and MCF7 cells started from 6 h of treatment at the concentration of 0.5 mg/mL DW. The lowest concentration (0.125 mg/mL DW) of the investigated extract expressed cytotoxicity after 72 hours following cancer cell treatment. In contrast to the cancer cells, in the case of the tested non-cancer cells, cytotoxic effect was not observed at the applied concentrations. The extract sub-cytotoxic concentration, in this case, was reported to be the 1 mg/mL DW. Further investigations are needed to confirm the selective cytotoxicity and possible action mechanisms of the leaf extract of *R. nigrum*.

Keywords: plant extract; Armenian flora; cytotoxicity; microglia; human adenocarcinoma; breast cancer

Abbreviations: ACOX1: acyl-CoA oxidase type 1; DMEM: Dulbecco's modified Eagle medium; DPPH: 1,1-diphenyl-2-picrylhydrazyl; EMEM: Minimal Essential Medium Eagle; DW: dry weight; EDTA: ethylenediamine tetraacetic acid; FBS: fetal bovine serum; GAE: gallic acid equivalent; MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; PBS: phosphate-buffered saline; QE: quercetin equivalent; VLCFA: very-long-chain fatty acids; WT: wild type

1. Introduction

Herbal preparations have a long-appreciated application, especially in medicine, due to their accessibility, powerful therapeutic value, synergistic effects with numerous clinically important medicines, and few side effects. Despite problems with bioavailability, currently, more than 20000 plant species are used in medicine and food production. On top of it, a huge number of medicinal preparations contain compounds derived from plants [1–4].

Ribes nigrum L. (black currant) leaf extracts are known for their beneficial effects on health. Due to their higher content of phenolic components, including flavonoids and anthocyanins, they possess anti-inflammatory, antioxidant, anticancer, analgesic, and hypotensive effects [5–8]. The black currant leaf is indicated as beneficial in many conditions, including arthrosis, bite, bladder stone, cancer, cold, cough, diarrhea, dysuria, fever, hepatitis, high blood pressure, infection, inflammation, pain, pertussis, *etc.* [6]. At the same time, there are no known contraindications for black currant leaf preparations concerning their acute toxicity, genotoxicity, reproductive or developmental toxicity, or carcinogenicity of this plant extract at doses relevant for human consumption [6,9]. The leaves of *R. nigrum* are also often used in European conventional remedies for the treatment and prophylaxis of numerous abnormalities in human metabolism [5,10]. Some investigations showed that the content of biologically active phenolic compounds is much higher in leaf extracts than in corresponding berry fruits [11]. The number of phenolic compounds present in *R. nigrum* leaf extracts exceeds 19, with epigallocatechin and epigallocatechin-3-gallate as the main antioxidant components [7,11–13].

Bearing in mind the above-mentioned data, we aimed to investigate the influence of the extract obtained from the leaves of Armenian *R. nigrum* growing at high altitudes on the viability of different cancer and normal cell lines. For this purpose, as non-cancer cells, we applied two types of neuroglial cell lines—BV-2 wild type (*Wt*) and recently generated acyl-CoA oxidase1 (ACOX1) deficient cells (*Acox1*^{-/-}) [14,15]. The BV-2 *Wt* cells were used to monitor the influence of the extract on normal cells. The *Acox1*-deficient microglial cells featured an accumulation of very-long-chain fatty acids (VLCFA) and induction of proinflammatory cytokines, were used as a cellular model of oxidative stress and inflammation, thus potentially responsive to antioxidants [16–18]. Since it is well-known that cancer initiation and progression are associated with oxidative stress in the cells, therefore, the BV-2 *Wt* and *Acox1*^{-/-} cell lines were chosen as models by which it could be possible to clarify whether the influence of the studied extract connected with the redox status of the cells [19,20].

Human colon adenocarcinoma HT29 and human breast cancer MCF7 cell lines were applied as cancer cell models [21–23]. This selection is based on the fact that HT29 cells serve not only as a colorectal cancer model, but also as an *in vitro* model in research on intestinal absorption, transport, and secretion involving cells that are in direct contact with consumed food components and medicinal preparations administered orally [24]. On the other hand, the application of MCF7 breast cancer cells was related to the high rate of this disease worldwide, including in the Armenian population. According to the WHO data published in 2020, breast cancer deaths in Armenia reached 444 per 100 thousand persons, representing 1.98% of total deaths [25,26].

2. Materials and methods

2.1. Chemicals and reagents

All applied chemicals and reagents were purchased from Sigma-Aldrich Co. Ltd.

2.2. Plant material

The plant material (*R. nigrum* L.) was collected from the Lori province (Armenia, 1600–1650 m a.s.l.) during its fruiting period (July 2019). Plant material was processed according to Hovhannisyan et al. [7]. Extraction was implemented using ethanol as solvent [27]. The obtained dried extracts were stored at 4 °C until further use. For the comet assay and MTT test involving cancerous cell lines, 50 mg DW/mL crude ethanol extract was prepared. The mass concentration of dissolved compounds in leaf extract was determined by drying 500 µL of extract and weighing each sample in three independent replicates. It was equal to 7.8 ± 0.57 mg/mL.

2.3. Cell cultures

Murine microglial BV-2 *Wt* and BV-2 Acyl-CoA oxidase type 1 (ACOX1) deficient mutant (*Acox1*^{-/-}) cell lines were provided by the Laboratory BioPeroxiL, University of Bourgogne, Dijon, France. BV-2 cells were from female C57BL/6J mice purchased from Banca-Biologica e Cell Factory (catalog no. ATL03001) were cultured in DMEM supplemented with 10% heat-inactivated FBS and 1% penicillin/streptomycin (Dutscher). Cultures were maintained at 37 °C in a humidified atmosphere with 5% of CO₂ [14].

BV-2 is derived from C57/BL6 murine. They are immortalized by v-raf/v-myc carrying J2 retrovirus. BV-2 express nuclear v-myc and the cytoplasmic v-raf oncogene products as well as the env gp70 antigen at the surface level. BV-2 microglia cell line retains microglia morphological and functional characteristics. In accordance with v-raf/v-myc expressed characters, the metabolic and proliferation rate of *in vitro* BV-2 greatly exceeds that of other microglia. BV-2 cells were seeded on 96-well microplates at 25×10^4 cells per well for viability assay and 6-well microplates at 5×10^5 cells per well—for enzymatic activity determination as described earlier [16,17].

Human colon adenocarcinoma HT29 and human breast cancer MCF7 cells were obtained from ATCC and maintained in McCoy's or EMEM (Minimal Essential Medium Eagle) medium, respectively, both supplemented with L-glutamine (2 mol/L), sodium pyruvate (200 g/L), fetal bovine serum (100 mL/L), and antibiotics (100 U/mL penicillin and 100 g/L streptomycin) [23,28]. Cells were maintained at 37 °C under a humidified atmosphere with 5% CO₂ in a Smart Cell Incubator (Heal Force, China) as described before by Koss-Mikołajczyk et al. [21]. Cultured cells were periodically checked for mycoplasma contamination (with Mycoplasma Detection Kit (ATCC, Monassas, VA, USA)).

2.4. MTT assay

BV-2 cells, plated in 96-wells plates, were treated for 24–72 h with various concentrations of *R. nigrum* ethanol extract (0.0001–10 mg/mL DW) and in the final step cell growth was evaluated

with 3-(4,5-dimethyltrazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay [29]. By this test we clarified the effect of *R. nigrum* extract on the viability and mitochondrial activity of the microglial cells.

The following concentrations of *R. nigrum* ethanol extract: 0.5, 0.25, 0.125, 0.06, 0.03, 0.015 mg/mL DW and exposure time for 3, 6, 24, or 72 h were used for the evaluation of HT29 and MCF7 cell growth inhibition. Treatments were performed as three independent replicates. Cytotoxicity was expressed as the percent cell growth of cells exposed to the plant extract normalized to control cells treated with the appropriate volume of solvent only (1% ethanol in the final mixture whose growth was regarded as 100%).

2.5. Genotoxic effects of *R. nigrum* leaf extract

Genotoxicity of *R. nigrum* leaf extract was evaluated using the comet assay [30]. About 10^5 HT29 cells were seeded in 24-well plates and treated with the plant extracts at different concentrations for 24 h [23]. Plant extract concentration was selected based on MTT cytotoxicity assay. The mean % DNA in tail was a measure of genotoxic potency of the extract. The damages on nuclei were analyzed with CometScore 2.0.0.38 software (Tritek Corp., Sumerduck, VA, USA). Two independent measurements were performed, in three technical replicates each.

2.6. Data processing

Statistical analyses were carried out by Student-t test and GraphPad Prism 8 software for calculation of the probability values. The statistical significance of the results of genotoxicity was examined by one-way ANOVA followed by Dunnett's test. The results are means \pm SD of three independent experiments. The results were considered statistically different at a *p*-value of 0.05 or less.

3. Results and discussion

The viability of cells under the influence of plant extracts is mostly connected with their chemical composition [2,16,17,31]. In our previous investigations, the content of total phenolic and flavonoid in *R. nigrum* leaf extracts were determined to amount to 84.1 ± 1.6 mg gallic acid equivalents (GAE)/g DW and 35.42 ± 1.52 mg quercetin equivalents (QE)/g DW, respectively. The main components of the extract comprised flavan-3-ols, flavonols, hydroxycinnamates, lignans, naphthols, furanocoumarins and quercetin derivatives [7]. Studies have shown, that extracts rich in phenolic components exhibit high antioxidant activity [17,31–35], which is aligned with our recent findings as the post-column derivatization of analytes with ABTS reagent indicated, that almost all of the major compounds identified in *R. nigrum* extract with LC-Q-Orbitrap HRMS analysis possess strong antioxidant activity [7] (See also Table 1). These findings served as a basis for further investigations on the influence of this extract on the viability of different cell lines, representing both normal and cancer models in *in vitro* studies. The results of current investigations showed the low cytotoxicity of the *R. nigrum* leaf extract on tested BV-2 normal cell lines (Figures 1 and 2), whereas it had considerable growth inhibitory activity against tested HT-29 and MCF7 cancer cell lines (Figure 3). According to the obtained results, the sub-cytotoxic concentration of investigated extract on both BV-2 cell lines was only reached at 1 mg/mL DW concentration (Figure 1).

Therefore, it can be stated that the investigated extract did not express toxicity against both BV-2 cell lines. This concentration of the extract was further applied to explore the effect of different exposure times on two BV-2 cell lines (Figure 2).

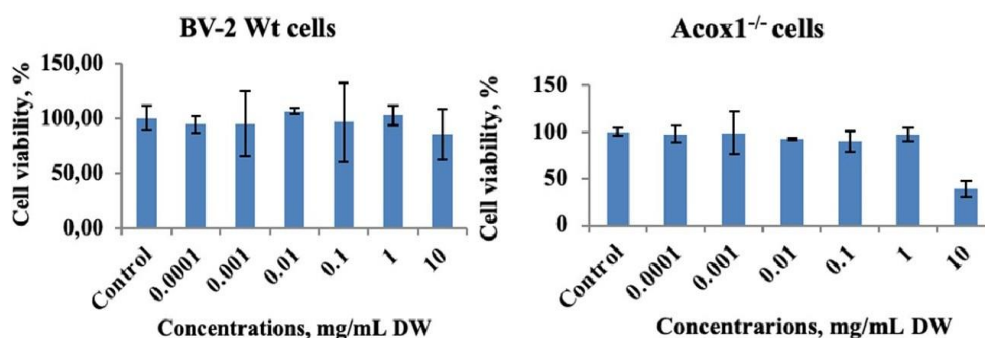


Figure 1. The viability of BV-2 *Wt* and *Acox1* deficient microglial cells under the treatment with *R. nigrum* extract. Cells were treated with different concentrations of extract (0.0001–10 mg/mL) for 24 h. Statistically significant difference was observed only at 10 mg/mL concentration of extract on *Acox1* deficient microglial cells ($p \leq 0.05$).

In case of the BV-2 *Wt* cells some statistically significant decrease in cell viability was observed after 48-hours of treatment with 1 mg/mL concentration of *R. nigrum* extract. This was not observed during the treatment of the *Acox1*^{-/-} cells (Figure 2), which could be due to the activation of some compensatory mechanisms [17]. Any significant changes were not detected in the viability of BV-2 *Acox1* cells following 24- or 48-hours' exposure times (Figure 2).

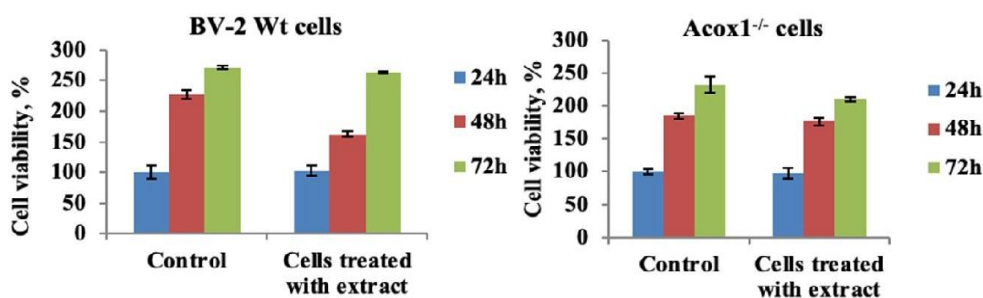


Figure 2. The influence of *R. nigrum* extract on viability kinetics of BV-2 *Wt* and *Acox1* deficient microglia cells (MTT assay). Cells were treated with 1 mg/mL of extract. Statistically significant differences were observed on BV-2 *Wt* cells only at 48-hour exposure times and BV-2 *Acox1* BV-2 at 72-hour exposure times ($p \leq 0.05$).

The ethanol extract of *R. nigrum* leaf showed considerable growth inhibitory effect towards HT29 and MCF7 cancer cell lines (Figure 3a,b) in contrast to the tested non-cancer BV-2 cell lines. In both cancer cell lines; the observed inhibition started from 6 h of treatment at the highest tested concentration (0.5 mg/mL DW) as well as after 72 h beginning from the lowest concentration (0.125 mg/mL DW). The growth inhibitory effect was thus both dose and time-dependent. In the case of the BV-2 *Wt*



and *Acox1* non-cancer cell lines, the exposure time had no significant effect on growth inhibition induced by *R. nigrum* extract. The 3-hour exposure time had only a slight inhibiting effect on the growth of HT29 cells, whereas it had not any influence on the growth of MCF7 cells at any tested concentration.

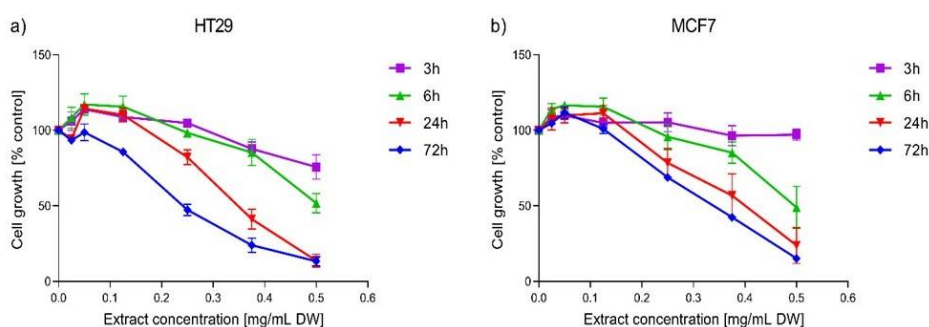


Figure 3. Growth inhibition of HT29 (a) and MCF7 (b) cells treated with *R. nigrum* ethanol extracts for 3, 6, 24 and 72 h (MTT test). The results are means \pm SD of three independent experiments carried out in triplicate ($p \leq 0.05$).

R. nigrum extract expressed noticeable cytotoxic activity against HT29 and MCF7 cells even at low concentrations. It can be of great interest, that *R. nigrum* leaf extract exhibited a rather low cytotoxicity against tested non-cancer cell lines in contrast to the documented in our study significant cytotoxicity against tested cancer cell lines. Ziemlewska et al. [36] also showed some beneficial effects of *R. nigrum* extract on the growth of two other normal cell lines: fibroblasts (BJ) and keratinocytes (HaCaT) at low concentrations and only low cytotoxicity was detected at high concentrations. It was also reported that *R. nigrum* leaf extract was not cytotoxic toward endothelial and normal kidney MDCK cells [9]. In another research, NCM460 normal colon epithelial cells maintained 90% viability even at 25 mg/mL concentration of *R. nigrum* leaf extract after 48 h exposure time [37].

Even though data obtained by us confirm that *R. nigrum* extract expressed strong cytotoxic activity towards tested cancer cell lines, there is very few literature data confirming this phenomenon for other cancer cell lines. Koss-Mikołajczyk et al. [28] showed the strong cytotoxic effect of *R. nigrum* fruits water extract on HT29 cells after 72-hour exposure time. Also, the noticeable cytotoxic effect on human gastric carcinoma MKN45 cells was reported by Liu et al. [37], however, only following high concentration exposures (about 15 mg/mL). Moreover, it was reported that the leaf extract of *R. nigrum* had no cytotoxic effect on A549 (lung carcinoma epithelial cell) and HeLa (cervical cancer cells) cells [9]. However, taking into account the obtained data and the presence of several major compounds with previously demonstrated anticancer and cytotoxic properties in the leaf extract of Armenian black currant which were identified in our previous research [7] (See Table 1), we presume that its active phytochemicals could possibly specifically target the cancerous cells. Nonetheless, further studies will be needed to elucidate the mechanisms behind this cytotoxicity and to confirm our presumptions.

To have some insight into the mechanism of cytotoxicity of *R. nigrum* leaf extract on HT29 cells, additionally, the genotoxicity of this extract was explored with the comet assay. According to the obtained results, *R. nigrum* extract exhibited some genotoxic activity on HT29 cells at 0.5 mg/mL DW concentration (Figure 4). According to one-way ANOVA with Dunnett's test, there were not observed significant differences between treated and non-treated cells at the lower tested concentrations.

Table 1. Major identified compounds in *R. nigrum* extract with possible antioxidant, anticancer and anti-inflammatory properties.

Tentative identification	Compound group	Post-column derivatization with ABTS (+/-)*	Biological (anticancer, antioxidant, anti-inflammatory) activities
Epicatechin gallate		+	Antioxidant, anticancer [23,38]
(epi)Galocatechin		+	Antioxidant, anticancer [23,39]
(+)-Catechin		+	Antioxidant, anticancer [23,39]
(-)-Epicatechin		+	Antioxidant, anticancer [23,39]
Procyanidins		+	Antioxidant, anticancer [40]
Caffeoylquinic acid	hydroxycinnamic acids and derivatives	+	Antioxidant, anticancer, anti-inflammatory [41,42]
Coumaric acid derivative		+	Antioxidant, anti-inflammatory, anticancer [39,43]
Rhamnetin glucoside	flavonols	+	N/A
Quercetin derivatives (Quercetin rutinoside, Quercetin hexoside, Quercetin glucuronide, Quercetin pentoside, Quercetin malonyl glucoside, Quercitrin)		+	Anticancer, antioxidant and anti-inflammatory [44–47]
Lariciresinol glucoside	lignan	+	N/A
Musizin glucoside and Musizin acetyl glucoside	naphthol	+	Anticancer, antioxidant [48]
Marmesin glucoside and Marmesin acetyl glucoside	furocoumarin (psoralen)	+	Anticancer [49,50]

* Antioxidant profiling was done by HPLC coupled post-column derivatization with ABTS reagent [7]. “+”—presence of antioxidant activity, “-”—absence of antioxidant activity, “N/A”—not applicable.

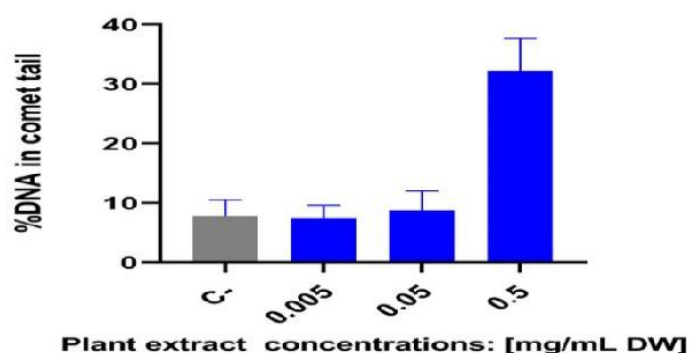


Figure 4. The genotoxicity of *R. nigrum* ethanol extract in HT29 cells. C- (Negative control), non-treated cells ($p \leq 0.05$).

As already mentioned, the extract was genotoxic at the tested highest concentration applied to HT29 cells based on comet assay results. There is no literature data concerning the genotoxicity of *R. nigrum* leaf extract. Our results are the first-time this phenomena is mentioned in these cell lines. The genotoxicity of the investigated extract in cancer cell lines can be due to the several compounds contained in the *R. nigrum* leaf extract, especially coumaric acid derivatives or furocoumarins (Table 1). These substances have been reported previously as anticancer agents which are acting through different mechanisms including the induction of DNA damage [43,50].

4. Conclusions

Based on the obtained data, the low cytotoxicity of *R. nigrum* leaf extract towards normal cell lines was confirmed. In contrast, the strong cytotoxic effect of the investigated extract was found against HT29 and MCF7 cancer cell lines. The extract also exhibited genotoxicity in HT29 cell line by inducing DNA fragmentation, which was detected using a comet assay. The presence of bioactive compounds, with possible anticancer and genotoxic properties, suggests the anticancer potential of the studied extract. Nonetheless, further investigations are needed to confirm the selective cytotoxicity and possible action mechanisms of the leaf extract of *R. nigrum* growing at high altitude in Armenian landscape.

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Conflict of interest

All authors declare no conflict of interest.

Author contributions

All authors contributed to the article formation equally. Authors revised and approved the final version of the manuscript.

References

1. Gupta PD, Birdi TJ (2017) Development of botanicals to combat antibiotic resistance. *J Ayurveda Integr Me* 8: 266–275. <https://doi.org/10.1016/j.jaim.2017.05.004>
2. Sahakyan N, Bartoszek A, Jacob C, et al. (2020) Bioavailability of tannins and other oligomeric polyphenols: a still to be studied phenomenon. *Curr Pharmacol Rep* 6: 131–136. <https://doi.org/10.1007/s40495-020-00217-6>
3. Sahakyan N, Petrosyan M, Koss-Mikołajczyk I, et al. (2019) The Caucasian flora: a still-to-be-discovered rich source of antioxidants. *Free Radical Res* 53: 1153–1162. <https://doi.org/10.1080/10715762.2019.1648799>

4. Sahakyan N, Petrosyan M, Trchounian A (2019) The activity of *Alkanna* species *in vitro* culture and intact plant extracts against antibiotic resistant bacteria. *Curr Pharm Design* 25: 1861–1865. <https://doi.org/10.2174/1381612825666190716112510>
5. Luzak B, Boncler M, Rywaniak J, et al. (2014) Extract from *Ribes nigrum* leaves *in vitro* activates nitric oxide synthase (eNOS) and increases CD39 expression in human endothelial cells. *J Physiol Biochem* 70: 1007–1019. <https://doi.org/10.1007/s13105-014-0370-z>
6. Duke JA (2002) *Handbook of Medicinal Herbs*, 2 Eds., Boca Raton: CRC Press. <https://doi.org/10.1201/9781420040463>
7. Hovhannisyanyan Z, Timotina M, Manoyan J, et al. (2022) *Ribes nigrum* L. extract-mediated green synthesis and antibacterial action mechanisms of silver nanoparticles. *Antibiotics* 11: 1415. <https://doi.org/10.3390/antibiotics11101415>
8. Magnavacca A, Piazza S, Cammisa A, et al. (2021) *Ribes nigrum* leaf extract preferentially inhibits ifn- γ -mediated inflammation in hacat keratinocytes. *Molecules* 26: 3044. <https://doi.org/10.3390/molecules26103044>
9. Jansone B, Laekeman G, Vlietinck A (2017) Assessment report on *Ribes nigrum* L., folium.
10. Sahakyan NZ, Petrosyan MT, Trchounian AH (2020) Increasing of the superoxide dismutase total activity in microglial cells under the treatment by *ribes nigrum* L. alcohol extract. *Proc YSU B Chem Biol Sci* 54: 216–222. <https://doi.org/10.46991/PYSU:B/2020.54.3.216>
11. Staszowska-Karkut M, Materska M (2020) Phenolic composition, mineral content, and beneficial bioactivities of leaf extracts from black currant (*Ribes nigrum* L.), raspberry (*Rubus idaeus*), and aronia (*Aronia melanocarpa*). *Nutrients* 12: 463. <https://doi.org/10.3390/nu12020463>
12. Butnariu M (2014) Detection of the polyphenolic components in *Ribes nigrum* L. *Ann Agr Env Med* 21: 11–14.
13. Paunović SM, Mašković P, Nikolić M, et al. (2017) Bioactive compounds and antimicrobial activity of black currant (*Ribes nigrum* L.) berries and leaves extract obtained by different soil management system. *Sci Horti-Amsterdam* 222: 69–75. <https://doi.org/10.1016/j.scienta.2017.05.015>
14. Raas Q, Gondcaille C, Hamon Y, et al. (2019) CRISPR/Cas9-mediated knockout of *Abcd1* and *Abcd2* genes in BV-2 cells: novel microglial models for X-linked adrenoleukodystrophy. *BBA-Mol Cell Biol L* 1864: 704–714. <https://doi.org/10.1016/j.bbali.2019.02.006>
15. Vamecq J, Andreoletti P, El Kebbij R, et al. (2018) Peroxisomal acyl-coA oxidase type 1: anti-inflammatory and anti-aging properties with a special emphasis on studies with LPS and argan oil as a model transposable to aging. *Oxid Med Cell Longev* 2018. <https://doi.org/10.1155/2018/6986984>
16. Ginovyan M, Andreoletti P, Cherkaoui-Malki M, et al. (2022) *Hypericum alpestre* extract affects the activity of the key antioxidant enzymes in microglial BV-2 cellular models. *AIMS Biophys* 9: 161–171. <https://doi.org/10.3934/biophys.2022014>
17. Sahakyan N, Andreoletti P, Cherkaoui-Malki M, et al. (2021) *Artemisia dracunculus* L. essential oil phytochemical components trigger the activity of cellular antioxidant enzymes. *J Food Biochem* 45: e13691. <https://doi.org/10.1111/jfbc.13691>
18. Kurutas EB (2015) The importance of antioxidants which play the role in cellular response against oxidative/nitrosative stress: current state. *Nutr J* 15: 71. <https://doi.org/10.1186/s12937-016-0186-5>

19. Jorgenson TC, Zhong W, Oberley TD (2013) Redox imbalance and biochemical changes in cancer. *Cancer Res* 73: 6118–6123. <https://doi.org/10.1158/0008-5472.CAN-13-1117>
20. Lash LH, Putt DA, Jankovich AD (2015) Glutathione levels and susceptibility to chemically induced injury in two human prostate cancer cell lines. *Molecules* 20: 10399–10414. <https://doi.org/10.3390/molecules200610399>
21. Koss-Mikołajczyk I, Kusznierevicz B, Namieśnik J, et al. (2015) Juices from non-typical edible fruits as health-promoting acidity regulators for food industry. *LWT-Food Sci Technol* 64: 845–852. <https://doi.org/10.1016/j.lwt.2015.06.072>
22. Karuna DS, Dey P, Das S, et al. (2018) *In vitro* antioxidant activities of root extract of *Asparagus racemosus* Linn. *J Tradit Complement Med* 8: 60–65. <https://doi.org/10.1016/j.jtcme.2017.02.004>
23. Baranowska M, Suliborska K, Chrzanowski W, et al. (2018) The relationship between standard reduction potentials of catechins and biological activities involved in redox control. *Redox Biol* 17: 355–366. <https://doi.org/10.1016/j.redox.2018.05.005>
24. Nautiyal J, Banerjee S, Kanwar SS, et al. (2011) Curcumin enhances dasatinib-induced inhibition of growth and transformation of colon cancer cells. *Int J Cancer* 128: 951–961. <https://doi.org/10.1002/ijc.25410>
25. Ferlay J, Colombet M, Soerjomataram I, et al. (2021) Cancer statistics for the year 2020: an overview. *Int J Cancer* 149: 778–789. <https://doi.org/10.1002/ijc.33588>
26. Bedirian K, Aghabekyan T, Mesrobian A, et al. (2022) Overview of cancer control in Armenia and policy implications. *Front Oncol* 11: 782581. <https://doi.org/10.3389/fonc.2021.782581>
27. Council of Europe European (2005) *Pharmacopoeia*, 5 Eds., France.
28. Koss-Mikołajczyk I, Kusznierevicz B, Bartoszek A (2019) The relationship between phytochemical composition and biological activities of differently pigmented varieties of berry fruits; comparison between embedded in food matrix and isolated anthocyanins. *Foods* 8: 646. <https://doi.org/10.3390/foods8120646>
29. Koss-Mikołajczyk I, Kusznierevicz B, Wiczowski W, et al. (2019) The comparison of betalain composition and chosen biological activities for differently pigmented prickly pear (*Opuntia ficus-indica*) and beetroot (*Beta vulgaris*) varieties. *Int J Food Sci Nutr* 70: 442–452. <https://doi.org/10.1080/09637486.2018.1529148>
30. Sassi A, Bouhleb I, Mustapha N, et al. (2016) Assessment *in vitro* of the genotoxicity, antigenotoxicity and antioxidant of *Ceratonia siliqua* L. extracts in murine leukaemia cells L1210 by comet assay. *Regul Toxicol Pharm* 77: 117–124. <https://doi.org/10.1016/j.yrtph.2016.02.009>
31. Sahakyan N, Andreoletti P, Petrosyan M, et al. (2022) Essential oils of basil cultivars selectively affect the activity of antioxidant enzymes in murine glial cells. *Curr Nutraceut* 3. <https://doi.org/10.2174/2665978602666211217143112>
32. Hambarzumyan S, Sahakyan N, Petrosyan M, et al. (2020) *Origanum vulgare* L. extract-mediated synthesis of silver nanoparticles, their characterization and antibacterial activities. *AMB Express* 10: 162. <https://doi.org/10.1186/s13568-020-01100-9>
33. Moghrovyan A, Sahakyan N, Babayan A, et al. (2019) Essential oil and ethanol extract of oregano (*Origanum vulgare* L.) from Armenian flora as a natural source of terpenes, flavonoids and other phytochemicals with antiradical, antioxidant, metal chelating, tyrosinase inhibitory and antibacterial activity. *Curr Pharm Design* 25: 1809–1816. <https://doi.org/10.2174/1381612825666190702095612>

34. Ginovyan MM, Sahakyan NZ, Petrosyan MT, et al. (2021) Antioxidant potential of some herbs represented in Armenian flora and characterization of phytochemicals. *Proc YSU B Chem Biol Sci* 55: 25–38. <https://doi.org/10.46991/PYSU:B/2021.55.1.025>
35. Avetisyan A, Markosian A, Petrosyan M, et al. (2017) Chemical composition and some biological activities of the essential oils from basil *Ocimum* different cultivars. *BMC Complem Altern M* 17: 60. <https://doi.org/10.1186/s12906-017-1587-5>
36. Ziemlewska A, Zagórska-Dziok M, Nizioł-Łukaszewska Z (2021) Assessment of cytotoxicity and antioxidant properties of berry leaves as by-products with potential application in cosmetic and pharmaceutical products. *Sci Rep-UK* 11: 3240. <https://doi.org/10.1038/s41598-021-82207-2>
37. Liu B, Li Z (2016) Black currant (*Ribes nigrum* L.) extract induces apoptosis of MKN-45 and TE-1 cells through MAPK-and PI3K/Akt-mediated mitochondrial pathways. *J Med Food* 19: 365–373. <https://doi.org/10.1089/jmf.2015.3521>
38. Wagh NS, Pai SR, Sonkamble VV (2020) Phytochemicals in the prevention and cure of cancers, In: Swamy, M.K., *Plant-derived Bioactives*, Singapore: Springer Singapore, 351–373. https://doi.org/10.1007/978-981-15-2361-8_16
39. Swamy MK (2020) *Plant-derived Bioactives*, Singapore: Springer Singapore. <https://doi.org/10.1007/978-981-15-1761-7>
40. Dong X, Zou B, Zhang Y, et al. (2013) Preparation of A-type proanthocyanidin dimers from peanut skins and persimmon pulp and comparison of the antioxidant activity of A-type and B-type dimers. *Fitoterapia* 91: 128–139. <https://doi.org/10.1016/j.fitote.2013.08.019>
41. Silva BA, Ferreres F, Malva JO, et al. (2005) Phytochemical and antioxidant characterization of *Hypericum perforatum* alcoholic extracts. *Food Chem* 90: 157–167. <https://doi.org/10.1016/j.foodchem.2004.03.049>
42. Liu W, Li J, Zhang X, et al. (2020) Current advances in naturally occurring caffeoylquinic acids: structure, bioactivity, and synthesis. *J Agr Food Chem* 68: 10489–10516. <https://doi.org/10.1021/acs.jafc.0c03804>
43. De P, Baltas M, Bedos-Belval F (2011) Cinnamic acid derivatives as anticancer agents-a review. *Curr Med Chem* 18: 1672–1703. <https://doi.org/10.2174/092986711795471347>
44. Azeem M, Hanif M, Mahmood K, et al. (2022) An insight into anticancer, antioxidant, antimicrobial, antidiabetic and anti-inflammatory effects of quercetin: a review. *Polym Bull.* <https://doi.org/10.1007/s00289-022-04091-8>
45. Cincin ZB, Unlu M, Kiran B, et al. (2015) Apoptotic effects of quercitrin on DLD-1 colon cancer cell line. *Pathol Oncol Res* 21: 333–338. <https://doi.org/10.1007/s12253-014-9825-3>
46. Massi A, Bortolini O, Ragno D, et al. (2017) Research progress in the modification of quercetin leading to anticancer agents. *Molecules* 22: 1270. <https://doi.org/10.3390/molecules22081270>
47. Moody R, Wilson K, Jaworowski A, et al. (2020) Natural compounds with potential to modulate cancer therapies and self-reactive immune cells. *Cancers* 12: 673. <https://doi.org/10.3390/cancers12030673>
48. Sun Y, Lenon GB, Yang AWH (2020) *Rumex japonicus* Houtt.: a phytochemical, pharmacological, and pharmacokinetic review. *Phyther Res* 34: 1198–1215. <https://doi.org/10.1002/ptr.6601>

49. Wang Q, Zhong S, Wu H, et al. (2022) *In vitro* anti-cancer effect of marmesin by suppression of PI3K/Akt pathway in esophagus cancer cells. *Esophagus-Tokyo* 19: 163–174. <https://doi.org/10.1007/s10388-021-00872-8>
50. Ivanov AA, Ukladov EA, Kremis SA, et al. (2022) Investigation of cytotoxic and antioxidative activity of 1,2,3-triazolyl-modified furocoumarins and 2,3-dihydrofurocoumarins. *Protoplasma* 259: 1321–1330. <https://doi.org/10.1007/s00709-022-01739-0>



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